

Hypovitaminosis D: A modifiable Risk Factor for Noncommunicable Diseases

By

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A thesis submitted to the Department of Mathematics and Natural Sciences in partial fulfillment of the requirements for the degree of Master of Science in Biotechnology

Department of Mathematics and Natural Sciences

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Declaration

It is hereby declared that

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Ethics Statement

This material is an original work, which has not been previously published elsewhere. It is my own research and analysis in a truthful and complete manner. The paper properly credits all the sources used (correct citation).

Abstract

Over the years, a broad spectrum of non-communicable diseases (NCDs), including osteoporosis, asthma, diabetes, cancer, hypertension, cardiovascular diseases, chronic kidney and liver diseases have been linked to poor vitamin D levels. This review presents the circumstances leading to hypovitaminosis D and how such a deficiency can eventually independently induce the development of NCDs. PubMed and EMBASE databases were used to identify all studies that assessed the association between vitamin D and various NCDs up until February 2022, without language restrictions. Available evidence from observational studies suggests a strong link between vitamin D and many NCDs. However, underlying molecular mechanisms of vitamin D mode of action that contribute to the pathophysiology of these non-communicable diseases are often complex and not fully understood. Vitamin D and its biologically active analogs have shown therapeutic potential for treatment or prevention of some NCDs. However, large-scale vitamin D supplementation trials have yet to confirm such a causal relationship. Further studies are required to clearly establish and understand the nature of NCDs association with vitamin D.

*This work is dedicated to
My Dear Family*

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Sincerely,

Afrin Hossain,

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List of Acronyms

7-DHC – 7-Dehydrocholesterol

CKD – Chronic kidney disease

COPD – Chronic obstructive pulmonary disease

CVD – Cardiovascular disease

ECF – Extracellular fluid

HDL – High density lipoprotein

IBS – Irritable bowel syndrome

LDL – Low density lipoprotein

MI – Myocardial infarction

MS – Multiple sclerosis

NAFLD – Non-alcoholic fatty liver disease

NCD – Non-communicable disease

NO – Nitric oxide

PTH – Parathyroid hormone

RANK – Receptor Activator of Nuclear Factor Kappa-B

RAS – Renin-angiotensin system

ROS – Reactive oxygen species

RXR – Retinoid X receptor

SPF – Sun protection factor

VDR – Vitamin D receptor

UVB – Ultra-violet B rays

WHO – World Health Organization

Introduction
And
Literature Review

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1. Introduction

For most of history, life expectancy was cut short by famine, injury, and infectious diseases. Economic development and improvement in living conditions conquered these conditions, completely annihilating many major infectious diseases like smallpox. Socioeconomic progress, however, has created an environment that favors the development of non-infectious diseases. The chronic non-infectious diseases are referred to as noncommunicable diseases (NCDs). NCDs are named as such because they cannot be communicated from person to person. These conditions have a long-term health consequence, last for a long period of time and create a need for long-term treatment and care. Every year NCDs account for almost 71% of all worldwide deaths which corresponds to 41 million early deaths (World Health Organization, 2021). Cardiovascular diseases embody most of these deaths, followed by cancers, respiratory diseases, and diabetes. Preventive measures for NCDs focus on reducing the risk factors associated with these diseases. WHO listed 4 modifiable behavioral and metabolic risk factors for NCDs:

Behavioral Risk Factors-

- Tobacco use
- Physical inactivity
- Alcohol abuse
- Unhealthy diets

Metabolic Risk Factors-

- Hypertension
- Hyperglycemia
- Hyperlipidemia
- Obesity

In the last few decades, the listed risk factor awareness propelled change of existing policies and development of new policies that address both behavioral and metabolic risk factors. As the society experiences unparalleled progress and dramatic changes in lifestyle, it is important to identify emerging risk factors contributing to the growing number of NCDs.

Over the years, poor vitamin D level has been recognized as a risk factor for multiple non-communicable diseases. This thesis manuscript will review the circumstances leading to hypovitaminosis D and how such a deficiency can eventually independently induce the development of non-communicable diseases.

2. Methods

PubMed and EMBASE databases were used to identify all studies that assessed the association between vitamin D and various non-communicable diseases up until February 2022, without language restrictions. We included studies that compared vitamin D levels between NCD cases and controls and also those that used vitamin D supplementation to improve NCD status. Reference lists of relevant original papers and review articles were also examined.

3. Vitamin D

Vitamin D was first identified by McCollum *et al* in 1922 as the antirachitic element in cod liver oil. Being the fourth discovered vitamin, it was naturally assigned the fourth alphabet 'D'. Several forms of Vitamin D have been identified after its initial discovery. In 1930s two forms of vitamin D found in food and diet- Ergocalciferol and Cholecalciferol were identified. Structure of biologically active form of Vitamin D- calcifediol and calcitriol were elucidated respectively in the late 1960s and early 1970s (Holick, 1994).

List of several forms of vitamin D (vitamers) and their site of synthesis:

Generic Name	Other Names	Site of synthesis
Ergocalciferol	Vitamin D ₂	Photochemically synthesized in plants
Cholecalciferol	Vitamin D ₃	Synthesized in the skin of animals and humans
Calcifediol/ Calcidiol	25(OH)D	Liver
	25(OH)D ₃	
	25-hydroxycholecalciferol	
	25-hydroxyvitamin D ₃	
Calcitriol	1,25(OH) ₂ D	Kidney (Primary)
	1,25(OH) ₂ D ₃	
	1,25-dihydroxycholecalciferol	Lymphocytes, microglia, keratinocytes, etc
	1,25-dihydroxyvitamin D	
	Vitamin D Receptor Activator	

Table 3.1: Vitamers of vitamin D and their site of synthesis

All forms of vitamin D act in concert to ensure optimal health.

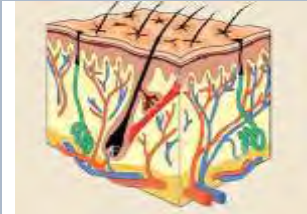



					
Cholecalciferol		Calcifediol		Calcitriol	
Cholecalciferol made in the skin is used by skin cells, which cannot rely on vitamin D from the blood	Taken directly into cells for local conversion and use	Form measured in the blood as indicator of vitamin D status	Hormonal vitamin D regulated by PTH for blood calcium control		
Decrease DNA damage	Most potent form of endothelial stability	Taken directly into cells and converted to calcitriol for local use	Increases calcium absorption in gut		
Decrease Cell death	Anti-inflammatory	Used as signaling molecule in most cells and tissues	Increases bone turnover		
Increase DNA repair	Form needed by breastfeeding mothers to transfer to baby through breastmilk		Increases calcium in the blood		

Table 3.2: Different forms of vitamin D and their functions

3.1 Biosynthesis of Vitamin D

For most of history sunlight exposure has been enough to cover our requirement of Vitamin D. Now-a-days vitamin D can be obtained through particular diet, fortified food and supplements, yet photosynthesis of Vitamin D under sunlight remains the primary source for most of the population.

7-dehydrocholesterol (7-DHC) is the prime precursor of the active form of vitamin D, calcitriol. Upon exposure to sunlight, high-energy UV photons (290-315nm) penetrate skin epidermis and cause photolysis of provitamin D₃ (7-dehydrocholesterol) to previtamin D₃. Within 2-3 days vitamin D₃ (calciferol or cholecalciferol) is spontaneously formed through a thermally induced isomerization of previtamin D₃ (Holick, 1987).

Cholecalciferol travels to the liver, and there, through the action of a hydroxylase enzyme it is converted to calcidiol. 1 α -hydroxylase enzyme in the kidney subsequently converts calcidiol to calcitriol, the biologically active vitamin D. From the kidney calcitriol enters the circulation.

