Mitochondrial 3243 A to G Mutation: Clinical Phenotypes on Different Stages of Life and Their Relevance to Heteroplasmy

By

Md. Abu Saleh 20146014

A thesis submitted to the School of Pharmacy in partial fulfillment of the requirements for the degree of Bachelor of Pharmacy (B. Pharm.)

School of Pharmacy Brac University September 2024

©2024. Brac University All rights reserved.

Declaration

It is hereby declared that

- 1. The thesis submitted is my own original work while completing degree at Brac University.
- 2. The thesis does not contain material previously published or written by a third party, except where this is appropriately cited through full and accurate referencing.
- 3. The thesis does not contain material which has been accepted, or submitted, for any other degree or diploma at a university or other institution.
- 4. I have acknowledged all main sources of help.

Student's Full Name & Signature:

Md. Abu Saleh 20146014

Approval

The thesis titled "Mitochondrial 3243 A to G Mutation: Clinical Phenotypes on Different Stages of Life and Their Relevance to Heteroplasmy" submitted by Md. Abu Saleh (20146014) of Spring, 2020 has been accepted as satisfactory in partial fulfillment of the requirement for the degree of Bachelor of Pharmacy (B.Pharm.) on September 2024.

Supervised By:

Supervised By:

Mohd. Raeed Jamiruddin, PhD Associate Professor School of Pharmacy Brac University

Approved By:

Dean:

A.F.M. Yusuf Haider, PhD Acting Dean, School of Pharmacy Professor, Department of Mathematics and Natural Sciences Brac University

Ethics Statement

This project does not involve any kind of animal and human trial.

Abstract

Mitochondrial 3243 A to G mutation is a major mutation, genetically causing mitochondrial diseases. It was first found in 1990 in a MELAS patient. One of the most prevalent pathogenic mutations, mitochondrial 3243 A to G mutation in mitochondrial DNA (mtDNA), is frequently linked to a variety of clinical phenotypes. The degree and prevalence of these symptoms differ, depending on variables including heteroplasmy levels (the percentage of mutant mtDNA), and phenotypes mainly include life stage. Clinical lactic acidosis, mitochondrial encephalomyopathy, stroke-like episodes (MELAS), maternally inherited diabetes and deafness, MERRF, CPEO and asymptomatic carriers as well as severe multisystem diseases. Till now, a lot of cohort studies have been performed to identify phenotypes connected with this mutation and a lot of phenotypes were found. Additionally, heteroplasmy level related studies were also performed. Different levels of heteroplasmy showed different phenotypes at different stages of life. The purpose of this study is to identify the clinical characteristics of the mutation at various phases of life and to emphasize the significance of heteroplasmy in the development and prognosis of disease.

Keywords: Mutation; Mitochondrial DNA; Point mutation; MELAS; MIDD; MERRF; CPEO; Heteroplasmy; Clinical phenotype; Diabetes; Deafness; Leukocyte; Cardiomyopathy.

Acknowledgement

I would like to thank the Almighty Allah at first, who is the Most Gracious, Most Merciful, the provider of all of our strength and knowledge which have enabled me to complete this project with full diligence.

Secondly, I would like to convey my regard, gratitude and honor to my most respected supervisor, Mohd. Raeed Jamiruddin, PhD, associate professor, Brac University. It was a great opportunity for me to work under his supervision and direction. His persuasive words have inspired me to take new challenges while doing research and to complete this project.

I would like to express my gratitude and gratefulness to all the faculties of the School of Pharmacy, Brac University.

I would like to also express my gratitude towards my parents who supported me morally and in every possible way throughout this long course.

Last but not the least, I would also like to acknowledge all the respected teaching assistants and lab officers.

Dedication

Dedicated to Syed Golamur Rahman Maizvandari

Table of Contents

Declarationii
Approvaliii
Ethics Statementiv
Abstractv
Acknowledgementvi
Dedicationvii
List of Tablesx
List of Figurexi
List of Acronymsxii
Chapter 1 Introduction1
1.1 Mitochondria1
1.2 Mitochondrial DNA1
1.3 Mitochondrial Point Mutation2
1.4 Mitochondrial 3243 A to G Mutation4
1.5 Aim of the study
Chapter 2 Methodology
2.1 Search Strategies
2.2 Criteria for Selection of Articles
Chapter 3 Result11
3.1 Tabulation of the selected studies

3.2 Statistical Presentation of Phenotypes	29
3.3 Identified Phenotypes and Their Prevalence	30
3.4 Clinical Phenotypes at Different Stages of Life	30
3.5 Correlation Between Phenotypes and Heteroplasmy Level	33
Chapter 4 Discussion	35
Chapter 5 Limitation of the Study	
Chapter 6 Conclusion	40

References

List of Tables

Table 1: Inclusion and Exclusion Criteria.	9
Table 2: Selected studies	12

List of Figures

Figure 1: Human mitochondrial DNA	2
Figure 2: Mitochondrial point mutations	4
Figure 3: Mitochondrial 3243 A to G mutation	5
Figure 4: Prisma flow diagram of identification of articles via database	10
Figure 5: Statistical presentation of phenotypes	29
Figure 6: Phenotypes at different stages of life	31
Figure 7: Correlation between phenotypes and heteroplasmy level	34

List of Acronyms

DNA	Deoxyribonucleic acid
ATP	Adenosine triphosphate
RNA	Ribonucleic acid
MELAS	Mitochondrial Encephalomyopathy, Lactic Acidosis and Stroke-like
	episodes
MIDD	Maternally Inherited Diabetes and Deafness
MERRF	Myoclonic epilepsy with ragged red fibers
CPEO	Chronic progressive external ophthalmoplegia
DM	Diabetes Mellitus
BMI	Body Mass Index
FPG	Fasting Plasma Glucose
ESRD	End Stage Renal Disease
RPA	Retinal Pigmentary Abnormalities
MRI	Magnetic Resonance Imaging
DGGE	Denaturing Gradient Gel Electrophoresis
Anti-GAD	Antibodies to Glutamic Acid Decarboxylase

Chapter 1

Introduction

1.1 Mitochondria

Mitochondria, which are membrane-bound organelles within cells, generate most of the chemical energy needed for cellular metabolism. This energy is stored in a small molecule called adenosine triphosphate (ATP), produced by the mitochondria. These organelles also have their own small set of chromosomes. Typically, mitochondrial DNA is only inherited from the mother. The size of mitochondria is generally between 0.75 to 3 micrometers, and unless they are stained, they are invisible under microscope (Newman, 2018). The number of mitochondria differs among various cell types. While they are often shown as oval-shaped organelles, mitochondria are constantly undergoing division (fission) and merging (fusion). Consequently, these organelles form dynamic, interconnected networks.

1.2 Mitochondrial DNA

Unlike other organelles in mammalian cells, mitochondria contain a small amount of DNA known as mitochondrial DNA (mtDNA), which encodes several key proteins necessary for mitochondrial respiration. A range of proteins, including prohibitins, mitochondrial transcription factor A (TFAM), ATPase family AAA domain-containing protein 3 (ATAD3), and POLG (DNA polymerase gamma, catalytic subunit), package mtDNA. While distinct from nuclear DNA (nDNA), mtDNA resembles bacterial chromosomes. These proteins and mtDNA together form a complex known as a nucleoid (Yan et al., 2019). The mtDNA's sense and antisense strands are referred to as the heavy (H) and light (L) strands, respectively. 16,569 base pairs of mtDNA are found in human cells which encode 37 different genes, including 13 polypeptides, 2 ribosomal RNAs, and 22 tRNAs (Nicholls & Gustafsson, 2018). The L strand contains 1 polypeptide (ND6) and 8 tRNAs, while the H strand encodes the remaining 12 polypeptides, 2 rRNAs, and 14 tRNAs. A noncoding element known as a displacement loop (D-loop), which is found in nearly all known cases of mtDNA transcription and replication, is also present in mtDNA (Sbisà et al., 1997).

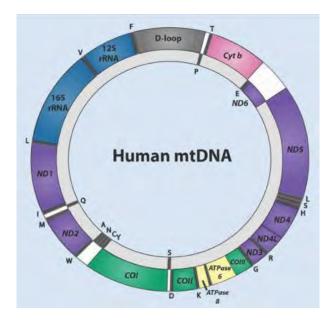


Figure 1: Human mitochondrial DNA

Note. This image was created to depict mitochondrial DNA. From "What is mitochondrial DNA?" by Home. (n.d.). Wellcome Trust Centre for Mitochondrial Research. https://www.newcastle-mitochondria.com/patient-and-public-

home-page/what-is-mitochondrial-dna/

1.3 Mitochondrial Point Mutation

Point mutations in the mtDNA are generally inherited from mothers. They could be found in rRNA, tRNA, or protein genes. Yet mt-tRNA genes contain more than half of the documented

point mutations linked to disease. From a phenotypic perspective, point mutations in genes that code for proteins in the mitochondria directly impact the functioning of the RC complex that the corresponding protein is a part of, whereas mutations in mt-tRNAs can alter the availability of functional mt-tRNAs, which may affect the general efficacy of mitochondrial translation. Point mutations are thought to be largely recessive, with the majority being heteroplasmic and exhibiting significant clinical variability. On the other hand, more and more harmful homoplasmic mutations are being identified; these mutations frequently impact only one tissue and exhibit partial penetration (Tuppen et al., 2010). There is a significant chance that mitochondrial DNA will come into contact with oxidative chemicals in this atmosphere. Superoxide radicals, which may damage DNA, are mostly found in the mitochondrial respiratory chain. Moreover, the mitochondrial membrane's monoamine oxidase produces hydrogen peroxide, which has the potential to be a genotoxic substance. It means that the high rate of mutation in mitochondrial DNA and the accumulation of mutations with age are not surprising (Rustin et al., 2000). A lot of mitochondrial mutations are identified. Among them mitochondrial 3243 A to G mutation is a remarkable one.

