# Prevalence of different variants of *Klebsiella pneumoniae* in urine samples: A Review

By-Mahi Tasnim Hossain 19136011

A thesis submitted to the Department of Mathematics and Natural Sciences in partial fulfillment of the requirements for the degree of Bachelor of Science in Biotechnology

Bachelor of Science in Biotechnology Department of Mathematics and Natural Science

> BRAC UNIVERSITY September 2023

© 2023. BRAC University All rights reserved.

## Declaration

It is hereby declared that,

1. The thesis submitted is my original work while completing the degree at BRAC University.

2. The thesis does not contain material previously published or written by a third party, except where this is appropriately cited through complete and accurate referencing.

3. The thesis does not contain material that has been accepted or submitted, for any other degree or diploma at a university or other institution.

4. I have acknowledged all the primary sources of help.

Student's Full name and signature:

Mahi Tasnim Hossain 19136011

# Approval

The thesis/project titled "Prevalence of different variants of *K.pneumoniae* in urine samples: A Review."

Submitted by

1. Mahi Tasnim Hossain (19136011)

of Spring, 2019 has been accepted as satisfactory in partial fulfillment of the requirement for the degree of Bachelor of Science in Biotechnology on the "date to be given" of August 2023.

### **Examining Committee:**

Supervisor: (Member)

> Fahim Kabir Monjurul Haque, PhD Associate Professor, Department of Mathematics and Natural Sciences Brac University

Program Director: (Member)

Dr Munima Haque, PhD Associate Professor, Department of Mathematics and Natural Sciences Brac University

Departmental Head: (Chairman)

A F M Yusuf Haider Professor and Chairperson,

Department of Mathematics and Natural Sciences

**BRAC** University

### Acknowledgment

First and foremost, I express my gratitude and appreciation to the divine being for the ongoing benefits that have enabled me to complete my thesis successfully. I would like to express my gratitude to my family for their unwavering support and encouragement since their contribution played an indispensable role in the successful completion of my thesis.

I express my utmost appreciation to Dr. Fahim Kabir Monjurul Haque, an Assistant Professor in the Department of Mathematics and Natural Sciences at BRAC University, for his invaluable contributions and mentorship that served as a source of inspiration during the course of my thesis. Working under his supervision presented a valuable opportunity that is anticipated to yield future benefits. Finally, I would like to express my gratitude to my acquaintances and supporters, namely Tahani Tabassum, Arko Roy, Nafisa Ahmed, Fariya Anny, Durdana Hossain, Nairita Ahsan Faruqi, Sadrina Afrin Mowna for their encouraging remarks that significantly contributed to enhancing my focus. I would also like to express my sincere appreciation to Ebtesham Shahalam, whose unwavering support has been invaluable to me.

I express profound gratitude towards the individuals indicated above for their contributions, as their presence has been crucial in facilitating my academic and psychological well-being, enabling the effective attainment of my initial objectives. This experience presented an opportunity for me to acquire knowledge within the scope of my personal interests. Therefore, I am confident that the knowledge I have acquired through my thesis will prove beneficial in the foreseeable future.

Maticamen

Mahi Tasnim Hossain.

19136011.

### Abstract

Klebsiella pneumoniae is a causative agent which is responsible for multiple infections along with pneumonia. Along with pneumonia, Urine infections are another symptom that is caused by to invasion of *Klebsiella*. It also causes blood steam infection (BSI), surgical site infection and pneumonia. *Klebsiella* was responsible for 7.4% of the overall prevalence of bacterial infections. In this study, we have reevaluated, summarized, and analyzed the prevalence of Klebsiella pneumoniae among pregnant, newborn, and different age groups all together as well as the extent of *Klebsiella pneumoniae* from urine samples worldwide. For example, Carbapenem-resistant bacteria According to a recent study, *Klebsiella pneumoniae* is the leading cause of human mortality owing to antibiotic resistance in Europe. Prevalence of *Klebsiella pneumoniae* is higher in Western, European, and African regions (100%, 98.70% and 91.67% in Ohio, Lisbon, and Algeria respectively). 3.63 % is the lowest percentage of *Klebsiella pneumoniae* found in urine samples in Taiwan. On the other hand, Enoxacin does not exhibit any antibiotic resistance in any of the samples that were obtained from the majority of nations, with the exception of Peshawar, Pakistan, where it exhibits 43.48% resistance. Conversely, urine samples obtained from patients in Saudi Arabia shown no resistance to Ampicillin and lacked multidrug resistance. With the exception of Malabo, patient urine samples from Ethiopia, Kenya, and Nigeria do not reveal multidrug resistance to Klebsiella pneumoniae. However, further research from Middle Eastern regions and third-world countries interprets the dangers of pneumonia as the main cause of urinary tract infection (UTI). Other causes of this symptom are inflammation, trouble in urination, Diabetes, hypertension, and many more hazards and consequences. *Klebsiella pneumoniae* causes trouble in breathing, tremor, blood infection, and bloody diarrhea. Different countries show the prevalence of *Klebsiella pneumoniae* and its antibiotic resistance along with risks.

# Keywords

Prevalence, Antibiotic susceptibility, Urinary tract infections, microtubules, microfilaments, Hospitalacquired infection, Community-acquired pneumonia, aspiration pneumonia, and blood infection.

# **Dedication (Optional)**

I intend to deliver this letter to my family; expressing gratitude for the motivation and support they have provided me.

# Table of contents

Declaration2
Approval
Acknowledgement4
Abstract5
Keywords6
Dedication7
Table of Contents
List of Tables10
List of Acronyms11
Chapter 1
Introduction12
1.1 Epidemiology
Chapter 214
Research Methodology14
2.1 Search strategy14
2.2 inclusion criteria14
2.3 Exclusion criteria14
Chapter 3
Klebsiella pneumoniae15-16
3.1 Morphology15
3.2 Resistant strains15-16

3.3 Signs and symptoms	16
Chapter 4	17-19
Prevalence of <i>K.pneumoniae</i> in urine samples	17-19
4.1 Highest prevalence of <i>K.pneumoniae</i> in urine samples	17-18
4.2 Prevalence of <i>K.pneumoniae</i> in Asian region	9
4.3 Prevalence of K.pneumoniae in Western and European Region	19
Chapter 52	20-23
5.1 Highest antibiotic resistance of K.pneumoniae in urine samples20	
5.2 Antibiotics resistant to urine samples in Asian region	20
5.3 Antibiotic resistance of K.pneumoniae of urine samples in African region	22-23
Chapter 6	24
Conclusion	24
Chapter 7	25-32
References	25-32