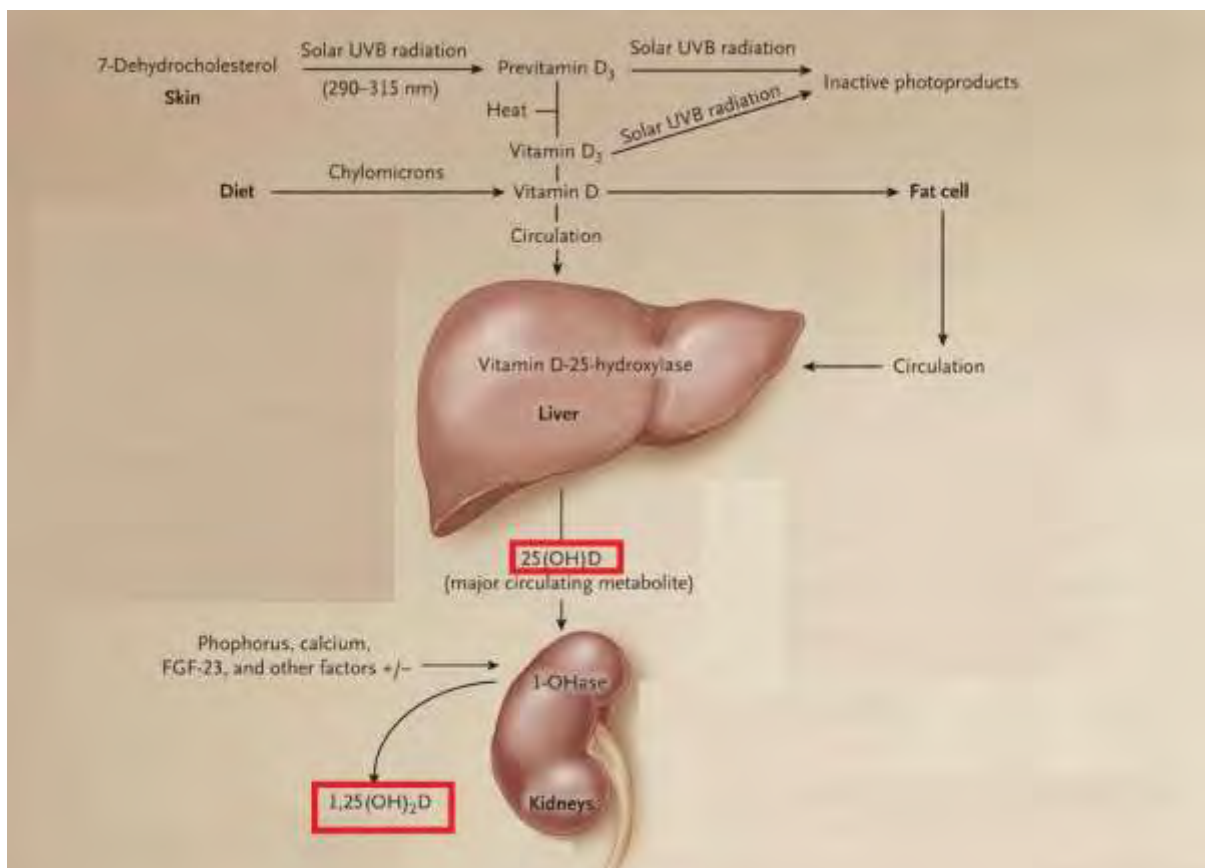


Figure 3.1: Stages of vitamin D biosynthesis (Holick, 2007)

Even if the skin is constantly exposed to sunlight vitamin D is not produced in excess. After initial exposure to UV radiation, provitamin D₃ is efficiently converted to previtamin D₃. However, previtamin D₃ is photosensitive, and sustained exposure to sunlight leads to isomerization into lumisterol. Consequently, only 10-20% of the initial provitamin D₃ concentrations eventually end up as previtamin D₃ (Holick, 1987).

In recent years several other tissues other than the kidneys were found to possess the ability to convert calcidiol to calcitriol. Extrarenal sites of calcitriol synthesis include lymphocytes, microglia of the brain (Eyles et al., 2005), keratinocytes (Lehmann & Meurer, 2003) and respiratory epithelial cells (Hansdottir et al., 2008). In vitro studies found many other nonrenal cells, such as bone, placenta, prostata, macrophages, and various cancer cells (e.g., from lung, prostata and skin) able to convert calcidiol to calcitriol (Lehmann & Meurer, 2003).

3.2 Vitamin D from natural diet, fortified food and supplements

Vitamin D can be sourced from diet in a very small amount (30%) as only limited number of foods contain it naturally (Holick, 2004). Egg yolks oily fish, fish liver oils, seal blubber, polar bear liver and whale blubber (Bills, 1927) (Kenny et al., 2004) are good sources of vitamin D. In many countries, food items like milk, margarine, and cereal are fortified with vitamin D. Industrial revolution across all continents has reduced our exposure to sunlight, thereby increasing our dependence on dietary sources of vitamin D. Vitamin D obtained from diet are not biologically active and must undergo two hydroxylation steps to become biologically active. Vitamin D₂ and vitamin D₃ absorbed from the intestine are first hydroxylated by 25-hydroxylase in the liver (Feldman et al., 2011). 1 α -hydroxylase in the kidney performs the second hydroxylation step, producing the final active metabolite.

As calcidiol and calcitriol are not considered as nutrients, they are not used for food fortification or dietary supplementation. There is evidence of ergocalciferol being too unstable for storing, and compared to cholecalciferol it has higher susceptibility to breakdown when cooked and baked. Therefore, studies concluded that the vitamin D form that ought to be considered in the context of food fortification and supplementation is cholecalciferol (Vieth, 2020).

3.3 Vitamin D Receptor (VDR)

Calcitriol exerts its effects via genomic and non-genomic pathway. The non-genomic effects (rapid responses, RR) are initiated at caveolae of the plasma membrane, whereas genomic effects are generated by binding of calcitriol to vitamin D receptor (VDR). VDR is an intracellular nuclear receptor that belongs to the thyroid hormone and retinoic acid receptor subfamily of nuclear hormone receptors. VDR can form heterodimers with retinoid X receptor (RXR) isoforms and control gene expression of variety of genes in many tissues. In these tissues VDR regulates cellular activities such as proliferation, differentiation, metabolism, ion transport, and apoptosis (Haussler et al., 2010).

VDR distribution in organs and tissues are as follows:

Cardiovascular system	Cardiomyocytes Smooth muscle cells
Endocrine system	Thyroid C-cells Parathyroid glands Langerhans islets
Epidermis	Hair follicles Keratocytes
Gastrointestinal System	Stomach Esophagus Intestine Liver
Immune System	Thymus T and B lymphocytes Bone marrow
Renal system	Ascending portion of Henle loop Juxtaglomerular cells
Respiratory system	Alveolar epithelium
Osteo-muscular system	Osteoblasts Chondrocytes Striated muscle
Reproductive system	Testis Ovary Uterus
Central nervous system	Neurons

Table 3.3: VDR distribution in organs and tissues (adapted from Vuolo et al., 2012)

3.4 Quantitation of vitamin D

In the past, vitamin D deficiency was diagnosed through clinical symptoms such as bone pain and proximal myopathy in adjunction to serum calcium, phosphate, and alkaline phosphate level quantitation (LIPS et al., 2006). More recently, advanced molecular techniques allow to assay serum vitamin D level with high level of accuracy. Clinical quantitation of vitamin D status is accomplished by measuring calcidiol, the primary circulating form of vitamin D. Serum calcidiol level is a better indicator of vitamin D status than calcitriol, because it has a relatively slower rate of clearance, and a half-life of approximately 15 days (Uribarri, 2007). Whereas the active form of vitamin D, calcitriol has a much shorter half-life (5–15 h). Additionally, conditions such as secondary hyperparathyroidism increase calcitriol concentrations making calcitriol an unreliable marker of vitamin D status (Feldman et al., 2011)

There has been poor consensus regarding definition of normal vitamin D status. Traditionally, calcidiol levels above 18-20 ng/ml were considered adequate or normal. However, due to growing body of evidence, a revised and more efficient classification has been suggested (Gómez Alonso et al., 2003). The revised cut-off levels indicated that, for a very long time wrong statistical approach was used to define "normal serum vitamin D levels". Human life begun in an ambient abundant in sunlight, and to this day, population living in such environment have a mean calcidiol concentration around 115 nmol/L (Luxwolda et al., 2013).