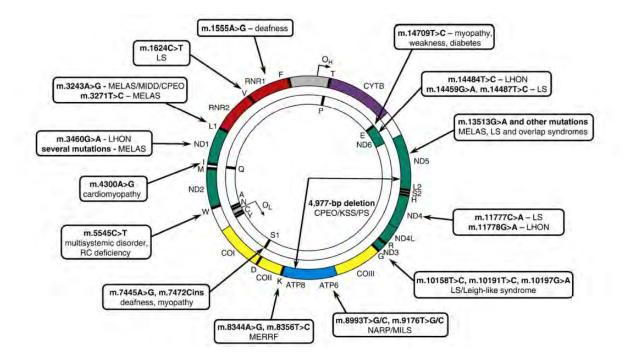


Figure 2: Mitochondrial point mutations.

Note. This image was created to depict mitochondrial point mutations. From "Mitochondrial DNA mutations and human disease" by Tuppen, H. A. L., Blakely, E. L., Turnbull, D. M., & Taylor, R. W. (2010). Mitochondrial DNA mutations and human disease. Biochimica et Biophysica Acta (BBA) - Bioenergetics, 1797(2), 113–128. https://doi.org/10.1016/j.bbabio.2009.09.005

1.4 Mitochondrial 3243 A to G Mutation

Mitochondrial 3243 A to G mutation is basically a nucleotide change from adenine to guanine at 3243 position in the mitochondrial tRNA leucine (UUR) gene (MT-TL1) of the mitochondrial genome. This is the most common point mutation which has been reported with Mitochondrial Encephalomyopathy, Lactic Acidosis and Stroke-like episodes (MELAS) syndrome(Gorman et al., 2015). The mutation was first identified by Goto and colleagues (Goto et al., 1990). The mutation is seen in heteroplasmic state mostly. Heteroplasmy is such a state where the mutations

are found in only a portion of the entire mtDNA population (Maassen, 2002). A cell that has both mutant and wild-type mtDNA genomes is said to be heteroplasmic. By means of somatic mutagenesis and continuous mtDNA replication, mutations can proliferate clonally by stochastic or selective processes, and manifest in different ratios or levels of heteroplasmy inside cells (Elson et al., 2001). Based on heteroplasmy level in different tissues, different types of phenotypes are seen. However, the mutation has significant molecular and functional consequences.

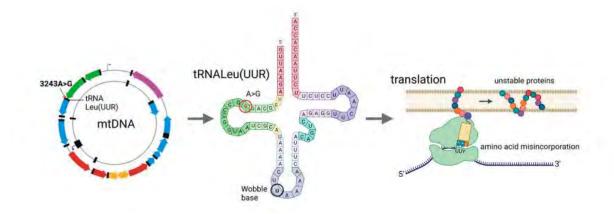


Figure 3: Mitochondrial 3243 A to G mutation.

Note. This image was created to depict mitochondrial 3243 A to G mutation. From "The Mitochondrial m.3243A>G Mutation on the Dish, Lessons from In Vitro Models," by Ryytty, S., & Hämäläinen, R. H. (2023). The Mitochondrial m.3243A>G Mutation on the Dish, Lessons from In Vitro Models. International Journal of Molecular Sciences, 24(17), 13478. <u>https://doi.org/10.3390/ijms241713478</u>

The MT-TL1 gene, which codes for the mitochondrial tRNA Leu(UUR), a 75 bp long tRNA for leucine, is impacted by the m.3243A>G mutation (Li et al., 2022a). The adenine-to-guanine mutation at position 3243 affects the D-loop of the tRNALeu(UUR) molecule, altering its stability, structure, and codon recognition. Specifically, the m.3243A>G mutation reduces UUG translation without significantly impacting UUA translation, causing amino acid misincorporation in all

mitochondrial proteins encoded by mtDNA (Chomyn et al., 2000). Additionally, this mutant lacks post-transcriptional taurine modifications at the wobble U base. The resulting unstable protein products lead to defects in the electron transport chain. The mutation primarily impacts proteins with abundant UUG codons, such as the ND6 subunit of complex I (Li et al., 2022b). As this mutation is heteroplasmic-that is, it coexists in cells with both mutant and non-mutated mtDNA—it is linked to a wide range of clinical outcomes. The amount of mutant to wild-type mtDNA in various tissues affects how severe the symptoms are. Fibroblast cells produced from patients offer a direct connection to the physiology of the patient; yet, their capacity to completely recapitulate disease phenotypes is limited, especially for complex tissues. In patient cultures, fibroblast proliferation is impacted. It has been observed that fibroblasts with a 30% mutation load proliferate at a slower rate (Chung et al., 2021), and that fibroblasts with a high mutation frequency (>95%) have much lower growth potential (Yokota et al., 2015). For the purpose of isolating effects specific to mitochondria, cybrid (cytoplasmic hybrid) cells are used, in which mutant mtDNA is inserted into a host cell with a different nuclear background. Also, highly proliferative cells can be utilized. However, these models might not accurately represent the interactions that take place in vivo between the nuclear and mitochondrial genomes. Additionally, the tolerance of various cell types to the m.3243A>G mutation varies (Ryytty & Hämäläinen, 2023).

1.5 Aim of the Study

The aim of the study is to find out prominent clinical phenotypes in individuals harboring mitochondrial 3243 A to G mutation during different stages of life and the correlation of the phenotype and heteroplasmy level. The study seeks most of the phenotypes and a statistical analysis of them. These phenotypes are found at different stages of life. So, building up a general concept about the onset of diseases is a goal. The study also aims to relate the phenotypes with the

heteroplasmy level of the mutation. Till date there is no established correlation found between heteroplasmy level and phenotype. So, the study intends to set a general correlation between heteroplasmy level and phenotype analyzing previous research in this field. The study contains information of 20 different studies conducted upon patients harboring the mutation. Next, it contains the phenotypes which are seen at different stages of life. Finally, it contains a relationship between phenotype and heteroplasmy level.

Chapter 2

Methodology

2.1 Search strategies

For collecting articles reputed and renowned sources were used. Sources were PubMed, Google Scholar, ELSEVIER, NCBI, Research gate, Springer and ScienceDirect. To get the desired articles, a search string was generated using the major keywords which were- mitochondrial 3243 A to G mutation, MELAS, MIDD, mitochondrial mutation, MERRF, CPEO and clinical phenotypes. Boolean logic using "and" and "or" was applied to combine the keywords, enhancing the accuracy of the search. As the mitochondrial 3243 A to G mutation was identified in 1990, there have been a few studies done. So, filters were not used.

2.2 Criteria for Selection of Articles

Certain inclusion and exclusion criteria were applied to select the journal articles. Below, table 1 represents the inclusion and exclusion criteria. The process of identifying articles through the database is illustrated with a PRISMA flow diagram in Figure 4.

Parameters	Inclusion Criteria	Exclusion Criteria
Patient	Patients harboring	Patients having phenotypes
	mitochondrial 3243 A to G	but not the mutation.
	mutation. Or those patients	
	who had phenotypes before	
	and the mutation was found	
	later.	
Relevance	Articles relevant to only	Articles relevant to
	mitochondrial 3243 A to G	mitochondrial mutations
	mutation.	other than 3243 A to G
		mutation.
Publication date range	Articles from 1990 to 2024.	Articles before 1990 as the
		mutation was first identified
		in 1990.
Availability	Articles that were available	Articles that were not
	through different ways.	available at all.
Language	Articles written in English	Articles in languages other
		than English.

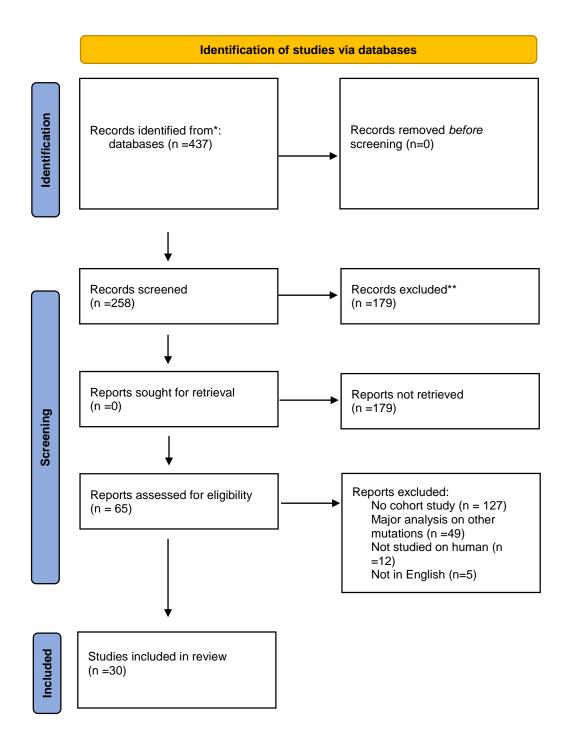


Figure 4: Prisma flow diagram of identification of articles via database

Chapter 3

Result

3.1 Tabulation of the selected studies

Table 2: Selected Studies

Se	Sampl	Control	Tested	Primary Outcomes	Secondary Outcomes	Phenotypes	Citation
ria	e	Populat	Populat				
1	Numb	ion	ion				
No	er	Numbe	Numbe				
		r	r				
1	1079	309	770	1.Mitochondrial 3243 A to G	1. Among 309 healthy people,	1.Diabetes	(Wang et
	subject	healthy	diabete	mutation was present in 13 patients.	no mitochondrial 3243 A to G	mellitus	al., 2013)
	s	controls	S	2. The mitochondrial 3243 A to G	mutation was present.		
			mellitus	mutation was more common in DM	2. Younger age was associated		
			(DM)	patients compared to healthy	with a higher likelihood of DM		
			patients	controls; this was correlated with	patients with the mitochondrial		
				DM patients' younger ages, lower	3243 A to G mutation.		
				BMIs, and higher FPG levels.			