# List of Tables

Serial Number	Title	Page number
1	The prevalence of <i>K.pneumoniae</i> in urine samples	17-18
2	Antibiotic resistance of K.pneumoniae in urine samples	21-22

# List of acronyms

Acronyms	Explanation
BSI	Bloodstream Infection
САР	Community Associated Pneumonia
НАР	Hospital Associated Pneumonia
MF	Microfilaments
МТ	Microtubules
UTI	Urinary Tract Infection
VAP	Ventilator Associated Pneumonia

# Chapter-1 Introduction

#### **1.1 Epidemiology**

*Klebsiella pneumoniae*, a well-known opportunistic pathogen that causes nosocomial pneumonia, bacteremia, and septicemia, also causes urinary tract infections (UTI) (Oelschlaeger & Tall,1997). *K.pneumoniae* biotypes are often acknowledged as opportunistic pathogens that function as causative agents of bacteremia's and respiratory and genitourinary infections. These infections are more prevalent in individuals experiencing physiological or psychological stress (Merino et al.,1992).

### 1.2. Hazards

Over the course of the past several decades, there has been an increase in the level of awareness regarding the role of *K.pneumoniae* as a significant opportunistic pathogen of the urinary tract in compromised individuals and hospitalized patients (Clegg & Murphy,2017). This bacterium is commonly found in the intestinal tract of humans, and rates of fecal carriage can increase threefold during hospitalization. Multiple genetic resistance mechanisms evolve due to the selective use of antibiotics in a hospital setting (Matovina et al.,2021).

### **1.3.** Pathophysiology

*Klebsiella* invasion in vivo hasn't been looked into yet, so it's not clear if it's another thing that makes *K.pneumoniae* dangerous (Merino et al.,1992). The advent of these bacteria, which display different antibiotic resistance phenotypes, has made treatment and management of *K.pneumoniae* urinary tract infections (UTIs) challenging (Clegg & Murphy,2017). But because more and more papers are talking

about how many bacterial pathogens can get into eukaryotic cells, we looked into *K.pneumoniae's* possible ability to do this (Merino et al.,1992). This is because UTIs are caused by *K.pneumoniae* (Clegg & Murphy,2017). In a gentamicin kill-invasion assay, this study shows how a *K.pneumoniae* isolate from a person with a UTI was able to invade the human bladder and ileocecal epithelial cells in a lab setting (Merino et al.,1992). Isolates of *K.pneumoniae* that did not show susceptibility to one or more antibiotics in three or more different antimicrobial groups were categorized as multidrug-resistant *K.pneumoniae* (Filgona et al,2015). We looked at the role of receptor-mediated endocytosis, endosome acidification, and the influence of microtubules (MT) and microfilaments (MF) in the internalization of *K.pneumoniae* using a number of inhibitors that work on eukaryotic cell structures or processes (Merino et al.,1992).

### **Chapter-2**

### **Research Methodology**

### 2.1 Search strategy

Using databases such as ScienceDirect, Google Scholar, PubMed, and Scopus, pertinent scientific literature was located. *K.pneumoniae*, pneumonia, prevalence, urine sample, urinary tract infection and UTI were used as search terms. Using the Boolean operators "AND", "OR", and "NOT", the search result was kept specific. Authentic research and review articles with a high number of citations were chosen to retrieve subject-related information.

#### 2.2 Inclusion criteria

The literature includes a description of the incidence of *K.pneumoniae* in urine samples from patients, an examination of the risk factors associated with urinary tract infections (UTIs) during pneumonia, and an analysis of the distribution of *Klebsiella* species among individuals of various age groups. Additionally, the literature briefly contained information regarding the symptoms and dangers associated with *K.pneumoniae* during urinary tract infections (UTIs).

### 2.3. Exclusion criteria

Literature that only stated the prevalence of bacterial nosocomial infection, the antibiotic resistance of *Klebsiella* spp., the diagnosis and treatment of *K.pneumoniae* were excluded. In addition, articles that merely reported the prevalence of *K.pneumoniae* among non-pregnant or men and women of reproductive age were excluded.

### **Chapter-3**

### Klebsiella pneumoniae

#### 3.1. Morphology

*Klebsiella* accounted for 7.4% of the total incidence of bacterial illnesses (Jay, 1983). It ranked as the third most prevalent etiology for lower respiratory tract infections and primary bacteremia across all categories (Jay,1983). *K.pneumoniae*, a member of the order Enterobacterales and widely known as a nosocomial pathogen that is capable of causing a variety of infectious syndromes, is one of the bacteria in this order (Matovina et al.,2021). According to the World Health Organization, new antibiotics are desperately needed to combat *K.pneumoniae*, a bacterium that causes serious infections in both hospitals and the population (David et al., 2019).

### 3.2. Resistant Strains

In previous extensive surveillance studies, *Klebsiella* has been identified as the predominant etiological agent responsible for nosocomial pneumonia and bacteremia resulting from pneumonia (Jay,1983). Carbapenem-resistant *K.pneumoniae* is the leading cause of human mortality due to antibiotic resistance in Europe, according to a recent study (David et al.,2019). *K.pneumoniae* causes suppurative lesions, bacteremia, and septicemia, which are responsible for a large number of nosocomial infections in newborns, people getting respiratory treatment, and people in the urology and burn wards of hospitals (Nassif & Sansonetti, 1986). The current study found that patients infected with KPC-producing *K.pneumoniae* had a mortality rate significantly higher than the pooled overall mortality rate (47.66 vs. 42.14%). This result may contribute to KPC-producing *K.pneumoniae* having higher invasiveness, and the KPC-encoding black always carries other drug-resistant genes, resulting in a

prominent drug-resistant phenotype in the bacteria (Xu et al.,2017). In previous extensive surveillance studies, *Klebsiella* has been identified as the predominant etiological agent responsible for nosocomial pneumonia and bacteremia resulting from pneumonia (Jay,1983). *K.pneumoniae*, which is one of the most prevalent *Klebsiella* species, is the most prevalent cause of hospital-acquired infections, meningitis, pneumonia, bacteremia, wound infections, and urinary tract infections (Pourbaghi et al., 2021).