Vitamin D status is classified in four ranges as described below-

Vitamin D status	25(OH)D level (ng/ml)	25(OH)D level (nmol/l)
Desirable	> 40 ng/ml	> 100 nmol/l
Hypovitaminosis D	20 - 40 ng/ml	50 - 100 nmol/l
Vitamin D insufficiency	10 - 20 ng/ml	25 - 50 nmol/l
Deficient	<10 ng/ml	<25 nmol/l

Table 3.4: Classification of vitamin D status

3.5 Causes of Vitamin D Deficiency

3.5.1 Sunblock

Sunscreens are intended to absorb solar UVB radiation. A sunscreen with a sun protection factor (SPF) of 30 confers a solar UVB radiation absorption ability of nearly 95–98%. Topical application of such sunblocker will, consequently, decrease the ability of the skin to produce vitamin D by the same amount (95–98%) (T. C. Chen et al., 2007). A case control study found that topical use of sunscreen with as little as SPF 8 can cause a significant drop in vitamin D level in blood (Matsuoka et al., 1987).

3.5.2 Skin pigment

Melanin is a natural sunscreen that humans developed due to constant exposure to sunlight (N. G. Jablonski & Chaplin, 2000). Melanin's absorption spectrum is 290-700 nm and therefore competes with provitamin D₃ for the UVB photons that cross the skin epidermis. This competitive action limits its photolysis 7-dehydrocholesterol (7-DHC) to previtamin D₃ (Holick, 1987). Multiple experiments provide evidence to melanin's inhibitory role in vitamin D synthesis. Compared to lighter skin, darker skin, which contains more melanin, requires much more time of sun exposure to reach healthy level of vitamin D production. For example, an experiment conducted in Boston, Massachusetts found significant differences in Caucasian and black skin previtamin D₃ level after exposure to noon sunlight in the month of June (T. C. Chen et al., 2007). The differences are illustrated below-

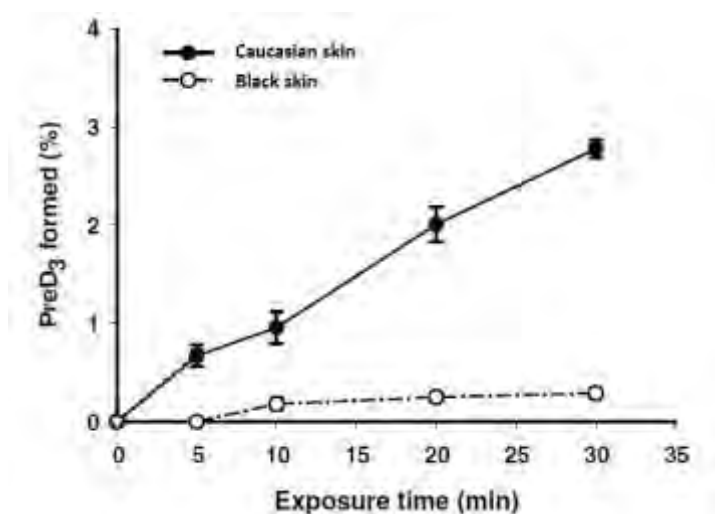


Figure 3.2: The conversion of 7-DHC to previtamin D₃ in Caucasian skin versus Black skin (T. C. Chen et al., 2007)

3.5.3 Clothing

Clothes can absorb most ultraviolet radiation. Studies found that any type of clothing covering the skin will block cutaneous synthesis of vitamin D (Matsuoka et al., 1992). A case control study found high prevalence of subclinical vitamin D deficiency among veiled Kuwaiti women (El-Sonbaty & Abdul-Ghaffar, 1996). Hypovitaminosis D was observed even among adolescent girls wearing concealing clothing in Turkey (Hatun et al., 2005).

3.5.4 Glass

Glass absorbs all UVB radiation. Therefore, skin exposed to sun UVB ray that passes through glass will not be able to produce any vitamin D. Plexiglass and plastic have similar UVB blocking effect on skin (Holick, 1994).

3.5.5 Aging

Aging decreases the capacity of human skin to synthesize vitamin D by greater than twofold. Data from clinical investigations indicate that there is an inverse relationship between age and concentrations of 7-dehydrocholesterol in human epidermis (MacLaughlin & Holick, 1985). A comparative study published in 1989 used UVB radiation in a tanning bed to compare the cutaneous synthesis of vitamin D₃ in healthy young people and older people. The results showed that the blood concentration of vitamin D in young subjects was at least three-fold higher than the older subjects (Holick et al., 1989).

3.5.6 Pollution

Air pollution has a significant and independent role in development of hypovitaminosis D. There is an inverse relationship between the level of air pollution and the amount of solar UVB radiation that can reach earth surface (Hosseinpahan et al., 2010). Therefore, less UVB can pass through more polluted areas, and as a consequence, less UVB is available for effective cutaneous vitamin D synthesis in the skin. Nitrous oxide, ozone and sulfur dioxide are some of the pollutants that absorb solar UVB radiation and decrease UVB radiation available for vitamin D synthesis (Spina Catherine Sheridan, 2005).

3.5.7 Altitude

Altitude has a dramatic impact on the extent of solar UVB that can make it to earth's surface. At high altitude the path length that UVB has to move through is shorter, thus more photons are available for skin to synthesize vitamin D (Wacker & Holick, 2013). An investigation comparing sun induced in vitro vitamin D₃ synthesis at different altitudes OF Mount Everest and Agra confirmed that higher altitude confers enhanced previtamin D synthesis. Mount Everest base camp and Agra are at same latitude but different altitude of 5300M and 169 M respectively. The results from the study demonstrated that there is an almost 5-fold increase in previtamin D₃ production in Mount Everest base camp compared to Agra (Wacker & Holick, 2013). Therefore, altitude is an independent factor that contributes to vitamin D deficiency.

3.5.8 Season and Latitude

The quality and quantity of solar radiation, especially UVB radiation, that reaches the earth's surface differs relative to latitude and season. Seasonal differences in vitamin D status of both children and adults is well documented (Holick, 2007). A study among 7437 Caucasian subjects revealed blood levels of calcidiol peaks in September (~30 ng/mL) and bottoms in February (~14 ng/mL) (Hyppönen & Power, 2007).

Latitude also has considerable influence on vitamin D synthesis (Webb et al., 1988). Mean circulating calcidiol in children and adults appears to be greater at equator (~40 ng/mL) and tends to drop in areas far North and South of the equator (~15 ng/mL) (Zittermann, 2006)

3.5.9 Cholecystectomy

Increased incidence of vitamin D deficiency has been reported in patients with prior cholecystectomy (Polat & Beyazal, 2018). A prior cholecystectomy confers a more than 2-fold heightened risk of deficiency (Stoker et al., 2012). This phenomenon may be due to reduced gut transit and augmented colonic passage after a cholecystectomy, resulting in suboptimal absorption (Fort et al., 1996). In addition, cholestyramine, a bile acid sequestrant often prescribed for post cholecystectomy diarrhea syndrome has been shown to impair vitamin D absorption (Thompson & Thompson, 1969).