2	113	0	113	1.32.8 years was the mean age of the	1. Maternal inheritance of	1.Materneally	(Suzuki et
	patient		patients	diagnosis of diabetes.	glucose intolerance was linked	inherited	al., 2003)
	S		harbori	2.12.9 years was the mean duration	to early middle-aged diabetes	diabetes and	
	harbor		ng	of diabetes.	development, decreased insulin	deafness.	
	ing		mitoch	3. Approximately 68% of the	secretory capability, and early	2. High	
	mitoch		ondrial	patients with the mutation who had	insulin therapy requirements.	incidence of	
	ondrial		3243 A	diabetes had maternal inheritance,	This was shown in 75% of	neurosensory	
	3243		to G	which was found to be substantially	mutant diabetic patients.	deafness	
	A to G		mutatio	greater than paternal inheritance.		3.Early middle	
	mutati		n.			aged onset of	
	on.					diabetes and	
						deafness.	

3	151	0	151	1.The number of symptom carrier	1.The Mitochondrial 3243 A to	1.Melas (7	(de Laat et
	patient		patients	patient is 124.	G mutation causes a slowly	patients)	al., 2021)
	S		harbori	2. The degree of heteroplasmy in	progressive disease.	2.MIDD (60	
	harbor		ng	leucocytes and Urea Electrolytes		patients)	
	ing		mitoch	and Creatinine (UEC) was only			
	mitoch		ondrial	weakly correlated to the severity of			
	ondrial		3243 A	the illness.			
	3243		to G				
	A to G		mutatio				
	mutati		n				
	on						
4	15	0	15	1.The degree of heteroplasmy	1.The heteroplasmy and age-	1.Hearing loss	(Sakata et
	patient		patients	ranged from 3% to 37% and the	corrected heteroplasmy levels	2.Diabetes	al., 2022)
	S		harbori	mean was 23.9%.	were linked with the age at	mellitus	
	harbor		ng	2. The mean age at which hearing	which hearing loss first	3.Balance-gait	
	ing		mitoch	loss first appeared was 28.6 years; in	appeared.	disorder.	

	mitoch	ondrial	the earliest cases, acquired hearing	2.The mitochondrial 3243 A to		
	ondrial	3243 A	loss was detected as early as age 10,	G mutation results in gradual		
	3243	to G	and in more latest cases, it was	hearing loss.		
	A to G	mutatio	detected as late as age 56.			
	mutati	n	3.In seven individuals, their hearing			
	on		rapidly declined to total deafness			
			between the ages of 40 and 63.			
			4. In all 15 patients the average rate			
			of the development of hearing loss			
			was 5.5db per year.			
5	53 0	53	1.Most patients (41.5%) were	1.Individuals with stable high	1.Fatigue	(Klein et
	patient	patients	categorized into the stable high	fatigue had worse clinical	2.Mental health	al., 2022)
	s	harbori	fatigue trajectory, followed by the	performance and more (severe)	issues	
	harbor	ng	fluctuating fatigue group at 35.9%,	organ system involvement than		
	ing	mitoch	and the stable low fatigue group at	individuals with a consistent		
	mitoch	ondrial	22.6%.	low fatigue trajectory.		

	ondrial		3243 A	2. The stable high fatigue group had	2.Patients experiencing severe		
	3243		to G	a higher percentage of female	fatigue have a heavier burden,		
	A to G		mutatio	patients (91%) compared to the	impacting various aspects of		
	mutati		n	fluctuating fatigue group (58%).	daily life such as mental health,		
	on			3. Patients who had consistent high	functioning, and overall quality		
				levels of fatigue had noticeably	of life compared to those with		
				greater issues with their	mild fatigue.		
				psychological and mental health.			
6	227	92	135	1.Among the 135 patients with	1. Type II diabetes was the	1.Diabetes	(Iwasaki
	individ	individ	patients	diabetes and end-stage renal disease	initial diagnosis of all	2.Neuropathy	et al.,
	uals	uals	with	(ESRD), 8 patients had the mutation	participants carrying the	3.Retinopathy	2001)
			diabete	and it was in heteroplasmic form.	mitochondrial 3243 A to G	4.Proteinuria	
			s and	2. Among the 92 patients with	mutation.	5.Deafness	
			End	ESRD alone, none had a		6.MELAS	
			stage	mitochondrial 3243 a to g mutation.			
			renal				

			disease(
			ESRD).				
7	14	0	14	1.Retinal pigmentary abnormalities	1. It is probably justified to	1.Retinal	(Sue et al.,
	patient		patients	(RPAs) are highly prevalent in	classify retinal pigmentary	Pigmentary	n.d.)
	S		harbori	patients with the mitochondrial 3243	abnormalities as a component	abnormality	
	harbor		ng	A to G mutation, occurring in 8	of the MELAS syndrome	2.Deafness	
	ing		mitoch	patients (57%) of these patients.	clinical spectrum.	3.Diabetes	
	mitoch		ondrial	2. Among the patients with	2. Concomitant optic atrophy	4.Epilepsy	
	ondrial		3243 A	pigmentary retinal abnormalities,	was absent in all individuals.	5.Stroke-like	
	3243		to G	only one had corrected visual acuity		episodes	
	A to G		mutatio	that was moderately impaired		6.Proximal	
	mutati		n	(20/60); all other patients had		myopathy	
	on			corrected visual acuity that was		7.External	
				20/30 or better.		ophthalmoplegia	

8	91	0	91	1.The mitochondrial 3243 A to G	1.Detectable mtDNA deletions	1.Progressive	(Moraes et
	patient		patients	mutation was found in 21 patients.	in muscle were present in none	external	al., 1993)
	s with		with	2.Progressive external	of the patients.	ophthalmoplegia.	
	mitoch		mitoch	ophthalmoplegia was present in 16		2.Ptosis	
	ondrial		ondrial	of the 21 patients (76%).		3.Proximal	
	myopa		myopat			weakness	
	thies		hies			4.Stroke	
						5.Deafness	
						6.Seizures	
						7.Ataxia	
						8.Retinopathy	
						9.Endocrinopath	
						у	
9	22	0	62	1.In the families of probands 14	1.In individuals with Fatigue	1.Limb weakness	(Hamman
	proban		individ	patients were found harboring	compared to those with	2.Deafness	s et al.,
	ds and		uals		MELAS syndrome, the	3.Ataxia	n.d.)

their	mitochondrial 3243 A to G	percentage of COX negative	4.Short stature
familie	mutation.	fibers were significantly higher,	5.Seizures
s (62	2. The mitochondrial 3243 A to G	whereas the frequency of	6.Ophthalmopleg
patient	mutation was identified in one or	highly COX positive fibers was	ia or ptosis
s total)	more tissues from 22 unrelated	the opposite.	7.Multiple
	index patients across 22 families.		stroke-like
	3. Age of onset and clinical severity	2. The blood's mutant	episodes
	score were significantly correlated	percentage of mitochondrial	8.Dementia
	with the amount of mutant mtDNA	3243 A to G mutation is	9.Retinopathy
	in muscle and blood.	probably going decline as	10.Diabetes
		individuals being old.	11.Neuropathy
			12.Cardiomyopat
		3. There could be significant	hy
		changes in heteroplasmy across	13.Wolff-
		generations.	Parkinson-White
			syndrome

						14.Gastrointestin	
						al pseudo-	
						obstruction	
10	41	25	16	1.Mitochondrial 3243A to G	1.Serum, plasma, and blood	1.Type 2	(Zhong,
	subject	healthy	patients	mutation score was positive in	leukocyte samples were	diabetes.	2000)
	s	subjects	with	seven plasma and serum samples	examined for their levels of		
			type 2	from patients with type 2 diabetes	mitochondrial 3243A to G		
			diabete	while the 25 samples from healthy	mutation heteroplasmy. The		
			S	subjects did not display any mutant	seven affected patients had		
				bands, also, the other nine samples	varying percentages of		
				from diabetic patients also showed	mitochondrial 3243 A to G		
				no mutant bands.	mutation in their blood		
					leukocytes (1.1%–13.5%),		
					serum (1.5%–35.2%), and		
					plasma (1.6%–36.5%).		