#### 3.3. Signs and symptoms

According to a survey, *Klebsiella* was identified as the primary etiological agent of pneumonia among those residing in nursing homes, accounting for a substantial 40% of the total reported cases (Jay,1983). It is common knowledge that *K.pneumoniae* is found everywhere in nature. It is considered to be one of the most significant opportunistic pathogens, and it is responsible for a wide variety of human illnesses, including bloodstream infection (BSI), urinary tract infection (UTI), surgical site infection, and pneumonia (Xu et al., 2017). *Klebsiella* species are highly prevalent Gram-negative bacilli that can be detected in various areas of the body, as well as in the environment and normal flora (Pourbaghi et al., 2021). *Klebsiella* is a prominent etiological agent responsible for pneumonia in elderly populations residing in nursing homes, chronic care facilities, and community settings (Jay, 1983).

## **Chapter-4**

### Prevalence of K.pneumoniae in urine samples

### 4.1. High prevalence of *K.pneumoniae* in urine samples

Numerous research has been conducted across diverse age groups and geographical regions worldwide, revealing that Ohio, located inside the United States, exhibits the highest prevalence of *K.pneumoniae* compared to other global regions. A study conducted by Brizendine et al. (2015) revealed a notable increase in the prevalence of positive microscopic results in Connecticut (4.89%) (So et al.,2015) and New Jersey (18.90%) (Popejoy et al.,2016) compared to other regions of the United States. Taiwan exhibited the most minimal prevalence rate among all nations, amounting to 3.63% (Lin et al.,2014)

				Sample numbe		ction thod	Percentage of	
Seria				r of			K.pneumonia	
1	Country	Area	Year	patient	PCR	Culture	e	Reference
	Banglades							(Chakraborty et
1	h	Sylhet	2016	108	Х	Yes	90%	al.,2016)
								(Ikeda et
2	Japan	Tokyo	2022	62	Yes	Х	45.16%	al.,2019)
3	Korea	Seoul	2016	81	Х	Yes	12.30%	(Kim et al.,2017)
4	Korea	Seoul	2010	185	Yes	Х	45.56%	(Ko et al.,2010)
								(Chander &
5	Nepal	Kathmandu	2013	145	Х	Yes	2.29%	shreshtha, 2013)
6	Nepal	Kathmandu	2017	39	Х	Yes	14.60%	(Nepal et al.,2017)
								(Fatima et
7	Pakistan	Baluchistan	2021	107	Х	Yes	20.80%	al.,2021)
8	Pakistan	Lahore	2021	100	Х	Yes	37%	(Iqbal et al.,2021)
								(Naeem et
9	Pakistan	Peshawar	2020	350	Yes	Yes	64.80%	al.,2021)
								(Nasomsong et
10	Thailand	Bangkok	2022	49	Х	Yes	90.50%	al.,2021)
								(Vachvanichsanon
11	Thailand	Songkhla	2020	324	Х	Yes	75.50%	g et al., 2021)
12	Taiwan	Tainan	2014	468	Yes	Х	3.63%	(Lin et al.,2014)
		Ilam,					39.4%,	(Ghafourian et
13	Iran	Tabriz,	2011	288	Yes	Yes	50.7%, 45.8%	al.,2012)

		Tehran						
14	Iran	Kashan	2015	250	Х	Yes	52.60%	(Moini et al.,2015)
15	Iran	Semnan	2018	173	X	Yes	32.94%	(Moghadas et al.,2018)
16	Iraq	Kufa	2016	464	Yes	Yes	12.26%	(Ahmed & Alaa.,2016)
17	Iraq	Kufa	2017	285	Yes	Yes	11.26%	(Ahmed & Haneen.,2017)
18	Saudia Arabia	Riyadh	2016	160	X	Yes	23.50%	(Al Yousef et al., 2016)
19	USA	Connecticu t	2014	143	X	Yes	4.89%	(So et al.,2015)
20	USA	New Jersey	2016	159	X	Yes	18.90%	(Popejoy et al.,2016)
21	USA	Ohio	2014	108	X	Yes	100%	(Brizendine et al.,2015)
22	Colombia	Valledupar	2012	123	Yes	X	12.70%	(Martinez et al.,2012)
23	Norway	Bærum	2012	101	Yes	Yes	44%	(Jørgensen et al.,2017)
24	Portugal	Braga, Vila Real	2022	102	Yes	Yes	39.20%	(Oliveira et al.,2022)
25	Portugal	Lisbon	2018	76	Yes	X	98.70%	(Marques et al.,2019)
26	Russia	Kazan	2018	10	Yes	Yes	100%	(Khaertynov et al.,2018)
27	Algeria	Tlemcen	2013	24	Yes	X	91.67%	(Yandai et al.,2019)
28	Ethiopia	Addis Ababa	2018	342	X	Yes	14.90%	(Abayneh et al.,2018)
29	Sudan	Khartoum	2020	250	Yes	Yes	55.60%	(Osman et al.,2020)
30	TChad	N'Djamena	2019	503	X	Yes	25%	(Bellifa et al.,2013)

Table 1: Prevalence of *K.pneumoniae* in urine samples. Data has been organized geographically.

### 4.2. Prevalence of K.pneumoniae in Asian region

Sylhet shows a rise of *K.pneumoniae* infection among individuals by 90% (Chakraborty et al.,2016). Moreover, moderate invasion of *Klebsiella* infection has been observed Korea by 45.56% (Ko et al.,2010) and 12.30% (Kim et al.,2017). Among the middle eastern regions, Nepal shows the lowest rate of prevalence, 2.29% in urine sample (Chander & shreshtha, 2013). In Iran, different provinces show moderate percentage of *K.pneumoniae* (Ghafourian et al.,2012). In Pakistan, estimation is still moderate in Baluchistan (Fatima et al.,2021) and Lahore (Iqbal et al.,2021). In 2020, percentage began to rise by 64.80% from patients' urine sample (Naeem et al.,2021). In Taiwan, urine samples show lowest percentage of *K.pneumoniae* percentage, 3.63% (Lin et al.,2014). Also, Bangkok shows dramatic rise of *K.pneumoniae* in urine samples which is 90.50% (Nasomsong et al.,2021).