4. Noncommunicable Diseases Linked to Hypovitaminosis D

4.1 Hypertension

The World Health Organization estimates 1.28 billion adults aged between 30-79 years to be suffering from hypertension globally. Essential hypertension, also defined as primary hypertension, is the most typical form of hypertension among hypertensive patients. Essential hypertension is often associated to low blood levels of ionized and ultrafiltrable calcium (McCarron et al., 1987) and elevated levels of parathyroid hormone (PTH) (Jorde et al., 2005). Both of these conditions, respectively known as hypocalcemia and hyperparathyroidism, are often secondary to vitamin D deficiency (Holick, 2005; Noto & Heller, 2009). Therefore, poor vitamin D levels may lead to the development of hypertension by creating conditions conducive to hypertension.

Activation of the renin–angiotensin system (RAS) links hypovitaminosis D to hypertension (Zittermann, 2006). The renin-angiotensin system (RAS) plays an essential role in the regulation of blood pressure. Inappropriate stimulation of the RAS has been linked to development of hypertension, stroke, and heart attack. Various studies suggest calcitriol may play a key role in preventing the unfavorable over-stimulation of the RAS. In a lot of epidemiological and clinical studies of the past, high plasma renin activity and/or high blood pressure has been observed in those with low calcitriol levels (Resnick et al., 1986; Rostand, 1997). Although the exact mechanism of how calcitriol affects the renin-angiotensin system was not known, results from interventional studies with calcitriol treatment had shown to reduce both plasma renin activity and blood pressure (Cheol Whee Park et al., 1999; Kimura et al., 1999). In 2003, a study found calcitriol to function as a potent repressor of renin gene expression (Li, 2003). Therefore, vitamin D contributes in maintaining appropriate blood pressure by regulating the expression of renin. This explains why deficiency of vitamin D leads to uncontrolled renin activity and ultimately high blood pressure.

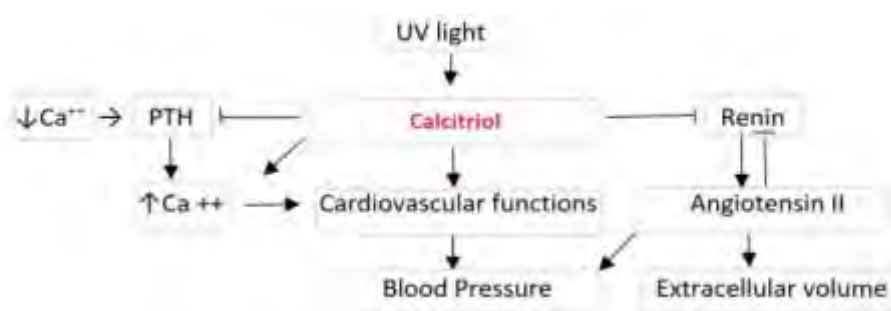


Figure 4.1: Outline of the interaction between the vitamin D and the renin–angiotensin system (RAS) leading to hypertension.

4.2 Cardiovascular Diseases

Vitamin D was first linked to cardiovascular disease in 1981, when Robert Scragg, hypothesized that the observed high CVD incidence in winter may be a consequence of low UV-B irradiation, with correspondingly low vitamin D status in that season (Scragg, 1981). After discovery of VDR in rat heart (Simpson R U et al., 1985), the relationship between vitamin D and CVD has become the focus of intense research. Numerous epidemiologic, observational, and clinical studies have found a strong relationship between serum calcidiol level and cardiovascular disease (CVD). Hypovitaminosis is consistently linked to cardiovascular events, such as myocardial infarction (MI), ST-elevation myocardial infarction (STEMI), non-ST-elevation myocardial infarction (NSTEMI), unstable angina, ischemic stroke, cardiovascular death, and higher mortality risk after acute stroke (Surdu et al., 2021).

Vitamin D deficiency plays multiple roles in development of atherosclerosis, the first step in CVD development. Low serum calcidiol has been found to be associated with more significant stenosis in the coronary arteries of postmenopausal women (Dziedzic et al., 2021). In observational and interventional studies, poor vitamin D levels was associated with unfavorable lipid profiles, and adequate vitamin D levels correlated with better lipid profiles (Challoumas, 2014). Unfavorable lipid profile (high total cholesterol, triglycerides, LDL and low HDL) is a known risk factors for CVD. Intervention studies found vitamin D repletion correlated with improved lipid profile (Manousopoulou et al., 2015).

A critical event in the development and progression of atherosclerosis is endothelial activation and rise in adhesion molecules, which increases with vitamin D deficiency and lowers with vitamin D supplementation (Salekzamani et al., 2017). Vitamin D has the ability to increase endothelial nitric oxide (NO) production (Surdu et al., 2021). Nitric oxide acts as a vasodilator and decreases the level of reactive oxygen species (ROS) which influences the vascular tone. On the other hand, hypovitaminosis D generates oxidative stress and triggers inflammation (Förstermann et al., 2017). The pathogenesis of atherosclerosis is attributed innate immune response cells (i.e., monocytes, macrophages) that cause chronic inflammation of the blood vessel walls (Wolf & Ley, 2019). Vitamin D mediates anti-inflammatory responses by reducing proinflammatory type 1 cytokines: IL-12, IL-6, IL-8, IFN-gamma, and TNF-alpha; and increasing anti-inflammatory type 2 cytokines: IL-4, IL-5, and IL-10 (Zhang et al., 2012). Vitamin D may also contribute in atherogenesis by inhibition action on proliferation of vascular smooth muscle cell (S. Chen et al., 2010).

Vitamin D action on atherosclerosis development is summarized in Table 4.1.

Lipid profile	<ul style="list-style-type: none"> • ↓ Total cholesterol • ↓ LDL-C • ↓ Triglycerides • ↑ HDL-C
Endothelial adhesion & activation	<ul style="list-style-type: none"> • ↓ Vascular cell adhesion molecule 1 (VCAM-1) • ↓ E-selectin
Vascular tone & endothelial function	<ul style="list-style-type: none"> • ↑ NO level • ↓ released ROS level
Inflammation & atherosclerosis	<ul style="list-style-type: none"> • ↓ Proinflammatory type 1 cytokines: IL-12, IL-6, IL-8, IFN-γ, TNF-α • ↑ Anti-inflammatory type 2 cytokines: IL-4, IL-5, & IL-10 • ↓ Oxidative stress by reducing cathepsin, IL-6 & adiponectin
Arterial smooth muscle cells	<ul style="list-style-type: none"> • ↓ Angiotensin II production • ↓ Oxidative stress • ↓ Tissue factor expression • Inhibits cellular senescence

Table 4.1: Vitamin D effects on atherosclerosis (adapted from Surdu et al., 2021).

A nested case-control study conducted over 10 years with 18,225 participants found low levels of calcidiol to be independently associated with higher risk of myocardial infarction in a graded manner, albeit other controlling for factors (Giovannucci et al., 2008). A recent experiment has shown that vitamin D is involved in lipid metabolism and diminishes the buildup of cholesterol by macrophages, making vitamin D in blood plasma a prognostic predictor of MI (Zhernakova et al., 2021). Evidence from decades of studies suggest vitamin D plays a multifactorial role in CVD development.