11	6	6	6	1. All of the patients and their	1. The six patients' parents were	1.Hypertrichosis	(Ma et al.,
	childre	individ	individ	mothers had a mitochondrial 3243 A	healthy non-consanguineous	2.Headache	2013)
	n and	uals	uals	to G mutation.	individuals.	3.Vomiting	
	their			2. Six patients had mutation loads	2. Four patients' brain magnetic	4.Blurred vision	
	mother			ranging from 43.6% to 58%.	resonance imaging (MRI)	5.Epilepsy	
	s			3.Among their mothers, 14.1% to	revealed hyperintense signals	6.Stroke-like	
	(Total			28.6% did not exhibit any	in the asymmetric parietal,	episode	
	12)			symptoms.	temporal, and occipital lobes on		
				4. In all patients hyperlactemia was	T2-weighted scans.		
				found.			
				5. All patients were found with			
				mixed neurogenic and			
				myopathic change and it was			
				detected by electromyography.			

12	212	90	122	1.By DGGE analysis, only one	1.In any of the healthy controls	1.Diabetes	(Klemm et
	individ	individ	diabetic	patient was found harboring	mitochondrial 3243 A to G	mellitus type 1	al., 2001)
	uals	uals	patients	mitochondrial 3243 A to G	mutation was not detected.	2. Hearing loss	
				mutation.		3.Diabetes	
				2. The affected patient was a woman		mellitus type 2	
				who was 63 years old. She was			
				diagnosed with diabetes at the age of			
				57.			
13	17	0	17	1.5 patients were found diagnosed	1. A MELAS patient died	1.MELAS	(Anan et
	unrelat		unrelate	with MELAS.	because he had severe	2.Left ventricular	al., 1995)
	ed		d	2.All of them 5 MELAS patients had	concentric hypertrophy of the	hypertrophy	
	patient		patients	a mitochondrial 3243 A to G	left ventricle which resulted in	3.Wall motion	
	s with		with	mutation.	and heart failure 2. It was	abnormalities	
	mitoch		mitoch		reported to have had 83% of	4.Occasional	
	ondrial		ondrial		mutant genomes in the heart.	palpitation	

	diseas		disease		2.In MELAS patients,		
	es		S		similarity was seen in the		
					percentage of mutant genomes		
					in clinically affected and		
					unaffected tissues.		
14	6	0	6	1.Two were diagnosed with MIDD	1.In patients with the	1.Vestibular	(Inoue et
	patient		patients	and four with MELAS.	mitochondrial 3243 A to G	symptoms	al., 2019)
	S		harbori	2. In peripheral leukocytes, the	mutation, hearing loss was	2.Diabetes	
	harbor		ng	heteroplasmy rate of the A3243G	attributed to cochlear	3.Hearing loss	
	ing		mitoch	mutation varied from 9 to 32%.	dysfunction rather than damage		
	mitoch		ondrial	3. During the initial examination	to the retrocochlear structures.		
	ondrial		3243 A	with caloric testing, two patients			
	3243		to G	(33%) exhibited normal responses in			
	A to G		mutatio	both ears, two patients (33%) had			
	mutati		n	reduced responses in one ear, and			
	on						

(Smith et
al., 1999)
tati
inal

mutati	minor pigmentary abnormalities that
on	were asymptomatic.
	4.All individuals with pigmentary
	retinal dystrophy and impaired
	electrophysiologic responses were
	older than 40 and had been suffering
	from diabetes over five years.
	5. Among the ten participants with
	abnormal retinal pigmentation,
	seven (70%) had both diabetes and
	deafness, while three (30%),
	including the two youngest
	individuals, were either diabetic or
	deaf.

16	3	0	3	1. All patients and their mothers had	1. Results of biochemical	1.Isolated	(Jiang et
	patient		patients	the mitochondrial tRNA-Leu gene	studies showed that urine	respiratory chain	al., 2015)
	s			mutation m.3243 A>G.	organic acid, ketone bodies,	complex III	
				2. Every patient was born following	carnitine esters, and blood	deficiency.	
				uneventful pregnancy.	glucose were all within normal	2.Hyperlactacide	
					ranges.	mia	
17	4	0	4	1.It was found that the original	1.One patient's best-corrected	1.Ophthalmopleg	(Latkany
	individ		individ	patient, her daughter, and son, as	visual acuities were found to be	ia	et al.,
	uals		uals	well as the sister with MELAS	20/25 and 20/30 on an ocular	2.Neurosensory	1999)
				syndrome, all four tested family	examination.	deafness	
				members had the A to G 3243 mt		3.Myopathy	
				DNA mutation heteroplasmic with		4.Macular retinal	
				wild-type mtDNA.		pigment	
						epithelial	
						atrophy	

18	1	0	1	1. The patient's serum creatinine was	1.While there was a small	1.Hyperlactatemi	(Cai et al.,
	patient		patient	70 umol/L, but his uric acid	amount of malformed	a	2022)
	harbor		harbori	increased unexpectedly to 1011	mitochondria in the renal	2.Proteinuria	
	ing		ng	umol/L when he was in the	tubules, there was no obvious	3.Membranous	
	mitoch		mitoch	emergency room.	accumulation of mitochondria	nephropathy	
	ondrial		ondrial	2. A thicker and more stiff	in the podocytes.		
	3243		3243 A	glomerular basement membrane			
	A to G		to G	was seen.			
	mutati		mutatio				
	on		n				
19	1	0	1	1.Mitochondrial 3243 A to G	1.The primary cause of the	1.Diabetes	(Yamamot
	patient		patient	mutation was found in white blood	onset of diabetes in this case	mellitus	o, 2003)
	harbor		harbori	cells.	was a mitochondrial DNA	2.Cardiomyopath	
	ing		ng	2.Anti-GAD antibody was negative.	mutation-related diabetes	у	
	mitoch		mitoch		mellitus, which is known to	3.Hypothyroidis	
	ondrial		ondrial			m.	

	3243		3243 A	3. The number of a-cells was	produce a severe and rapid drop		
	A to G		to G	growing while that of p-cells was	in insulin secretion.		
	mutati		mutatio	decreasing.			
	on		n				
20	1	0	1	1. 1. The first biopsy of the deltoid	1.Quantification of the	1.MERRF	(Fabrizi et
	patient		patient	muscle revealed 8%–10% of ragged	mitochondrial 3243 A to G		al., 1996)
	harbor		harbori	red fibers.	mutation showed that 78% of		
	ing		ng	2. In the second biopsy of the vastus	the proband's muscle mtDNA		
	mitoch		mitoch	lateralis, ragged red fibers were	was mutated.		
	ondrial		ondrial	reduced to 1%–2%, yet the succinate			
	3243		3243 A	dehydrogenase staining in many of			
	A to G		to G	these fibers was more intense than in			
	mutati		mutatio	normal fibers.			
	on		n				

3.2 Statistical presentation of phenotypes

Table 1 encompasses 20 different studies conducted on individuals harboring mitochondrial 3243 A to G mutation. This includes sample number, control population number, tested population number, primary outcomes, secondary outcomes and phenotypes. Various phenotypes have been found. Statistical presentation is shown in the chart below.

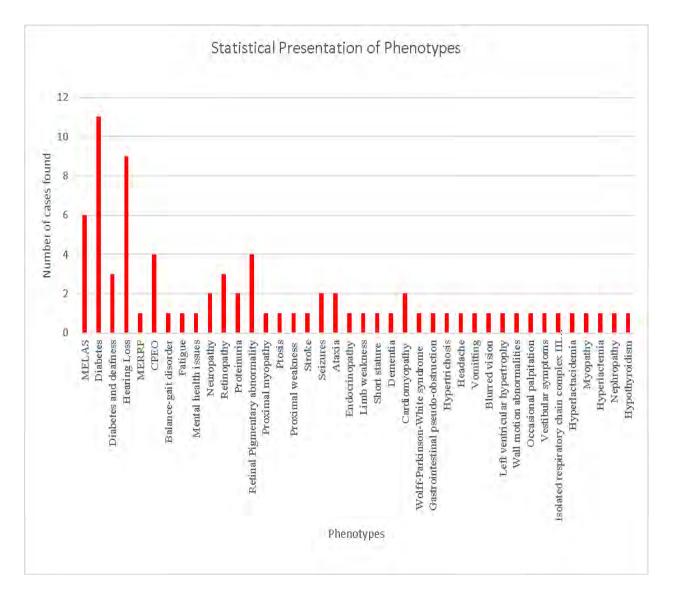


Figure 5: Statistical Presentation of phenotypes

3.3 Identified phenotypes and their prevalence

From the tabulated 20 different studies, 40 different phenotypes were identified. Among them, diabetes is found, the highest, 11 times. Second, hearing loss/deafness is found 9 times. Third, MELAS is found 6 times. Fourth, CPEO and retinal pigmentary abnormality are found 4 times. Fifth, diabetes and deafness as well as retinopathy are found 3 times. Diseases those are found 2 times are- neuropathy, proteinuria, seizure, ataxia and cardiomyopathy. Finally, the diseases those found only once are- MERRF, balance-gait disorder, fatigue, mental health issues, proximal myopathy, ptosis, proximal weakness, stroke, endocrinopathy, limb weakness, short stature, dementia, Wolff-Parkinson-White syndrome, gastrointestinal pseudo-obstruction, hypertrichosis, headache, vomiting, blurred vision, left ventricular hypertrophy, wall motion abnormalities, occasional palpitation, vestibular symptom, isolated respiratory chain complex III deficiency, hyperlactacidemia, myopathy, hyperlactemia, nephropathy and hypothyroidism.