#### **4.3.** Prevalence of K.pneumoniae in Western and European Region

However, the highest prevalence is noticed 100% according to Table-1 by (Brizendine et al.,2015) and (Khaertynov et al.,2018). To elaborate, the USA shows 100% prevalence in Ohio (Brizendine et al.,2015) but lowest percentage in Connecticut, 4.89% (So et al.,2015) and New Jersey,18.90% (Popejoy et al.,2016). Similarly, Colombia shows a decrease in percentage compared to other western regions (Martinez et al.,2012). In Norway, percentage of *K.pneumoniae* is moderately lower but observed both in PCR and culture samples (Jørgensen et al.,2017). In Portugal, the percentage of *K.pneumoniae* is 98.70% in Lisbon, PCR detection confirmed by (Marques et al.,2019). Compared to other European regions, Braga and Vila Real samples confirmed *K.pneumoniae* percentage, 39.20% both in PCR and culture method (Oliveira et al.,2022). In Kazan, patients' urine samples confirmed 100% prevalence of *K.pneumoniae* both in PCR and culture detection test (Khaertynov et al.,2018).

In Algeria, 91.67% prevalence is observed in a study conducted by (Yandai et al.,2019) from the PCR samples of patients' urine samples. Among Ethiopia, Sudan and TChad, prevalence is comparatively low in Addis Ababa, 14.90% a study conducted by (Abayneh et al.,2018). Similarly, TChad shows moderately lower percentage of *K.pneumoniae*,25% (Bellifa et al.,2013) than in Sudan, 55.60% (Osman et al.,2020). However, further studies should be administered and carried on for more information as there were very few research done in African regions.

#### Chapter-5

#### Antibiotic resistance of *K.pneumoniae* in urine samples

#### 5.1. Highest antibiotic resistance of *K.pneumoniae* in urine samples

Multiple studies had identified the multidrug resistance and antibiotic resistance caused by *K.pneumoniae* from urine samples in which the prevalence of *K.pneumoniae* was the highest in the respective study. Besides, several antibiotics show different percent estimation in different antibiotics. However, the percent estimation of Ampicillin is 100% all the time in urine samples of almost all countries except in some province of Iran such as Shahrekord (Amraie et al.,2014), Shiraz (Mansury et al.,2016), Sari (Shadkam etal.,2021). On the other hand, samples of urine collected from patients in Saudia Arabia showed no resistance to Ampicillin and no multidrug resistance (Kader & Kumar,2005).

### 5.2. Antibiotics resistant to urine samples in Asian region

Enoxacin shows no antibiotic resistance in any samples collected from maximum countries except in Peshawar, Pakistan, 43.48% (Ullah et al,2009). In Bangladesh, Cefpirome and Cefradine did not show any resistance to *K.pneumoniae*, from samples collected from Dhaka (Aminul et al 2021) and Sylhet (Chakraborty et al 2016). Moreover, 41.30% of antibiotic resistance has been observed from antibiotic Gatifloxacin from urine samples collected from Peshawar, Pakistan (Ullah et al,2009). In Rohtak, a study conducted by (Gupta et al.,2007) shows that *K.pneumoniae* shows 100% multidrug resistance collected from the patient's urine samples. 93.33% antibiotic resistance by Imipenem has been observed in Peshawar, Pakistan (Ullah et al,2009).

				Sa mpl		Name of Antibiotics															
				e																Multi	
				nu			Amox icillin													drug	
				mbe			+ clavul					Co-					Gatif			Resis	_
Serial	Country	Area	ar		Ampi cillin			Ceftaz idime					Ciprofl oxacin					Imipen em	Merop enem	tance	Reference
1	Banglad esh	Dhaka	20 21	150	-	50.30 %			72.80 %	-	-	-	-	-		60.30 %	-	-	44.70 %	82%	(Aminul et al 2021)
2	Banglad esh	Sylhet	20 16	500	100%	-	-	-	50%	-	-	44%	37.50%	-	-	31%	-	-	-	56%	(Chakrabort y et al 2016)
3		Karna taka		417	75.60 %	14.60 %	-	-	-	-	-	-	36.50%	-	-	36.50 %	-	0%	-	90.20 %	(Manjula et al.,2014)
4		Kolen chery	20 16		100%	32.30 %	68%	-	-	_	-	60%	-	-	-	40%	-	14.30%	34.30 %	-	(Varghese et al 2016)

		Rohta	20	575																	(Courts at
5	India	k k	20 07	90	-	68%	-	-	60%	-	-	-	-	-	-	40%	-	-	-		(Gupta et al.,2007)
6	Indonesi a	Klate n	20 19	167	100%		36.69 %	_	_	_	_	_	38.75%	-	_	_	-	_	_		(Nirwati et al.,2019)
		~																			
7	Pakistan		20 09				17.39 %		32.61 %				45.65%			17.39 %	41.3 0%	93.33%			(Ullah et al,2009)
8	Iran	Tabriz	20 20	421	-	19.70 %		18.18 %	41.83 00%	-	-		56.970 %	-		87.80 %	-	6.06%	-	53.30	(Jafari- Sales et al.,2020)
9	Iran	Tehra n	20 20	88	-				45.40 %	-		52.20 %	26%	-		27.20 %	-	11.30%	-	48.80	(Moghada m et al.,2020)
10	Iran	Shahr ekord	20 14	195		21.96 %			41.61 %	-	-	54.33 %	15.60%	-		32.94 %	-	20.80%	-		(Amraie et al.,2014)
11	Iran	Shiraz	20 16	144	-	15.90 %	1	43.70 %	-	-	-	43.70 %	-	-	-	19.40 %	-	15.90%	11.80 %		( <u>Mansury</u> et al.,2016)

12	Iran	Sari	20 21	100	-	10%	-		43.30 %	-	-	-	65.60%	-	-	-	-	35.80%	41.70 %	74.30 %	( <mark>Shadkam</mark> etal.,2021)
13	Iran	Kasha n		250	96%	48.10 %	46.30 %	50%	52%	-	-	-	46%	-	-	59.3%	-	0%		46.60 %	(Moini et al.,2015) [50]
14		Alkho bar. Dam mam	20 05	245 5	-	38%	-	-	-	-	-	-	43%	-	-	40%	-	7%	7%	-	(Kader & Kumar, 2005)
15	Portugal	Lisbo n	20 18		96.50 %	0%	-	8.80%	-	-	-	-	14%	-	-	14%	-	0%	0%	30%	(Marques et al.,2019)
16	Equatori al Guinea	Malab		785	100%		93.30 %		81.30 %	-	-	-	62.50%	85.80 %		86.20 %	-	3.30%		91.70 %	(Shatalov, 2015)
17	Ethiopia	Jimm a		228	100%	-	-	-	25%	-	-	-	0%	-	-	25%	-	-	-	-	(Beyene & Tsegaye, 2011)
18	Kenya	Nairo bi		336	100%	-		90.60 %	-	_	-	-	83%	_		63.60 %	-	0.60%	0.60%	_	(Maina et al 2013)
19	Nigeria	Anvig ba		200	-	-	83.30 %	41.70 %	-	-	-	-	75%	-	-	66.70 %	-	100%	-	-	(Mofolorun sho et al 2021)
20		Kamp ala, Kayu nga, Mpigi	20 16	144 8	100%	-	-	-	3%	-	-	-	11%	-	-	11%	-	-	-	60%	( <u>Najjuka</u> et al 2016)