4.3 Cancer

The first association between cancer and sun exposure was reported in 1915 (Hoffman, 1915). The study compared cancer deaths between 1908 and 1912. The results showed that with growing distance from the equator, cancer death rate also increased. A multitude of epidemiologic studies followed these initial observations that led to the theory that vitamin D produced after sun exposure has anti-carcinogenic effects. A moderate, inverse correlation with solar radiation was observed for cancers of the organs in the digestive system including esophagus, stomach, pancreas, gallbladder, colon and rectum (Mizoue, 2004). Epidemiological data shows depleted levels of calcidiol are linked to an almost 50% increased risk of colon, prostate, or breast cancer (Garland et al., 2009; Holick, 2007). While adequate vitamin D status had a protective effect and lowered risk of cancer (Giovannucci, 2009). Epidemiological surveys have continually revealed that UVB exposure and subsequent increase in circulating levels of vitamin D are linked to decline in incidence and death in different histological types of cancer (Vuolo et al., 2012).

Although metabolizing enzymes of vitamin D are primarily located in liver and kidney, catabolic enzymes can be found in several tissues. Tissues, such as, lung, pancreas, colon, breast, and prostate can locally synthesize and degrade calcitriol and this phenomenon has been proposed to represent a key factor in numerous types of human cancer (Cross, 2007; Haussler et al., 1998; Schwartz et al., 2004). Multiple levels of evidence bolster the relationship between vitamin D and cancer (Vuolo et al., 2012):

- (1) Low circulating levels of calcifediol confer increased risk for developing cancer
- (2) High consumption of vitamin D is linked to a reduced risk of cancer
- (3) Cancer is less aggressive in summer when the synthesis of vitamin D is higher
- (4) Genetic polymorphisms of genes transcribing proteins involved in the vitamin D signal pathway influence the risk of developing cancer

In the last decades, advancement in molecular studies, allowed epidemiological data to be combined with cellular and molecular studies data to shed more light on the role of vitamin D in cancer. Calcitriol is now recognized as a powerful regulator of cell growth and differentiation. Accumulating evidence suggest it has also a role on cell death, tumor invasion, and angiogenesis. Murine models with prostate and lung metastasis, showed decreased cancer growth when treated with calcitriol (Getzenberg et al., 1997; Nakagawa, 2004).

Calcitriol is deemed to be analogous to a master switch controlling a great number of genes that modulate cell cycle progression, DNA replication and repair, etc. The antiproliferative effects of calcitriol have been repeatedly confirmed through many studies. The first instant was in 1981 where calcitriol was found to inhibit melanoma cell proliferation (Colston et al., 1981). The same year calcitriol was reported to be involved in bone marrow cell differentiation (E. Abe et al., 1981). Some studies suggest that vitamin D analogues have the ability to inhibit IGF-I signaling pathways, and therefore promote apoptosis in breast cancer cells (Xie et al., 1999).

Calcitriol shows substantial antitumoral activity *in vivo* such as human prostatic adenocarcinoma, human breast, pancreatic, and colon cancer as well as *in vitro* such as in leukemia, myeloma, and lymphoma lines (Reichel et al., 1989; Uhmman et al., 2011; Zhou et al., 1990). Calcitriol regulates transcription of several genes involved in various pathways that reduce tumorigenesis. Calcitriol’s role as a potent inhibitor of inflammation farther enhances its anti-tumoral activity (Vanoirbeek et al., 2011). Calcitriol inhibits the synthesis of prostaglandin, a significant stimulus for cell growth. Calcitriol is also able to induce prostaglandin degradation, and thus significantly reduce prostaglandin activity (Moreno et al., 2005).

Vitamin D Receptor (VDR) is expressed by both benign and malignant proliferative cells (Vuolo et al., 2012). Calcitriol binds to VDR and forms heterodimers with another receptor-ligand, and together they occupy specific nucleotide sequences known as vitamin D response elements (VDREs). Along some other transcription factors this complex transcribes vitamin D responsive genes. Some genes that are implicated in cancer are transcriptionally activated by calcitriol. For example, BGLAP (osteocalcin), the growth arrest and DNA-damage-inducible gene, GADD45 gene (Deeb et al., 2007).

Vitamin D action on cancer development is summarized in Table 4.2.

Promotes	Apoptosis
Inhibits	Angiogenesis
	Metastasis
	Tumorigenesis

Table 4.2: Vitamin D action on cancer development

A novel cancer etiology model proposes vitamin D and calcium as elements that can prevent and even potentially arrest cancer pathogenesis (Garland et al., 2009). The model is known as DINOMIT, abbreviation of disjunction, initiation, natural selection, overgrowth, metastasis, involution, and transition. According to the model vitamin D metabolites prevent disjunction of cells and have a positive impact in other phases of DINOMIT as well (Garland et al., 2007). The reported 10 mechanisms that account for the role of vitamin D and calcium in reducing cancer rate and death and the sources is summarized in Table 4.3:

Mechanism	Studies
1. Up-regulation of adherence & signaling between epithelial cells	(Johansen et al., 2000) (Ordóñez-Morán et al., 2008)
2. Contact inhibition of proliferation	(Johansen et al., 2000) (Ordóñez-Morán et al., 2008)
3. Differentiation	(Abe et al., 1986) (Ordóñez-Morán et al., 2008) (LAMPRECHT & LIPKIN, 2001)
4. Cell cycle stabilization	(Jensen et al., 2001)
5. Promotion of apoptosis	(Welsh, 1994) (Welsh et al., 1998) (Mathiasen et al., 1999)
6. Anti-neoangiogenesis	(Majewski et al., 1996) (Pendás-Franco et al., 2008) (Mantell et al., 2000)
7. Down-regulation of glycogen synthase kinase 3 (GSK-3) *reduces proliferation of colorectal, prostate, & pancreatic cancers in vitro	(Ougolkov & Billadeau, 2006)
8. Downregulation of the canonical Wnt signaling pathway *Wnt signaling pathway is active in colorectal and other cancers	(Ordóñez-Morán et al., 2008)
9. Increased expression of DKK-1 protein *DKK-1 is a tumor suppressor protein in colon cancer cells	(González-Sancho et al., 2005)
10. Down-regulation of DKK-4 transcription *DKK-4 is up-regulated in colorectal cancer	(González-Sancho et al., 2005)

Table 4.3: Proposed mechanisms that account for the role of vitamin D and calcium in reduction of cancer rate and death

A study relying on data from observational studies and randomized trials predicted that elevation in blood vitamin D to 40-60 ng/mL would prevent nearly 58k new cases of breast cancer and 49k new cases of colorectal cancer in the USA and Canada. They also predicted such levels may cut down cancer mortality to three fourths (Garland et al., 2009).

4.4 Asthma

Asthma is a top noncommunicable disease that can afflict people at any age. In fact, it is the leading chronic illness among children. In 2019 an estimated 262 million were affected and 461000 died because of asthma (“Global Burden of Disease: GBD Cause and Risk Summaries,” 2020). Last decade has seen the highest increase in asthma patients, and accumulating evidence link vitamin D deficiency to its pathogenesis and prognosis (Litonjua & Weiss, 2007). Asthma is characterized by inflammation and tightening of the muscles around the small airways causing air passages in the lungs to become narrow. In patients with asthma, vitamin D level represents a critical marker of asthma severity and treatment response.

There are many theories on the underlying mechanism of calcitriol’s role in the pathogenesis of asthma. One theory points to calcitriol’s impact in immune modulation. Respiratory infections are presumed to serve as trigger for a worsening of asthma and vitamin D is known to inhibit the development of respiratory infections (Urashima et al., 2010). Genetic variations in Vitamin D Receptor (VDR) have also been identified as a risk factor for asthma in patient cohorts (Poon et al., 2004).