3.4 Clinical phenotypes at different stages of life

The mitochondrial 3243 A to G mutation is the cause of various diseases which manifest at different stages of life. From previous studies it has been found that different clinical features, generally, are seen during different phases. So, a classification is made based on age which are early stage (age<20), middle stage (age 20-40) and end stage (age>40). The following chart represents phenotypes at different stages of life.

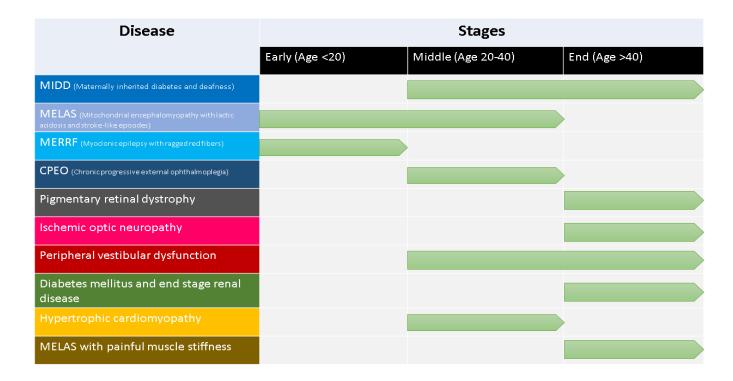


Figure 6: Phenotypes at different stages of life

One of the main clinical phenotypes of this mutation is maternally inherited diabetes and deafness(MIDD). From a study it is found that most patients harboring mitochondrial 3243 A to G mutation develop diabetes around age 35 (Maassen, 2002). From another study it is found that, the average age of developing diabetes due to mitochondrial 3243 A to G mutation is 40.23 ± 3.30 (Wang et al., 2013). The mean age of hearing loss was found 28.6 from another study (Sakata et al., 2022). Which ultimately results in deafness. So, generally MIDD is found at the middle stage and end stage of life in patients harboring the mutation. Another main phenotype of this disease is Mitochondrial encephalomyopathy with lactic acidosis and stroke-like episodes (MELAS). MELAS is usually developed between ages of 2 and 15 (Pia & Lui, 2024). In another study, from 42 MELAS patients the average age of diagnosis was found 33 (Cox et al., 2023). So, generally MELAS is found at the early stage and middle stage of life. Another notable phenotype of the mutation is myoclonic epilepsy with ragged red fibers (MERRF). Generally, it is hard to claim any

specific stage of life to develop MERRF. There has been too little information found. But, a study shows that MERRF is found in patients at an early stage of life (Fabrizi et al., 1996). Next, a disorder of the eye known as chronic progressive external ophthalmoplegia (CPEO) is characterized by a gradually worsening incapacity to move the eyebrows and eyes. In many cases, it is the only indication of mitochondrial disease. From a study, it is observed that the age of onset of CPEO in a patient harboring the mutation is 39 (Greaves et al., 2010). This indicates that CPEO is found in the middle stage of life. Another phenotype of the mutation is pigmentary retinal dystrophy. A study found that all patients with pigmentary retinal dystrophy and impaired electrophysiological responses were over 40 years old (Smith et al., 1999), suggesting that pigmentary retinal dystrophy occurs at the end stage of life. The condition known as ischemic optic neuropathy refers to optic nerve disease resulting from a temporary or permanent interruption of blood flow to any part of the optic nerve (Patil et al., 2022). From a case study, this disease was found in a patient harboring the mutation as first presentation. And, it is observed that ischemic optic neuropathy is found at the end stage of life (Scarcella et al., 2023). The mitochondrial A3243G mutation may influence the peripheral vestibular systems due to the strong connections between the cochlea and the peripheral vestibular end organs in terms of embryology, physiology, and morphology (Jin et al., 2006). Research indicates that patients with this mutation typically begin to experience peripheral vestibular symptoms around the age of 36.5 ± 13.2 years (Inoue et al., 2019), suggesting that peripheral vestibular dysfunction is prevalent in the middle to late stages of life. Recently, this mutation was identified in three diabetic individuals with progressive kidney disease, indicating a potential role in the development of renal disease in diabetic patients. Consequently, a study was conducted to assess the impact of this mutation on the progression to end-stage renal disease (ESRD) among diabetic patients. The study identified eight patients with

the mutation, diabetes, and ESRD, all of whom were over 40 years old (Iwasaki et al., 2001). So, ESRD is found in the end stage of life in patients harboring the mutation. Another phenotype found of the mutation is hypertrophic cardiomyopathy. This phenotype is found from a case study which involves a patient of 36 years old. The patient was harboring the mutation and had hypertrophic cardiomyopathy as phenotype (Hsu et al., 2008). This indicates hypertrophic cardiomyopathy is found at the middle stage of life. Lastly, MELAS with muscle stiffness is found in another case study as the phenotype of the mutation. According to the study, MELAS with muscle stiffness is found at the end stage of life (Deschauer et al., 1999).

3.5 Correlation between phenotypes and heteroplasmy level

Heteroplasmy plays a crucial role in mitochondrial diseases, which are caused by malfunctioning mitochondria (Gorman et al., 2016). The development of these diseases depends on the proportion of mutant mtDNA relative to wild-type mtDNA, known as the heteroplasmy level (Parakatselaki & Ladoukakis, 2021). Phenotypes are mostly heteroplasmy level dependent. The following chart represents the general heteroplasmy level of different phenotypes.

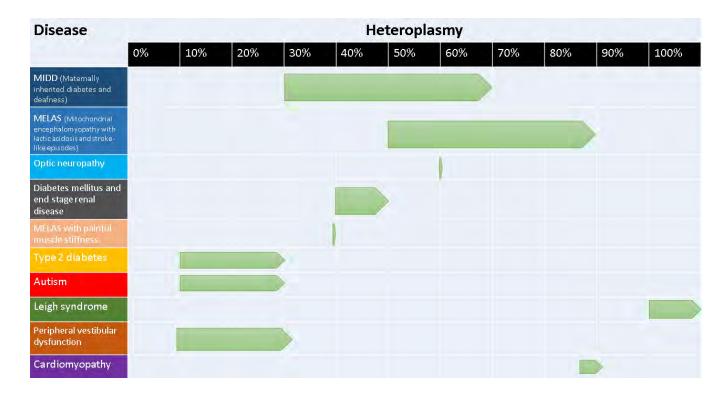


Figure 7: Correlation between phenotypes and heteroplasmy level

Here, discussed levels are not tissue specific as it varies depending upon tissues. Heteroplasmy level for MIDD is 30%-70% (Kyriakidou et al., 2023). Next, heteroplasmy level for MELAS is 50%-90% (Tranah et al., 2018). A study shows that, heteroplasmy level for optic neuropathy is 60% (Motlagh Scholle et al., 2020). Proceeding, heteroplasmy level for diabetes mellitus and end stage renal disease is 40%-50% (Iwasaki et al., 2001). Thereafter, heteroplasmy level for MELAS with painful muscle stiffness is 39% (Deschauer et al., 1999). Then, heteroplasmy level for type 2 diabetes is 10%-30% (van den Ouweland et al., 1992). Next, heteroplasmy level for autism is 10%-30% (Pons et al., 2004). Further, heteroplasmy level for leigh syndrome is (Picard et al., 2014). Next, heteroplasmy level for peripheral vestibular dysfunction is 9%-32% (Inoue et al., 2019). At last, the heteroplasmy level for cardiomyopathy is 87%-91% (Gallego-Delgado et al., 2015).

Chapter 4

Discussion

The mitochondrial 3243 A to G mutation is among the most common causes of neurogenetic disorders, with an estimated prevalence of about 1 in 15,000 individuals (Majamaa et al., 1998). Historically some of the clinical manifestations have been ascribed with this mutation which includes mitochondrial encephalopathy with lactic acidosis and stroke-like episodes (MELAS), maternally inherited deafness and diabetes (MIDD) and chronic progressive external ophthalmoplegia (CPEO) (Nesbitt et al., 2013). The mutation was first identified by Goto and colleagues in 1990 and was identified as MELAS mutation (Goto et al., 1990). With the progression of time diseases associated with the mutation have increased. Many mitochondrial 3243 A to G mutation carriers, however, do not exhibit the whole MELAS syndrome. Rather than developing the strokelike episodes typical of MELAS, they may either be asymptomatic or exhibit a wide range of clinical symptoms which indicate multiorgan engagement but differ in their clinical severity from mild to severe involvement (Damian et al., 1995). The A3243G mutation appears to be more common than previously believed, and its clinical symptoms are likely underestimated(Manwaring et al., 2007).

The objective of this study was to better understand the clinical manifestations connected to this mutation across several phases of life and analyze the significance of heteroplasmy, or the percentage of mutant mitochondrial DNA (mtDNA), in defining these manifestations. The results illustrate the complexity of mitochondrial disease by emphasizing how age and heteroplasmy levels affect the clinical manifestations. The study showed that the 3243 A to G mutation is linked to a wide range of clinical characteristics that differ markedly during different phases of life.