# **Table 2: Antibiotic resistance of** *K.pneumoniae* **in urine samples.** Data has been arranged geographically.

### 5.3 Antibiotic resistance of K.pneumoniae of urine samples in African region.

From urine samples collected from Anyigba, Nigeria, the samples show 100% antibiotic resistance to Imipenem but zero signs of multidrug resistance (Mofolorunsho et al 2021). 85.80% antibiotic resistance by Doxycycline has been observed in Equatorial Guinea, Malabo (Shatalov,2015). A study conducted by (Kader&Kumar,2005) illustrates no multidrug resistance in Saudia Arabia. Except, Malabo (Shatalov,2015), African regions such as Ethiopia (Beyene&Tsegaye,2011), Kenya (Maina et

al2013) and Nigeria (Mofolorunsho et al 2021) shows no multidrug resistance to *K.pneumoniae* in urine samples collected from patients.

### **Chapter-6**

### **Conclusion:**

To recapitulate, K.pneumoniae is causing several infections around the globe including Hospital Acquired Pneumonia (HAP), Ventilator Associated Pneumonia (VAP), Community Associated Pneumonia (CAP), and Aspiration Pneumonia. Among the countries, Iran, Iraq, and some countries in Africa showed invasion of Pneumonia more compared to other Western countries. However, the countries are more associated with antibiotic resistance compared to Asian regions as well. K.pneumoniae is more invasive and toxic having multiple variants of its own. Hypervirulent K.pneumoniae are more susceptible in different healthcare sectors along with the expansion of Carbapenem-Resistant K.pneumoniae. Moreover, K.pneumoniae are tough to recognize and diagnosis is quite challenging due to its multiple variants and mutative modification in genetic material. Therefore, I think extensive field research needs to be administered more in the public health care sector and more research needs to be done in life science facilities. More life science sectors need to work on their samples and identify the dangers and treatment of disease caused by K.pneumoniae. In a nutshell, the prevalence and antibiotic susceptibility of K.pneumoniae need to be studied in in broader sense and the dangers of its multiple variants need to be figured out more in general especially in thirdworld countries to prevent the risk factors and other health hazards.

# Chapter-7 Reference

1. Ikeda, M., Mizoguchi, M., Oshida, Y., Tatsuno, K., Saito, R., Okazaki, M., ... & Moriya, K. (2018). Clinical and microbiological characteristics and occurrence of K.pneumoniae infection in Japan. *International Journal of General Medicine*, 293-299. http://dx.doi.org/10.2147/IJG

2. Kim, Y. J., Kim, S. I., Kim, Y. R., Wie, S. H., Lee, H. K., Kim, S. Y., & Park, Y. J. (2017). Virulence factors and clinical patterns of hypermucoviscous K.pneumoniae isolated from urine. Infectious diseases, 49(3), 178-184. https://doi.org/10.1080/23744235.2016.1244611

3. Chakraborty, S., Mohsina, K., Sarker, P. K., Alam, M. Z., Karim, M. I. A., & Sayem, S. A. (2016). Prevalence, antibiotic susceptibility profiles and ESBL production in K.pneumoniae and Klebsiella oxytoca among hospitalized patients. *Periodicum biologorum*, *118*(1). https://doi.org/10.18054/pb.v118i1.3160

4. Nasomsong, W., Nulsopapon, P., Changpradub, D., Pongchaidecha, M., Pungcharoenkijkul, S., Juntanawiwat, P., ... & Santimaleeworagun, W. (2021). The potential use of ceftazidime-avibactam against Carbapenem resistant K.pneumoniae clinical isolates harboring different carbapenemase types in a Thai University Hospital. *Drug Design, Development and Therapy*, 3095-3104. https://doi.org/10.2147/DD

5. Ghafourian, S., Sekawi, Z., Neela, V., Khosravi, A., Rahbar, M., & Sadeghifard, N. (2012). Incidence of extended-spectrum beta-lactamase-producing K.pneumoniae in patients with urinary tract infection. *Sao Paulo Medical Journal*, *130*, 37-43. https://doi.org/10.1590/S1516-31802012000100007

6. Al Yousef, S. A., Younis, S., Farrag, E., Moussa, H. S., Bayoumi, F. S., & Ali, A. M. (2016).
Clinical and laboratory profile of urinary tract infections associated with extended spectrum β-lactamase producing Escherichia coli and K.pneumoniae. *Annals of Clinical & Laboratory Science*, 46(4), 393-400. <u>https://doi.org/10.15537/smj.2018.12.23273</u>

7. Ko, K. S., Lee, J. Y., Baek, J. Y., Suh, J. Y., Lee, M. Y., Choi, J. Y., ... & Song, J. H. (2010). Predominance of an ST11 extended-spectrum β-lactamase-producing K.pneumoniae clone causing bacteraemia and urinary tract infections in Korea. *Journal of Medical Microbiology*, *59*(7), 822-828. https://doi.org/10.1099/jmm.0.018119-0

8. Marques, C., Menezes, J., Belas, A., Aboim, C., Cavaco-Silva, P., Trigueiro, G., ... & Pomba, C. (2019). K.pneumoniae causing urinary tract infections in companion animals and humans: population structure, antimicrobial resistance and virulence genes. *Journal of Antimicrobial Chemotherapy*, 74(3), 594-602. <u>https://doi.org/10.1093/jac/dky499</u>

9. Naeem, S., Bilal, H., Muhammad, H., Khan, M. A., Hameed, F., Bahadur, S., & Rehman, T. U. (2021). Detection of blaNDM-1 gene in ESBL producing Escherichia coli and K.pneumoniae isolated from urine samples. *The Journal of Infection in Developing Countries*, *15*(04), 516-522. https://doi.org/10.3855/jidc.12850

10. Moghadas, A. J., Kalantari, F., Sarfi, M., Shahhoseini, S., & Mirkalantari, S. (2018). Evaluation of virulence factors and antibiotic resistance patterns in clinical urine isolates of K.pneumoniae in Semnan, Iran. *Jundishapur Journal of Microbiology*, *11*(7).