Asthma is managed with inhaled medications such as glucocorticoids to decrease inflammation present in the air passages. However, asthma patients respond variably to inhaled corticosteroids. Results from a population-based study suggest that children with low Vitamin D levels require more inhaled corticosteroids (Brehm et al., 2009). A clinical investigation conjectured that vitamin D influences glucocorticoid response in asthma (Sutherland et al., 2010). The results from the investigation supported the theory and found that adequate levels of Vitamin D enhance glucocorticoid response. When stimulated by calcitriol, human CD⁴⁺ T cells increase their production of IL-10, which can potentially help overcome the poor corticosteroids responsiveness in glucocorticoid resistant (Xystrakis et al., 2006). In addition, vitamin D was found to improve lung function and decrease airway hyperresponsiveness (Sutherland et al., 2010). Accumulating evidence suggest that pregnant women can reduce the risk of early childhood asthma by almost 40% by taking vitamin D during their gestational period (Litonjua & Weiss, 2007)

4.5 Chronic Obstructive Lung Disease (COPD)

The World Health Organization (WHO) ranked Chronic Obstructive Pulmonary Disease (COPD) as third among the world's leading diseases causing mortality. In 2019 alone it was responsible for 3.23 million deaths. COPD is characterized by persistent and progressive respiratory distress including breathing difficulties, coughing, and production of phlegm. Such symptoms are result of anomaly in the small airways of the lungs that limit the airflow in and out of the lungs. The airways can become narrow due to blockage by mucus, and/or injury of the lung causing inflammation and distension of the airway lining.

Observational studies and epidemiologic evidence link COPD to vitamin D status. Adequate vitamin D status correlates to better pulmonary function (Black & Scragg, 2005) and vitamin D deficiency is predominant among patients awaiting lung transplantation (Førli et al., 2004). A study published in 2010 found high prevalence of subnormal vitamin D levels in COPD that correlates with disease severity (Janssens et al., 2010).

The mechanisms linking vitamin D depletion with the development of COPD are mostly hypothetical. Vitamin D deficiency is known to cause osteoporosis and fractures. Alterations in the thoracic skeleton may result in malfunction of the respiratory muscles which is conducive to the pathophysiology of COPD. In women, multiple thoracic vertebral fractures was found to decline vital capacity as well as total lung capacity (Leech et al., 1990). Another study of COPD patients observed a strong link between COPD severity and fractures (Nuti et al., 2009).

COPD is exacerbated by growth of an abnormal flora composed of viruses or bacteria or a combination of both (Papi et al., 2006). Vitamin D deficiency may alter host immunity of the lung, allowing the growth of such microbes which ultimately trigger inflammation.

Calcitriol maintains the extracellular matrix homeostasis of bone tissue and the lung. Matrix metalloproteinase-9 (MMP-9) has been suggested contribute in the development of COPD (Culpitt et al., 2005). In induced sputum of subjects suffering from COPD, MMP-9 was found to be elevated. Adequate levels vitamin D in keratinocytes inhibits TNF-alpha from inducing upregulation of MMP-9 (Bahar-Shany et al., 2010). Therefore, it is possible that deficiency of vitamin D decreases the attenuation of MMP-9 activity, subsequently increasing degradation of lung parenchyma.

4.6 Diabetes

Diabetes is one of the four major non communicable diseases. It has long been noticed that as humans age, serum calcifediol falls (McKenna, 1992) and serum glucose rises (Harris et al., 1998). Comparative studies revealed calcifediol concentration in blood is lower in subjects diagnosed with type 2 diabetes compared to nondiabetic control subjects (Pietschmann et al., 1988; Scragg et al., 1995). A study published in 1995 revealed that patients at risk for diabetes had lower calcifediol levels compared to those who were not at risk for diabetes (Boucher et al., 1995). The same study also found that among subjects at high risk for diabetes, poor vitamin D levels was associated with impaired insulin secretion. In a prospective population-based study of elderly men, hypovitaminosis D was linked to hyperresponsive insulin secretion after an oral glucose tolerance test (Baynes et al., 1997). Consequently, vitamin D has long been suspected of contributing to the pathogenesis of type 2 diabetes by triggering insulin resistance and reducing insulin secretion (Boucher, 1998).

It has been long established that onset of insulin resistance triggers diabetes. When cells become resistant to insulin, β -cells release more insulin in an attempt to prevent hyperglycemia. As this hyperactivity grows, β cells experience acute elevation in Ca^{2+} and reactive oxygen species (ROS) signaling that ultimately results in cell death and marks the beginning of diabetes (Newsholme et al., 2016). In normal physiological condition, vitamin D acts to preserve the normal levels of both Ca^{2+} and ROS that increase in β -cells during diabetes (Berridge, 2015). Therefore, in hypovitaminosis D, with depleted reserves, vitamin D cannot efficiently reduce inflammation, which eventually initiates insulin resistance.

The discovery of VDR in pancreatic β cells and Vitamin D dependent calcium-binding proteins (calbindins) in pancreatic tissue, suggested a role for vitamin D in insulin secretion (Johnson et al., 1994). Currently numerous VDR gene polymorphisms have been recognized to be associated to type 2 diabetes mellitus and insulin release (Ortlepp et al., 2001). It has been hypothesized that vitamin D affects several pathways involved in insulin secretion. According to one experimental data, calcitriol affects β -cell insulin secretion and synthesis by increasing intracellular concentration of calcium through non-selective voltage-dependent calcium channels (Sergeev, 2016). Vitamin D, likely, also assists in maintaining β -cell calcium-dependent endopeptidases involved in cleavage of proinsulin to insulin (Boucher, 1998). Calcium not only helps with insulin exocytosis but also with β -cell glycolysis, which signals concentration of glucose in circulation (Boucher, 1998)