Generally, common childhood symptoms include MELAS and MERRF. MELAS is generally diagnosed below age 20. The disease is diagnosed when heteroplasmy is generally 50%-90%. Also, MERRF is diagnosed at an early stage of life. With the progression of age MIDD is generally diagnosed with a heteroplasmy level of 30%-70%. Also, CPEO, Peripheral vestibular dysfunction and hypertrophic cardiomyopathy is generally diagnosed at the middle stage of life which is under 40. Lately, retinal pigmentary abnormalities, muscle stiffness and end stage renal diseases are diagnosed at the end stage of life which is basically after 40 years. These phenotypes show different levels of heteroplasmy. It is to mention that, age of onset of these diseases may vary as case of exception and heteroplasmy level is not tissue specific.

Our findings align with the majority of existing research on the 3243 A to G mutation, particularly regarding the wide range of clinical symptoms and the significant role heteroplasmy plays in determining disease severity. Additionally, the mutation has been linked in the past to a variety of illnesses, including maternally inherited diabetes and deafness (MIDD), MELAS syndrome, and others. Our research supports the idea that heteroplasmy is essential for this variability.

Research by Geng and colleagues demonstrates that the m.3243A>G mutation in mitochondrial DNA (mtDNA) impairs mitochondrial function, especially in peripheral blood mononuclear cells (PBMCs). Increased oxidative stress and decreased energy generation are associated with this disorder. More prominent clinical symptoms and more obvious functional impairment are correlated with higher heteroplasmy levels (Geng et al., 2019). It is seen that a heteroplasmy of 90% - 100% causes Leigh syndrome. This syndrome is a devastating neurodegenerative disease which is generally diagnosed at an early stage of life (Baertling et al., 2014). This sets an example of the consistency of our findings with previous studies.

It is possible to partially explain the variation in clinical symptoms linked to the 3243 A to G mutation by considering the molecular mechanisms behind mitochondrial malfunction, especially when considering different life stages. Due to the mutation's impact on mitochondrial tRNA, there will be a reduction in the production of mitochondrial proteins and consequent energy shortages. The brain, muscles, and pancreas are high-energy tissues that are disproportionately affected by this energy imbalance, which accounts for the early onset of symptoms in these organs (Goto et al., 1990). This image is further complicated by heteroplasmy. The concept of heteroplasmy suggests that cells contain a mix of normal and mutated mtDNA, with varying proportions of mutated mtDNA across different tissues. There is a good chance that this tissue-specific heterogeneity adds to the variety of clinical manifestations seen. For instance, tissues like neurons and muscle cells, which require more energy, might be more susceptible to the negative consequences of high heteroplasmy levels, which could result in severe symptoms like muscle weakness and stroke-like episodes. Conversely, organs that require less energy could be able to handle higher concentrations of mutant mtDNA, which could account for some patients' milder symptoms even though they have significant heteroplasmy (Wallace, 1999). In the study, these facts appeared as a generalized form. Heteroplasmy level is presented without specification of tissues so that a standard level can be formed. For these phenotypes, different levels of heteroplasmy are observed in different tissues. But, heteroplasmy in leukocyte generally shows a standard expression (Tranah et al., 2018).

The diagnosis and treatment of patients with the mitochondrial 3243 A to G mutation will have a notable benefit by the study's findings. Firstly, findings emphasize the importance of prompt and precise diagnosis, especially for patients with high heteroplasmy levels who can be susceptible to

severe phenotypes. When combined with heteroplasmy quantification, genetic testing for the mitochondrial 3243 A to G mutation can yield important prognostic data and direct therapeutic care.

From the study, the best course of treatment may involve a customized strategy that considers the patient's stage of life as well as their amount of heteroplasmy. For example, young children with high heteroplasmy levels require early management to avoid or lessen severe consequences like MELAS syndrome. Contrarily, patients at middle or end stage with medium heteroplasmy level may take treatment approaches for treating diseases like MIDD, cardiomyopathy and nephropathy(Parikh et al., 2015).

The result of the study enlists the identified clinical phenotypes which appear at different stages of life and their correlation to heteroplasmy level. Age of onset and age of diagnosis are not always the same. Because, disease is not diagnosed clinically just after the onset often. However, the idea of classification of age in the study may help to enforce the initiation of the diagnosis process. Also, the idea of heteroplasmy may predict the phenotype which may appear in the individual. To sum up, determination of heteroplasmy level at a certain age may predict the appearance of phenotype and can be beneficial in the treatment.

Chapter 5

Limitation of the study

Though the study provides important insights on the mutation and its phenotypes, there are some limitations which should be acknowledged. First, most of the the selected study reports lack control population number. Second, three studies had only 1 individual as tested population. Still, those studies were selected to ensure the listing of phenotypes. Third, the classification based on age can not be considered as standard. Because, those information were collected from only a few papers. Additionally, there are not so many studies available based on age. Fourth, heteroplasmy levels mentioned are not tissue specific. Because, till date there has no study taken place which ensures the constant level of heteroplasmy in certain tissues. In different tissues, different levels of heteroplasmy were seen. Moreover, they don't have any direct relation with each other. So, this study can't claim the mentioned heteroplasmy levels as standards.

Chapter 6

Conclusion

This study concludes by illustrating the complex relationship between heteroplasmy, clinical phenotypes and the mitochondrial 3243 A to G mutation. These findings are collected from different studies conducted on individuals harboring the mutation. There were limitations in those studies as well as in this study as a consequence. To establish a standard classification of age and heteroplasmy level more research is required. The results highlight the significance of taking the patient's life stage and heteroplasmy levels into account when diagnosing and treating mitochondrial disorders. Understanding of the variation in heteroplasmy levels among tissues and how these variations affect clinical variability may help the progression of targeted therapy. Finding nuclear genes that interact with the mitochondrial mutation may help explain the disease's pathophysiology and identify new targets for treatment. Even though there is still a lot to learn about the variables that affect clinical variability, this work lays the groundwork for further investigations targeted at enhancing the treatment and prognosis of individuals with mitochondrial illnesses.

References

1.Anan, R., Nakagawa, M., Miyata, M., Higuchi, I., Nakao, S., Suehara, M., Osame, M., & Tanaka,
H. (1995). Cardiac Involvement in Mitochondrial Diseases: A Study on 17 Patients With
Documented Mitochondrial DNA Defects. *Circulation*, 91(4), 955–961.
https://doi.org/10.1161/01.CIR.91.4.955

2.Baertling, F., Rodenburg, R. J., Schaper, J., Smeitink, J. A., Koopman, W. J. H., Mayatepek, E., Morava, E., & Distelmaier, F. (2014). A guide to diagnosis and treatment of Leigh syndrome. *Journal of Neurology, Neurosurgery, and Psychiatry*, 85(3), 257–265. https://doi.org/10.1136/jnnp-2012-304426

3.Cai, M., Yu, Q., & Bao, J. (2022). A case report of mitochondrial myopathy with membranous nephropathy. *BMC Nephrology*, *23*(1), 87. https://doi.org/10.1186/s12882-022-02710-0

4.Chomyn, A., Enriquez, J. A., Micol, V., Fernandez-Silva, P., & Attardi, G. (2000). The mitochondrial myopathy, encephalopathy, lactic acidosis, and stroke-like episode syndrome-associated human mitochondrial tRNALeu(UUR) mutation causes aminoacylation deficiency and concomitant reduced association of mRNA with ribosomes. *The Journal of Biological Chemistry*, *275*(25), 19198–19209. https://doi.org/10.1074/jbc.M908734199

5.Chung, C.-Y., Singh, K., Kotiadis, V. N., Valdebenito, G. E., Ahn, J. H., Topley, E., Tan, J., Andrews, W. D., Bilanges, B., Pitceathly, R. D. S., Szabadkai, G., Yuneva, M., & Duchen, M. R. (2021). Constitutive activation of the PI3K-Akt-mTORC1 pathway sustains the m.3243 A > G mtDNA mutation. *Nature Communications*, *12*(1), 6409. https://doi.org/10.1038/s41467-021-26746-2

6.Cox, B. C., Pearson, J. Y., Mandrekar, J., & Gavrilova, R. H. (2023). The clinical spectrum of MELAS and associated disorders across ages: A retrospective cohort study. *Frontiers in Neurology*, *14*, 1298569. https://doi.org/10.3389/fneur.2023.1298569

7.Damian, M. S., Seibel, P., Reichmann, H., Schachenmayr, W., Laube, H., Bachmann, G., Wassill, K. H., & Dorndorf, W. (1995). Clinical spectrum of the MELAS mutation in a large pedigree. *Acta Neurologica Scandinavica*, *92*(5), 409–415. https://doi.org/10.1111/j.1600-0404.1995.tb00156.x

8.de Laat, P., Rodenburg, R. R., Roeleveld, N., Koene, S., Smeitink, J. A., & Janssen, M. C. (2021).
Six-year prospective follow-up study in 151 carriers of the mitochondrial DNA 3243 A>G variant. *Journal of Medical Genetics*, 58(1), 48–55. https://doi.org/10.1136/jmedgenet-2019-106800

9.Deschauer, M., Wieser, T., Neudecker, S., Lindner, A., & Zierz, S. (1999). Mitochondrial 3243 A→G mutation (MELAS mutation) associated with painful muscle stiffness. *Neuromuscular Disorders*, 9(5), 305–307. https://doi.org/10.1016/S0960-8966(99)00019-X