15. Al Yousef, S. A., Younis, S., Farrag, E., Moussa, H. S., Bayoumi, F. S., & Ali, A. M. (2016). Clinical and laboratory profile of urinary tract infections associated with extended spectrum  $\beta$ -lactamase producing Escherichia coli and K.pneumoniae. *Annals of Clinical & Laboratory Science*, 46(4), 393-400. <u>https://doi.org/10.5812/jjm.63637</u>

11. Iqbal, Z., Mumtaz, M. Z., & Malik, A. (2021). Extensive drug-resistance in strains of Escherichia coli and K.pneumoniae isolated from paediatric urinary tract infections. *Journal of Taibah University Medical Sciences*, *16*(4), 565-574.

https://doi.org/10.1016/j.jtumed.2021.03.004

12. Chander, A., & Shrestha, C. D. (2013). Prevalence of extended spectrum beta lactamase producing Escherichia coli and K.pneumoniae urinary isolates in a tertiary care hospital in Kathmandu, Nepal. *BMC Research notes*, *6*(1), 1-6. DOI:10.1186/1756-0500-6-487

13. Ahmed, A. J. A., & Alaa, H. A. A. (2016). Virulence factors and antibiotic susceptibility patterns of multidrug resistance K.pneumoniae isolated from different clinical infections. *African Journal of Microbiology Research*, *10*(22), 829-843. DOI: 10.5897/AJMR2016.8051

14. Khaertynov, K. S., Anokhin, V. A., Rizvanov, A. A., Davidyuk, Y. N., Semyenova, D. R., Lubin, S. A., & Skvortsova, N. N. (2018). Virulence factors and antibiotic resistance of K.pneumoniae strains isolated from neonates with sepsis. *Frontiers in medicine*, *5*, 225. <u>https://doi.org/10.3389/fmed.2018.00225</u>

15. Popejoy, M. W., Paterson, D. L., Cloutier, D., Huntington, J. A., Miller, B., Bliss, C. A., ... & Kaye, K. S. (2016). Efficacy of ceftolozane/tazobactam against urinary tract and intra-abdominal infections caused by ESBL-producing Escherichia coli and K.pneumoniae: a pooled analysis of Phase 3 clinical trials. *Journal of Antimicrobial Chemotherapy*, 72(1), 268-272. https://doi.org/10.1093/jac/dkw374

16. Jørgensen, S. B., Søraas, A., Sundsfjord, A., Liestøl, K., Leegaard, T. M., & Jenum, P. A. (2017). Fecal carriage of extended spectrum  $\beta$ -lactamase producing Escherichia coli and K.pneumoniae after urinary tract infection–a three year prospective cohort study. *PloS one*, *12*(3), e0173510. <u>https://doi.org/10.1086/519865</u>

17. So, W., Crandon, J. L., & Nicolau, D. P. (2015). Effects of urine matrix and pH on the potency of delafloxacin and ciprofloxacin against urogenic Escherichia coli and K.pneumoniae. *The Journal of Urology*, *194*(2), 563-570.https://doi.org/10.1016/j.juro.2015.01.094

18. Lin, W. H., Kao, C. Y., Yang, D. C., Tseng, C. C., Wu, A. B., Teng, C. H., ... & Wu, J. J. (2014). Clinical and microbiological characteristics of K.pneumoniae from community-acquired recurrent urinary tract infections. *European journal of clinical microbiology & infectious diseases*, *33*, 1533-1539. DOI 10.1007/s10096-014-2100-4

19. Nepal, K., Pant, N. D., Neupane, B., Belbase, A., Baidhya, R., Shrestha, R. K., ... & Jha, B. (2017). Extended-spectrum beta-lactamase and metallo beta-lactamase production among Escherichia coli and K.pneumoniae isolated from different clinical samples in a tertiary care hospital in Kathmandu, Nepal. *Annals of clinical microbiology and antimicrobials*, *16*(1), 1-7. DOI 10.1186/s12941-017-0236-7

20. Oliveira, R., Castro, J., Silva, S., Oliveira, H., Saavedra, M. J., Azevedo, N. F., & Almeida, C. (2022). Exploring the antibiotic resistance profile of clinical K.pneumoniae isolates in Portugal. *Antibiotics*, *11*(11), 1613.<u>https://doi.org/10.3390/antibiotics11111613</u>

21. Vachvanichsanong, P., McNeil, E. B., & Dissaneewate, P. (2021). Extended-spectrum betalactamase Escherichia coli and K.pneumoniae urinary tract infections. *Epidemiology & Infection*, *149*, e12. <u>https://doi.org/10.1017/S0950268820003015</u> 22. Ahmed, J., & Haneen, M. R. J. A. (2017). Phenotypic and molecular characterization of multidrug resistant K.pneumoniae isolated from different clinical sources in Al-Najaf province-Iraq. *Pak. J. Biol. Sci*, *20*(5), 217-232. DOI: 10.3923/pjbs.2017.217.232

23. Brizendine, K. D., Richter, S. S., Cober, E. D., & van Duin, D. (2015). Carbapenem-resistant K.pneumoniae urinary tract infection following solid organ transplantation. *Antimicrobial agents and chemotherapy*, *59*(1), 553-557. <u>https://doi.org/10.1128/aac.04284-14</u>

24. Fatima, S., Liaqat, F., Akbar, A., Sahfee, M., Samad, A., Anwar, M., ... & Khan, A. (2021). Virulent and multidrug-resistant K.pneumoniae from clinical samples in Balochistan. *International Wound Journal*, *18*(4), 510-518. <u>https://doi.org/10.1111/iwj.13550</u>

25. Martinez, P., Garzón, D., & Mattar, S. (2012). CTX-M-producing Escherichia coli and K.pneumoniae isolated from community-acquired urinary tract infections in Valledupar, Colombia. *The Brazilian Journal of Infectious Diseases*, *16*(5), 420-425. https://doi.org/10.1016/j.bjid.2012.05.001