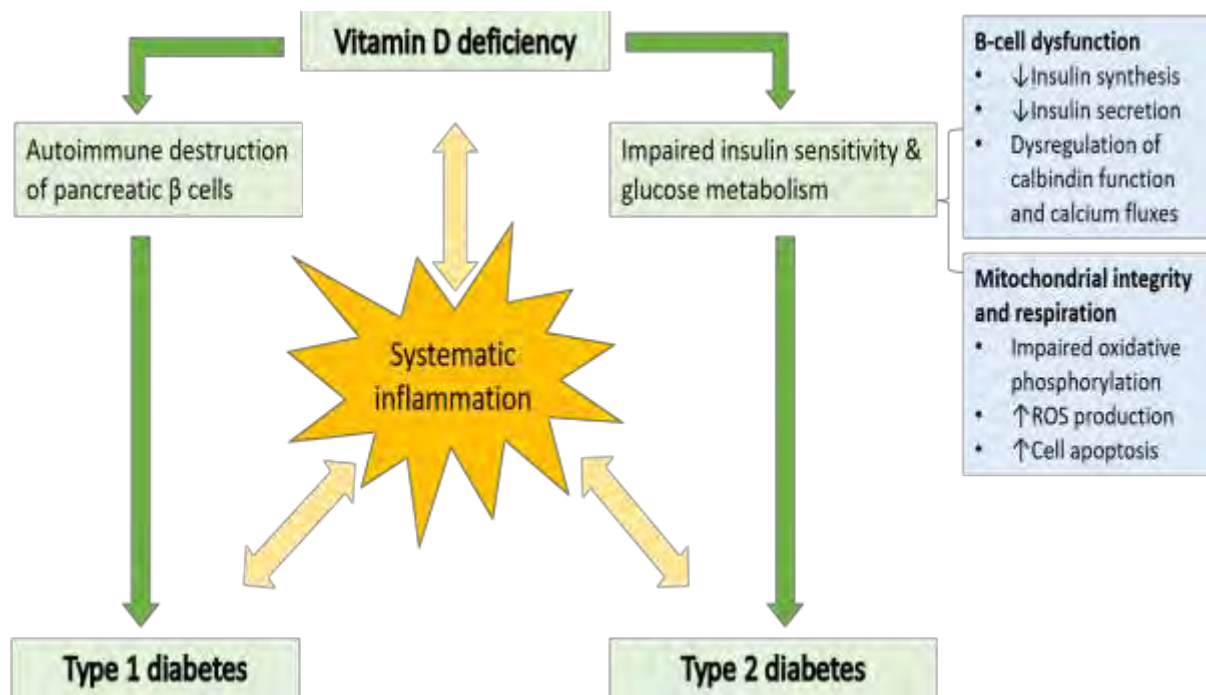


Figure 4.2: Vitamin D deficiency and development of diabetes

Vitamin D plays a significant role in preserving the epigenome (Berridge, 2017). Epigenetic alterations are a hallmark of diabetes where hypermethylation inactivates many diabetes-related genes. Vitamin D prevents hypermethylation of several gene promoter regions of numerous diabetes-related genes by enhancing the expression of the enzyme DNA demethylase.

Vitamin D repletion has been shown to ameliorate and even prevent type 1 diabetes mellitus (Stene et al., 2000; The EURODIAB Substudy 2 Study Group, 1999). More recent studies found that Vitamin D supplementation may be able to prevent the onset of diabetes type 2 (Wimalawansa, 2018). Supplementation with active vitamin D₃ might help improve the insulin resistance and decrease risk in predisposed patients (Berridge, 2017). In diabetic patients with established hypovitaminosis D, vitamin D replenishment was found to improve both glycemia and insulin secretion (Kumar et al., 1994). Frequent replenishment with large enough doses of vitamin D was able to enhance insulin sensitivity among insulin resistant and vitamin D deficient patients (von Hurst et al., 2010).

4.7 Bone diseases (Osteoporosis, Osteomalacia, Rickets)

Because of its key role in calcium metabolism, Vitamin D sufficiency is critical for maintaining skeletal health. Chronic inadequacy of calcitriol leads to (Lips, 2001)-

- Secondary hyperparathyroidism
- High bone turnover
- Progressive bone loss
- Mineralization defects
- Fragility fractures

Among numerous functions attributed to vitamin D, the main function is to aid the processes necessary to maintain healthy, mineralized bones. Vitamin D enhances intestinal absorption of calcium and phosphorus, therefore depleted vitamin D levels cause calcium malabsorption. Combined results from multiple studies establish that at serum calcidiol concentrations below ~80 nmol/L there is calcium malabsorption and thus enhanced risk of fracture (Heaney, 2004). Osteoporosis, osteomalacia and rickets are major hypovitaminosis D related bone diseases. The figure below depicts the relationship between the level of serum calcidiol concentration and the risk of various hypovitaminosis D related bone disease.

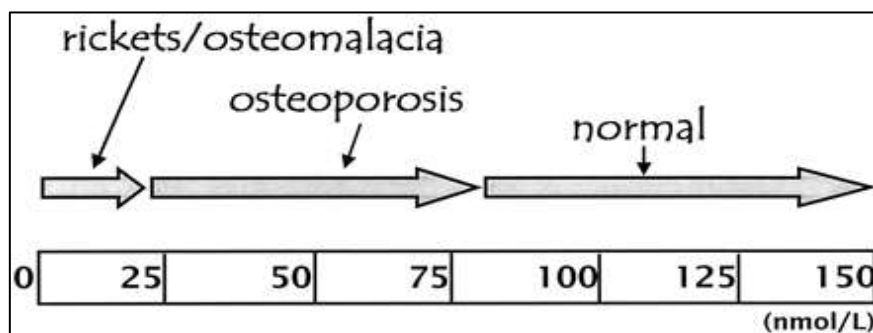


Figure 4.3. Proposed mapping of hypovitaminosis D induced bone diseases onto the serum calcidiol concentration continuum (Heaney, 2004)

Osteoporosis is a major NCD and the most prevalent bone disease, globally present at rate of 1 in 3 among women and 1 in 5 among men above the age of 50 (Kanis et al., 2000; Melton et al., 1992, 1998). Osteoporosis results in fragility fractures that often lead to other conditions such as myocardial infarction, cancer or death. In children, severe vitamin D deficiency leads to rickets, a bone disease characterized by bone pain and skeletal deformities. In adults, severe vitamin D deficiency leads to the development of a condition named osteomalacia. Osteomalacia is characterized by incomplete osteoid mineralization (Lips et al., 2006).

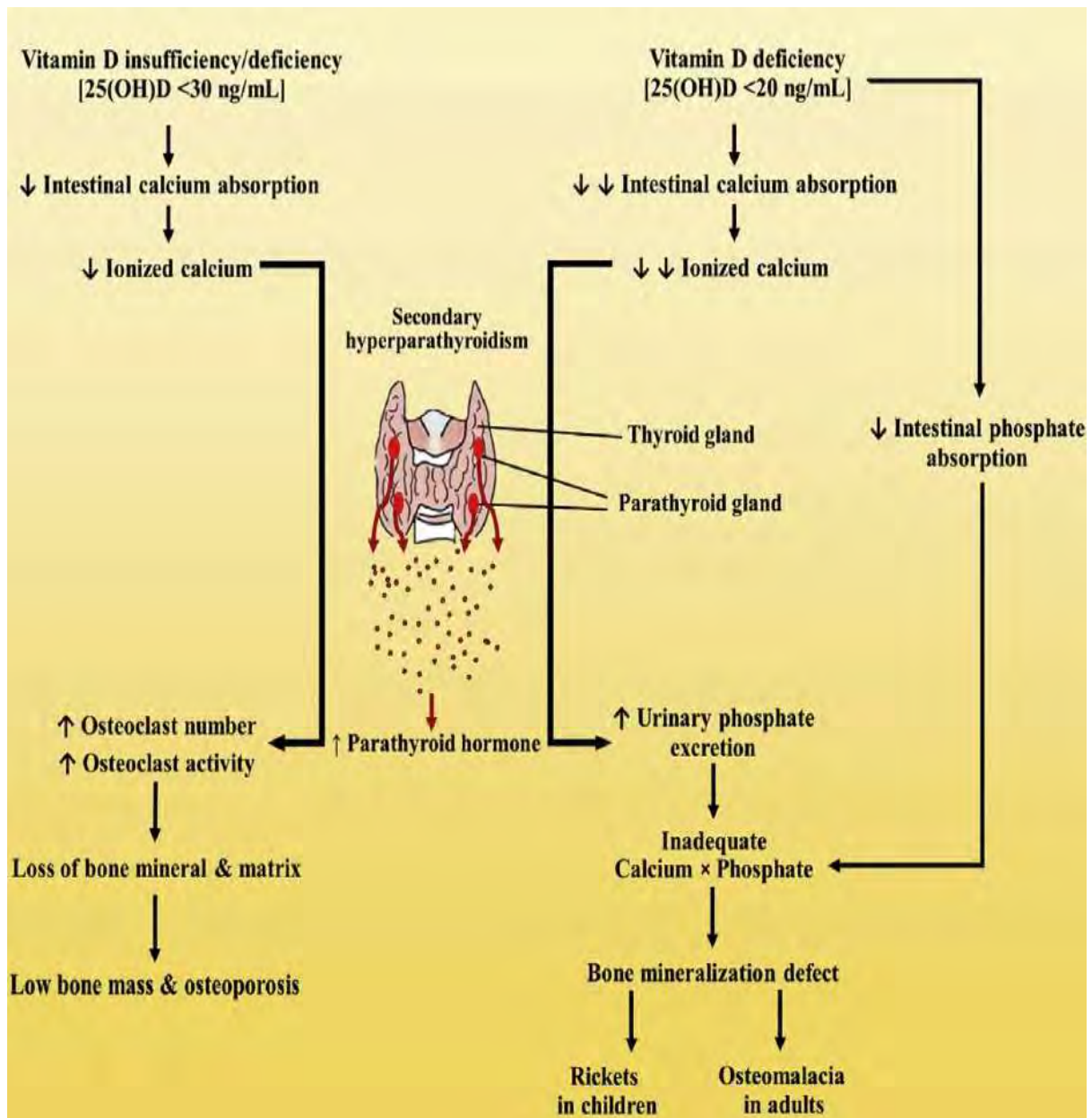


Figure 4.4: Vitamin D deficiency pathway to Osteoporosis, osteomalacia and Rickets development (Charoenngam et al., 2019)