10.Elson, J. L., Samuels, D. C., Turnbull, D. M., & Chinnery, P. F. (2001). Random intracellular drift explains the clonal expansion of mitochondrial DNA mutations with age. *American Journal of Human Genetics*, *68*(3), 802–806. https://doi.org/10.1086/318801

11.Fabrizi, G. M., Cardaioli, E., Grieco, G. S., Cavallaro, T., Malandrini, A., Manneschi, L., Dotti, M. T., Federico, A., & Guazzi, G. (1996). The A to G transition at nt 3243 of the mitochondrial tRNALeu(UUR) may cause an MERRF syndrome. *Journal of Neurology, Neurosurgery & Psychiatry*, *61*(1), 47–51. https://doi.org/10.1136/jnnp.61.1.47

12.Gallego-Delgado, M., Cobo-Marcos, M., Bornstein, B., Hernández-Laín, A., Alonso-Pulpón,L., & Garcia-Pavia, P. (2015). Mitochondrial cardiomyopathies associated with the m.3243A>G

mutation in the MT-TL1 gene: Two sides of the same coin. *Revista Espanola De Cardiologia* (*English Ed.*), 68(2), 153–155. https://doi.org/10.1016/j.rec.2014.09.007

13.Geng, X., Zhang, Y., Yan, J., Chu, C., Gao, F., Jiang, Z., Zhang, X., Chen, Y., Wei, X., Feng, Y., Lu, H., Wang, C., Zeng, F., & Jia, W. (2019). Mitochondrial DNA mutation m.3243A>G is associated with altered mitochondrial function in peripheral blood mononuclear cells, with heteroplasmy levels and with clinical phenotypes. *Diabetic Medicine*, *36*(6), 776–783. https://doi.org/10.1111/dme.13874

14.Gorman, G. S., Chinnery, P. F., DiMauro, S., Hirano, M., Koga, Y., McFarland, R., Suomalainen, A., Thorburn, D. R., Zeviani, M., & Turnbull, D. M. (2016). Mitochondrial diseases. *Nature Reviews. Disease Primers*, *2*, 16080. https://doi.org/10.1038/nrdp.2016.80

15.Gorman, G. S., Schaefer, A. M., Ng, Y., Gomez, N., Blakely, E. L., Alston, C. L., Feeney, C., Horvath, R., Yu-Wai-Man, P., Chinnery, P. F., Taylor, R. W., Turnbull, D. M., & McFarland, R. (2015). Prevalence of nuclear and mitochondrial DNA mutations related to adult mitochondrial disease. *Annals of Neurology*, *77*(5), 753–759. https://doi.org/10.1002/ana.24362

16.Greaves, L. C., Yu-Wai-Man, P., Blakely, E. L., Krishnan, K. J., Beadle, N. E., Kerin, J., Barron, M. J., Griffiths, P. G., Dickinson, A. J., Turnbull, D. M., & Taylor, R. W. (2010).
Mitochondrial DNA Defects and Selective Extraocular Muscle Involvement in CPEO. *Investigative Opthalmology & Visual Science*, *51*(7), 3340. https://doi.org/10.1167/iovs.09-4659

17.Hammans, S. R., Sweeney, M. G., Hanna, M. G., Brockington, M., Morgan-Hughes, J. A., & Harding, A. E. (n.d.). *The mitochondria! DNA transfer RNALeu*<*UUR*).

18.Hsu, P.-C., Chu, C.-S., Lin, T.-H., Lu, Y.-H., Lee, C.-S., Lai, W.-T., & Sheu, S.-H. (2008). Adult-onset hypertrophic cardiomyopathy manifested as initial major presentation of mitochondrial disease with A-to-G 3243 tRNA Leu(UUR) point mutation. *International Journal of Cardiology*, *129*(3), 441–443. https://doi.org/10.1016/j.ijcard.2007.06.098

19.Inoue, A., Iwasaki, S., Fujimoto, C., Kinoshita, M., & Yamasoba, T. (2019). Progression of Peripheral Vestibular Dysfunctions in Patients With a Mitochondrial A3243G Mutation. *Otology* & *Neurotology*, 40(3), 359–364. https://doi.org/10.1097/MAO.000000000002091

20.Iwasaki, N., Babazono, T., Tsuchiya, K., Tomonaga, O., Suzuki, A., Togashi, M., Ujihara, N., Sakka, Y., Yokokawa, H., Ogata, M., Nihei, H., & Iwamoto, Y. (2001). Prevalence of A-to-G mutation at nucleotide 3243 of the mitochondrial tRNALeu(UUR) gene in Japanese patients with diabetes mellitus and end stage renal disease. *Journal of Human Genetics*, *46*(6), 330–334. https://doi.org/10.1007/s100380170068

21.Jiang, J., Wang, X. L., & Ma, Y. Y. (2015). Respiratory chain complex III deficiency in patients with tRNA-leu mutation. *Genetics and Molecular Research*, *14*(4), 18629–18636. https://doi.org/10.4238/2015.December.28.12

22.Jin, Y., Nakamura, M., Shinjo, Y., & Kaga, K. (2006). Vestibular-evoked myogenic potentials in cochlear implant children. *Acta Oto-Laryngologica*, *126*(2), 164–169. https://doi.org/10.1080/00016480500312562

23.Klein, I., Verhaak, C. M., Smeitink, J. A. M., De Laat, P., Janssen, M. C. H., & Custers, J. A. E. (2022). Identifying trajectories of fatigue in patients with primary mitochondrial disease due to the m.3243A > G variant. *Journal of Inherited Metabolic Disease*, 45(6), 1130–1142. https://doi.org/10.1002/jimd.12546

24.Klemm, T., Neumann, S., Trülzsch, B., Pistrosch, F., Hanefeld, M., & Paschke, R. (2001). Search for mitochondrial DNA mutation at position 3243 in German patients with a positive family history of maternal diabetes mellitus. *Experimental and Clinical Endocrinology & Diabetes*, 109(05), 283–287. https://doi.org/10.1055/s-2001-16348

25.Kyriakidou, A., Hadjivassiliou, M., Papapostolou, A., & Picolos, M. K. (2023). Maternally Inherited Diabetes and Deafness (MIDD)—Atypical Clinical Diabetes Features Leading to the Diagnosis. *JCEM Case Reports*, 1(3), luad047. https://doi.org/10.1210/jcemcr/luad047

26.Latkany, P., Ciulla, T. A., Cucchillo, P., & Malkoff, M. D. (1999). Mitochondrial maculopathy: Geographic atrophy of the macula in the MELAS associated A to G 3243 mitochondrial DNA point mutation. *American Journal of Ophthalmology*, *128*(1), 112–114. https://doi.org/10.1016/S0002-9394(99)00057-4

27.Li, D., Liang, C., Zhang, T., Marley, J. L., Zou, W., Lian, M., & Ji, D. (2022a). Pathogenic mitochondrial DNA 3243A>G mutation: From genetics to phenotype. *Frontiers in Genetics*, *13*, 951185. https://doi.org/10.3389/fgene.2022.951185

28.Li, D., Liang, C., Zhang, T., Marley, J. L., Zou, W., Lian, M., & Ji, D. (2022b). Pathogenic mitochondrial DNA 3243A>G mutation: From genetics to phenotype. *Frontiers in Genetics*, *13*, 951185. https://doi.org/10.3389/fgene.2022.951185

29.Ma, Y.-Y., Wu, T.-F., Liu, Y.-P., Wang, Q., Li, X.-Y., Song, J.-Q., Shi, X.-Y., Zhang, W.-N., Zhao, M., Hu, L.-Y., Yang, Y.-L., & Zou, L.-P. (2013). Heterogeneity of six children and their mothers with mitochondrial DNA 3243 A>G mutation. *Mitochondrial DNA*, *24*(3), 297–302. https://doi.org/10.3109/19401736.2012.760071

30.Maassen, J. A. (2002). Mitochondrial diabetes: Pathophysiology, clinical presentation, and genetic analysis. *American Journal of Medical Genetics*, *115*(1), 66–70. https://doi.org/10.1002/ajmg.10346

31.Majamaa, K., Moilanen, J. S., Uimonen, S., Remes, A. M., Salmela, P. I., Kärppä, M., Majamaa-Voltti, K. A. M., Rusanen, H., Sorri, M., Peuhkurinen, K. J., & Hassinen, I. E. (1998). Epidemiology of A3243G, the Mutation for Mitochondrial Encephalomyopathy, Lactic Acidosis, and Strokelike Episodes: Prevalence of the Mutation in an Adult Population. *The American Journal of Human Genetics*, *63*(2), 447–454. https://doi.org/10.1086/301959

32.Manwaring, N., Jones, M. M., Wang, J. J., Rochtchina, E., Howard, C., Mitchell, P., & Sue, C.
M. (2007). Population prevalence of the MELAS A3243G mutation. *Mitochondrion*, 7(3), 230–233. https://doi.org/10.1016/j.mito.2006.12.004

33.Moraes, C. T., Ciacci, F., Silvestri, G., Shanske, S., Sciacco, M., Hirano, M., Schon, E. A., Bonilla, E., & DiMauro, S. (1993). Atypical clinical presentations associated with the MELAS mutation at position 3243 of human mitochondrial DNA. *Neuromuscular Disorders*, *3*(1), 43–50. https://doi.org/10.1016/0960-8966(93)90040-Q