26. Osman, E. A., El-Amin, N., Adrees, E. A., Al-Hassan, L., & Mukhtar, M. (2020). Comparing conventional, biochemical and genotypic methods for accurate identification of K.pneumoniae in Sudan. *Access microbiology*, 2(3). <u>https://doi.org/10.1099%2Facmi.0.000096</u>

27. Bellifa, S., Hassaine, H., Balestrino, D., Charbonnel, N., M'hamedi, I., Terki, I. K., ... & Forestier, C. (2013). Evaluation of biofilm formation of K.pneumoniae isolated from medical devices at the University Hospital of Tlemcen, Algeria. *Afr J Microbiol Res*, 7(49), 5558-64. DOI: 10.5897/AJMR12.2331

28. Yandai, F. H., Ndoutamia, G., Nadlaou, B., & Barro, N. (2019). Prevalence and resistance profile of Escherichia coli and K.pneumoniae isolated from urinary tract infections in N'Djamena, Tchad. *International Journal of Biological and Chemical Sciences*, *13*(4), 2065-2073. https://doi.org/10.4314/ijbcs.v13i4.13\_

29. Moini, A. S., Soltani, B., Ardakani, A. T., Moravveji, A., Erami, M., Rezaei, M. H., & Namazi, M. (2015). Multidrug-resistant Escherichia coli and K.pneumoniae isolated from patients in Kashan, Iran. *Jundishapur journal of microbiology*, 8(10). https://doi.org/10.5812%2Fjjm.27517

30. Abayneh, M., Tesfaw, G., & Abdissa, A. (2018). Isolation of extended-spectrum  $\beta$ -lactamase-(ESBL-) producing Escherichia coli and K.pneumoniae from patients with community-onset urinary

tract infections in Jimma University Specialized Hospital, Southwest Ethiopia. *Canadian Journal of Infectious Diseases and Medical Microbiology*, 2018. <u>https://doi.org/10.1155/2018/4846159</u>

31. Ullah, F., Malik, S. A., & Ahmed, J. (2009). Antimicrobial susceptibility pattern and ESBL prevalence in K.pneumoniae from urinary tract infections in the North-West of Pakistan. *Afr J Microbiol Res*, *3*(11), 676-80.

https://academicjournals.org/journal/AJMR/article-full-text-pdf/18C6B9B14587

32. Jafari-Sales, A., Soleimani, H., & Moradi, L. (2020). Antibiotic resistance pattern in K.pneumoniae strains isolated from children with urinary tract infections from Tabriz hospitals. *Health Biotechnology and Biopharma*, *4*(1), 38-45. DOI: 10.22034/HBB.2020.03

33. Manjula, N. G., Math, G. C., Nagshetty, K., Patil, S. A., Gaddad, S. M., & Shivannavar, C. T. (2014). Antibiotic susceptibility pattern of ESβL producing K.pneumoniae isolated from urine samples of pregnant women in Karnataka. *Journal of Clinical and Diagnostic Research: JCDR*, 8(10), DC08. https://doi.org/10.7860%2FJCDR%2F2014%2F9594.5048

34. Gupta, N., Kundra, S., Sharma, A., Gautam, V., & Arora, D. R. (2007). Antimicrobial susceptibility of uropathogens in India. *J Infect Dis Antimicrob Agents*, 24(1), 13-18.
<u>https://www.researchgate.net/profile/Dr-Naveen-Gupta/publication/</u>
<u>283256069\_Antimicrobial\_susceptibility\_of\_uropathogens\_in\_India/links/</u>
<u>562f33c108ae518e3484073d/Antimicrobial-susceptibility-of-uropathogens-in-India.pdf</u>

35. Shatalov, A. (2015). Prevalence and antibiotic resistance pattern of Escherichia coli and K.pneumoniae in urine tract infections at the La Paz Medical center, Malabo, Equatorial Guinea. *Open Journal of Medical Microbiology*, 5(04), 177. <u>http://dx.doi.org/10.4236/ojmm.2015.54022</u>

36. Moghadam, M. T., Shariati, A., Mirkalantari, S., & Karmostaji, A. (2020). The complex genetic region conferring transferable antibiotic resistance in multidrug-resistant and extremely drug-resistant K.pneumoniae clinical isolates. *New Microbes and New Infections*, *36*, 100693. https://doi.org/10.1016/j.nmni.2020.100693

37. Chakraborty, S., Mohsina, K., Sarker, P. K., Alam, M. Z., Karim, M. I. A., & Sayem, S. A. (2016). Prevalence, antibiotic susceptibility profiles and ESBL production in K.pneumoniae and Klebsiella oxytoca among hospitalized patients. *Periodicum biologorum*, *118*(1). https://doi.org/10.18054/pb.v118i1.3160 38. Amraie, H., Shakib, P., Rouhi, S., Bakhshandeh, N., & Zamanzad, B. (2014). Prevalence assessment of magA gene and antimicrobial susceptibility of K.pneumoniae isolated from clinical specimens in Shahrekord, Iran. *Iranian Journal of Microbiology*, *6*(5), 311. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4385570/pdf/IJM-6-311.pdf

39. Marques, C., Menezes, J., Belas, A., Aboim, C., Cavaco-Silva, P., Trigueiro, G., ... & Pomba, C. (2019). K.pneumoniae causing urinary tract infections in companion animals and humans: population structure, antimicrobial resistance and virulence genes. *Journal of Antimicrobial Chemotherapy*, 74(3), 594-602. <u>https://doi.org/10.1093/jac/dky499</u>

40. Aminul, P., Anwar, S., Molla, M. M. A., & Miah, M. R. A. (2021). Evaluation of antibiotic resistance patterns in clinical isolates of K.pneumoniae in Bangladesh. *Biosafety and Health*, *3*(6), 301-306. <u>https://doi.org/10.1016/j.bsheal.2021.11.001</u>

41. Nirwati, H., Sinanjung, K., Fahrunissa, F., Wijaya, F., Napitupulu, S., Hati, V. P., ... & Nuryastuti, T. (2019, December). Biofilm formation and antibiotic resistance of K.pneumoniae isolated from clinical samples in a tertiary care hospital, Klaten, Indonesia. In *BMC proceedings* (Vol. 13, No. 11, pp. 1-8). BioMed Central. <u>https://doi.org/10.1186/s12919-019-0176-7</u>

42. Mofolorunsho, K. C., Ocheni, H. O., Aminu, R. F., Omatola, C. A., & Olowonibi, O. O. (2021). Prevalence and antimicrobial susceptibility of extended-spectrum beta lactamases-producing Escherichia coli and K.pneumoniae isolated in selected hospitals of Anyigba, Nigeria. *African Health Sciences*, *21*(2), 505-512. https://doi.org/10.4314/ahs.v21i2.4

43. Najjuka, C. F., Kateete, D. P., Kajumbula, H. M., Joloba, M. L., & Essack, S. Y. (2016). Antimicrobial susceptibility profiles of Escherichia coli and K.pneumoniae isolated from outpatients in urban and rural districts of Uganda. *BMC research notes*, *9*, 1-14. DOI 10.1186/s13104-016-2049-8

44. Maina, D., Makau, P., Nyerere, A., & Revathi, G. (2013). Antimicrobial resistance patterns in extended-spectrum  $\beta$ -lactamase producing Escherichia coli and K.pneumoniae isolates in a private tertiary hospital, Kenya. *Microbiology Discovery*, *1*(5), 1.<u>http://dx.doi.org/10.7243/2052-6180-1-5</u>

45. Mansury, D., Motamedifar, M., Sarvari, J., Shirazi, B., & Khaledi, A. (2016). Antibiotic susceptibility pattern and identification of extended spectrum  $\beta$ -lactamases (ESBLs) in clinical isolates of K.pneumoniae from Shiraz, Iran. *Iranian journal of microbiology*, 8(1), 55.

https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4833741/pdf/IJM-8-55.pdf

46. Shadkam, S., Goli, H. R., Mirzaei, B., Gholami, M., & Ahanjan, M. (2021). Correlation between antimicrobial resistance and biofilm formation capability among K.pneumoniae strains isolated from hospitalized patients in Iran. *Annals of Clinical Microbiology and Antimicrobials*, 20, 1-7. https://doi.org/10.1186/s12941-021-00418-x

47. Varghese, A., George, S., Gopalakrishnan, R., & Mathew, A. (2016). Antibiotic susceptibility pattern of K.pneumoniae isolated from cases of urinary tract infection in a tertiary care setup. *Journal of Evolution of Medical and Dental Sciences*, *5*(29), 1470-1474. DOI: 10.14260/jemds/2016/346

48. Kader, A. A., & Kumar, A. (2005). Prevalence and antimicrobial susceptibility of extendedspectrum β-lactamase-producing Escherichia coli and K.pneumoniae in a general hospital. *Annals of Saudi medicine*, 25(3), 239-242. <u>https://doi.org/10.5144/0256-4947.2005.239</u>

49. Beyene, G., & Tsegaye, W. (2011). Bacterial uropathogens in urinary tract infection and antibiotic susceptibility pattern in jimma university specialized hospital, southwest ethiopia. *Ethiopian journal of health sciences*, *21*(2), 141-146. <u>https://doi.org/10.4314/ejhs.v21i2.69055</u>

50. Moini, A. S., Soltani, B., Ardakani, A. T., Moravveji, A., Erami, M., Rezaei, M. H., & Namazi, M. (2015). Multidrug-resistant Escherichia coli and K.pneumoniae isolated from patients in Kashan, Iran. *Jundishapur journal of microbiology*, 8(10). <u>https://doi.org/10.5812%2Fjjm.27517</u>

51. David, S., Reuter, S., Harris, S. R., Glasner, C., Feltwell, T., Argimon, S., ... & Grundmann, H. (2019). Epidemic of carbapenem-resistant K.pneumoniae in Europe is driven by nosocomial spread. *Nature microbiology*, *4*(11), 1919-1929. https://doi.org/10.1038/s41564.019.0492.8

https://doi.org/10.1038/s41564-019-0492-8

52. Clegg, S., & Murphy, C. N. (2017). Epidemiology and virulence of K.pneumoniae. *Urinary Tract Infections: Molecular Pathogenesis and Clinical Management*, 435-457. https://doi.org/10.1128/9781555817404.ch18

53. Matovina, M., Abram, M., Repac-Antić, D., Knežević, S., & Bubonja-Šonje, M. (2021). An outbreak of ertapenem-resistant, carbapenemase-negative and porin-deficient ESBL-producing K.pneumoniae complex. *Germs*, *11*(2), 199. <u>https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8373410/</u>

54. Oelschlaeger, T. A., & Tall, B. D. (1997). Invasion of cultured human epithelial cells by K.pneumoniae isolated from the urinary tract. *Infection and immunity*, 65(7), 2950-2958. https://doi.org/10.1128/iai.65.7.2950-2958.1997

55. Merino, S., Camprubí, S., Albertí, S., Benedí, V. J., & Tomas, J. (1992). Mechanisms of K.pneumoniae resistance to complement-mediated killing. *Infection and immunity*, *60*(6), 2529-2535. https://doi.org/10.1128/iai.60.6.2529-2535.1992

56. Jay, S. J. (1983). Nosocomial pneumonia: The challenge of a changing clinical spectrum. *Postgraduate Medicine*, 74(2), 221-235.<u>https://doi.org/10.1080/00325481.1983.11698392</u>

57. Nassif, X. A. V. I. E. R., & Sansonetti, P. J. (1986). Correlation of the virulence of K.pneumoniae K1 and K2 with the presence of a plasmid encoding aerobactin. *Infection and immunity*, *54*(3), 603-608. <u>https://doi.org/10.1128/iai.54.3.603-608.1986</u>

58. Pourbaghi, E., Doust, R. H., Rahbar, M., & Farzami, M. R. The Frequency of Integrons and OXA Genes in Uropathogenic Isolates of K.pneumoniae: Transmission of antimicrobial resistance in K.pneumoniae. *Archives of Medical Laboratory Sciences*, *7*, 1-8. https://doi.org/10.22037/amls.v7.35178

59. Xu, L., Sun, X., & Ma, X. (2017). Systematic review and meta-analysis of mortality of patients infected with carbapenem-resistant K.pneumoniae. *Annals of clinical microbiology and antimicrobials*, *16*, 1-12. DOI: 10.1186/s12941-017-0191-3

60. Filgona, J., Banerjee, T., & Anupurba, S. (2015). Role of efflux pumps inhibitor in decreasing antibiotic resistance of K.pneumoniae in a tertiary hospital in North India. *The Journal of Infection in Developing Countries*, *9*(08), 815-820.

https://doi.org/10.3855/jidc.6216\_