34.Motlagh Scholle, L., Zierz, S., Mawrin, C., Wickenhauser, C., & Lehmann Urban, D. (2020). Heteroplasmy and Copy Number in the Common m.3243A>G Mutation—A Post-Mortem Genotype–Phenotype Analysis. *Genes*, *11*(2), 212. https://doi.org/10.3390/genes11020212

35.Nesbitt, V., Pitceathly, R. D. S., Turnbull, D. M., Taylor, R. W., Sweeney, M. G., Mudanohwo, E. E., Rahman, S., Hanna, M. G., & McFarland, R. (2013). The UK MRC Mitochondrial Disease Patient Cohort Study: Clinical phenotypes associated with the m.3243A>G mutation--implications for diagnosis and management. *Journal of Neurology, Neurosurgery, and Psychiatry*, 84(8), 936–938. https://doi.org/10.1136/jnnp-2012-303528

36.Nicholls, T. J., & Gustafsson, C. M. (2018). Separating and Segregating the Human Mitochondrial Genome. *Trends in Biochemical Sciences*, *43*(11), 869–881. https://doi.org/10.1016/j.tibs.2018.08.007

37.Parakatselaki, M.-E., & Ladoukakis, E. D. (2021). mtDNA Heteroplasmy: Origin, Detection,
Significance, and Evolutionary Consequences. *Life*, *11*(7), 633.
https://doi.org/10.3390/life11070633

38.Parikh, S., Goldstein, A., Koenig, M. K., Scaglia, F., Enns, G. M., Saneto, R., Anselm, I., Cohen, B. H., Falk, M. J., Greene, C., Gropman, A. L., Haas, R., Hirano, M., Morgan, P., Sims, K., Tarnopolsky, M., Van 39.Hove, J. L. K., Wolfe, L., & DiMauro, S. (2015). Diagnosis and management of mitochondrial disease: A consensus statement from the Mitochondrial Medicine Society. *Genetics in Medicine: Official Journal of the American College of Medical Genetics*, *17*(9), 689–701. https://doi.org/10.1038/gim.2014.177

40.Patil, A. D., Biousse, V., & Newman, N. J. (2022). Ischemic Optic Neuropathies: Current Concepts. *Annals of Indian Academy of Neurology*, 25(Suppl 2), S54–S58. https://doi.org/10.4103/aian.aian_533_22

41.Pia, S., & Lui, F. (2024). Melas Syndrome. In *StatPearls*. StatPearls Publishing. http://www.ncbi.nlm.nih.gov/books/NBK532959/

42.Picard, M., Zhang, J., Hancock, S., Derbeneva, O., Golhar, R., Golik, P., O'Hearn, S., Levy, S., Potluri, P., Lvova, M., Davila, A., Lin, C. S., Perin, J. C., Rappaport, E. F., Hakonarson, H., Trounce, I. A., Procaccio, V., & Wallace, D. C. (2014). Progressive increase in mtDNA 3243A>G heteroplasmy causes abrupt transcriptional reprogramming. *Proceedings of the National Academy*

of Sciences of the United States of America, 111(38), E4033-4042. https://doi.org/10.1073/pnas.1414028111

43.Pons, R., Andreu, A. L., Checcarelli, N., Vilà, M. R., Engelstad, K., Sue, C. M., Shungu, D., Haggerty, R., de Vivo, D. C., & DiMauro, S. (2004). Mitochondrial DNA abnormalities and autistic spectrum disorders. *The Journal of Pediatrics*, *144*(1), 81–85. https://doi.org/10.1016/j.jpeds.2003.10.023

44.Rustin, P., Von Kleist-Retzow, J.-C., Vajo, Z., Rotig, A., & Munnich, A. (2000). For debate: Defective mitochondria, free radicals, cell death, aging-reality or myth-ochondria? *Mechanisms of Ageing and Development*, *114*(3), 201–206. https://doi.org/10.1016/S0047-6374(00)00102-0

45.Ryytty, S., & Hämäläinen, R. H. (2023). The Mitochondrial m.3243A>G Mutation on the Dish, Lessons from In Vitro Models. *International Journal of Molecular Sciences*, *24*(17), 13478. https://doi.org/10.3390/ijms241713478

46.Sakata, A., Kashio, A., Koyama, H., Uranaka, T., Iwasaki, S., Fujimoto, C., Kinoshita, M., & Yamasoba, T. (2022). Long-Term Progression and Rapid Decline in Hearing Loss in Patients with a Point Mutation at Nucleotide 3243 of the Mitochondrial DNA. *Life*, *12*(4), 543. https://doi.org/10.3390/life12040543

47.Sbisà, E., Tanzariello, F., Reyes, A., Pesole, G., & Saccone, C. (1997). Mammalian mitochondrial D-loop region structural analysis: Identification of new conserved sequences and their functional and evolutionary implications. *Gene*, 205(1–2), 125–140. https://doi.org/10.1016/s0378-1119(97)00404-6

48.Scarcella, S., Dell'Arti, L., Gagliardi, D., Magri, F., Govoni, A., Velardo, D., Mainetti, C., Minorini, V., Ronchi, D., Piga, D., Comi, G. P., Corti, S., & Meneri, M. (2023). Ischemic optic

neuropathy as first presentation in patient with m.3243 A > G MELAS classic mutation. *BMC Neurology*, 23(1), 165. https://doi.org/10.1186/s12883-023-03198-3

49.Smith, P. R., Bain, S. C., Good, P. A., Hattersley, A. T., Barnett, A. H., Gibson, J. M., & Dodson, P. M. (1999). *Pigmentary Retinal Dystrophy and the Syndrome of Maternally Inherited Diabetes and Deafness Caused by the Mitochondrial DNA 3243 tRNALeu A to G Mutation. 106*(6).

50.Sue, C. M., Mitchell, P., Crimmins, D. S., Moshegov, C., Byme, E., & Morris, J. G. L. (n.d.). *Pigmentary retinopathy associated with the mitpoociahnotnmdruitaa1t..iDoNnA 3243*.

51.Suzuki, S., Oka, Y., Kadowaki, T., Kanatsuka, A., Kuzuya, T., Kobayashi, M., Sanke, T., Seino, Y., & Nanjo, K. (2003). Clinical features of diabetes mellitus with the mitochondrial DNA 3243 (AÁ/G) mutation in Japanese: Maternal inheritance and mitochondria-related complications. *Diabetes Research and Clinical Practice*.

52.Tranah, G. J., Katzman, S. M., Lauterjung, K., Yaffe, K., Manini, T. M., Kritchevsky, S., Newman, A. B., Harris, T. B., & Cummings, S. R. (2018). Mitochondrial DNA m.3243A > G heteroplasmy affects multiple aging phenotypes and risk of mortality. *Scientific Reports*, 8(1), 11887. https://doi.org/10.1038/s41598-018-30255-6

53.Tuppen, H. A. L., Blakely, E. L., Turnbull, D. M., & Taylor, R. W. (2010). Mitochondrial DNA mutations and human disease. *Biochimica et Biophysica Acta (BBA) - Bioenergetics*, *1797*(2), 113–128. https://doi.org/10.1016/j.bbabio.2009.09.005

54.van den Ouweland, J. M., Lemkes, H. H., Ruitenbeek, W., Sandkuijl, L. A., de Vijlder, M. F., Struyvenberg, P. A., van de Kamp, J. J., & Maassen, J. A. (1992). Mutation in mitochondrial tRNA(Leu)(UUR) gene in a large pedigree with maternally transmitted type II diabetes mellitus and deafness. *Nature Genetics*, *1*(5), 368–371. https://doi.org/10.1038/ng0892-368

55.Wallace, D. C. (1999). Mitochondrial diseases in man and mouse. *Science (New York, N.Y.)*, 283(5407), 1482–1488. https://doi.org/10.1126/science.283.5407.1482

56.Wang, S., Wu, S., Zheng, T., Yang, Z., Ma, X., Jia, W., & Xiang, K. (2013). Mitochondrial DNA mutations in diabetes mellitus patients in Chinese Han population. *Gene*, *531*(2), 472–475. https://doi.org/10.1016/j.gene.2013.09.019

57. Yamamoto, M. (2003). With Mitochondrial DNA 3243 (A-*G) Mutation. 12.

58.Yan, C., Duanmu, X., Zeng, L., Liu, B., & Song, Z. (2019). Mitochondrial DNA: Distribution, Mutations, and Elimination. *Cells*, 8(4), 379. https://doi.org/10.3390/cells8040379

59.Yokota, M., Hatakeyama, H., Okabe, S., Ono, Y., & Goto, Y. (2015). Mitochondrial respiratory dysfunction caused by a heteroplasmic mitochondrial DNA mutation blocks cellular reprogramming. *Human Molecular Genetics*, 24(16), 4698–4709. https://doi.org/10.1093/hmg/ddv201

60.Zhong, S. (2000). Presence of mitochondrial tRNA Leu(UUR) A to G 3243 mutation in DNA extracted from serum and plasma of patients with type 2 diabetes mellitus. *Journal of Clinical Pathology*, *53*(6), 466–469. https://doi.org/10.1136/jcp.53.6.466