There is strong consensus over necessity of vitamin D in preventing rickets and osteomalacia (Anderson & Atkins, 2008). Vitamin D's most studied action is its ability to maintain physiological levels of serum calcium and phosphate through calcium and phosphate absorption, renal tubular calcium reabsorption, and calcium mobilization from the bone (Charoenngam et al., 2019). Calcitriol is a strong stimulant for active transport of calcium in the intestine (O'Loughlin & Morris, 1998) and its primary role on bone is to mediate fluctuations in the extracellular fluid (ECF) concentration of calcium and phosphate, which indirectly impacts bone mineralization.

Decreased calcium and phosphate absorption due to vitamin D deficiency elevates parathyroid hormone (PTH) causing secondary hyperparathyroidism. These events trigger the release of calcium from the bones in order to maintain calcium homeostasis in a process known as bone resorption (Lips, 2001). During bone resorption, existing bone is broken down by osteoclasts, and replaced with new bone by osteoblasts (Jilka, 2003; Parfitt, 2002; Zaidi, 2007). Calcitriol and PTH stimulate osteoblasts to express RANKL, which in turn activates osteoclasts. Increased osteoclast activity results in enhanced bone resorption, loss of bone mineral and matrix, and ultimately decreased bone mass and causes osteoporosis (Charoenngam et al., 2019). Incessant bone turnover and resorption weakens bone architecture and increases fracture risk (Lips, 2001). Increase of PTH has a phosphaturic effect in the body, i.e., increase in phosphate excretion through urine. Decline in intestinal phosphate absorption combined with urinary phosphate excretion results in inadequate calcium-phosphate product, leading to defective bone mineralization and development of osteomalacia and rickets (Charoenngam et al., 2019).

It is difficult to assess whether beneficial effect of vitamin D on skeletal health is solely due to this “calcemic effect”. In vivo, administration of calcium and phosphate alone can correct the effects of vitamin D deficiency on skeleton (Underwood & DeLuca, 1984). Such finding signifies that at least for bone mineralization, calcitriol is not an absolute requirement. However, calcitriol may exert a direct response on cartilage and bone by stimulating normal skeletal development and turnover (Bikle, 2014). Increasing evidence suggest that vitamin D plays a direct role throughout normal bone metabolism by regulating bone cell activity (Atkins et al., 2007; Driel et al., 2006).

Supplementation with vitamin D suppresses PTH function and therefore bone turnover (Prestwood et al., 1996). Results from a meta-analysis of multiple randomized controlled trials suggest a positive dose response association between supplementation with vitamin D and prevention of fractures (Bischoff-Ferrari et al., 2009). Although vitamin D has not been proved to directly cause bone diseases, it is evident that maintaining adequate vitamin D level throughout life is critical to prevent osteomalacia, and to reduce or avoid the onset of osteoporosis in old age.

4.8 Chronic Liver Disease

Hypovitaminosis D is a common phenomenon in subjects with chronic liver disease, manifested by 92% of all patients (Arteh et al., 2010). Cholestatic liver disorders cause impaired intestinal absorption, hence, in the past, hypovitaminosis D was believed to be predominant only in cholestatic liver disorders (Collier et al., 2002). Growing body of evidence, however, demonstrate its widespread presence in chronic liver disease, irrespective of etiology.

Non-alcoholic fatty liver disease (NAFLD) has long surpassed viral hepatitis and alcoholic fatty liver disease as the most common form of chronic liver disease (Lazo & Clark, 2008). NAFLD encompasses an extensive spectrum of liver damage conditions including steatosis, fibrosis and cirrhosis that can advance to liver failure and hepatocellular carcinoma (Marchesini, 2003). NAFLD pathogenesis is incompletely understood but it has been hypothesized that a number of diverse parallel processes involving extrahepatic factors may lead to the development and progression of liver inflammation (Tilg & Moschen, 2010). One of those extrahepatic factors is vitamin D level. A study confirmed that relative to control subjects, subjects with biopsy-proven NAFLD have lower serum 25-hydroxycholecalciferol concentrations (Targher et al., 2007). Another study showed that, irrespective of sex, age, BMI, race, history of diabetes, peripheral vascular disease, renal disease, liver diseases and hypertension, there is an inverse relationship between low serum 25-hydroxycholecalciferol levels and NAFLD (K. L. Jablonski et al., 2013)

Vitamin D deficiency has been recognized as an independent risk factor for severe liver fibrosis (Petta et al., 2010). Serum levels of 25-hydroxyvitamin D represent a critical marker predicting mortality in advanced liver cirrhosis (Paternostro et al., 2017).

Intervention studies with vitamin D supplementation, however, have failed to provide enough evidence on the positive effect of vitamin D supplementation on liver related morbidity and quality of life (Bjelakovic et al., 2017).

While it is evident that a relationship between vitamin D and liver disease exists, it is unclear whether it's deficiency of vitamin D that confers an enhanced risk to liver disease or it's the other way around where disease of the liver causes vitamin D deficiency.

4.9 Chronic Kidney Disease

Chronic kidney disease (CKD) is an emerging non communicable disease and one of the most important predictors of premature cardiovascular disease. Patients with CKD exhibit dramatically high rate of severe vitamin D deficiency, which is further exacerbated by the reduced ability to convert calcifediol into the hormone form calcitriol (Jones, 2007). The kidneys have a significant role in vitamin D metabolism. They not only offer the enzymatic system for the calcitriol synthesis but they are also involved in the uptake of filtrated calcifediol from the urine (Nykjaer et al., 1999). Consequently, it is not surprising that impaired renal function leads to vitamin D deficiency. In fact, even early-stage CKD patients experience vitamin D deficiency (Levin et al., 2007). While it is unclear whether hypovitaminosis D triggers kidney disease, it is certain vitamin D deficiency accelerates the progression of kidney disease. Growing body of evidence have established a relationship between vitamin D deficiency and progression of CKD, and they have also recognized plasma vitamin D status to be an independent inverse predictor of disease progression and mortality in patients with chronic kidney disease (S. Williams et al., 2009). Vitamin D and its receptor are involved in regulation of the renin–angiotensin system (RAS) (Li et al., 2004) and the nuclear factor (NF)- κ B pathway (Sun et al., 2006). Both pathways are involved in a broad range of pathological process affecting the kidneys. Vitamin D has been recognized for its impressive therapeutic effects in prevention and intervention of various kidney diseases. Strong data suggest that adequate replenishment of vitamin D in deficient populations could potentially reduce premature morbidity and mortality in chronic kidney disease (Li, 2010).

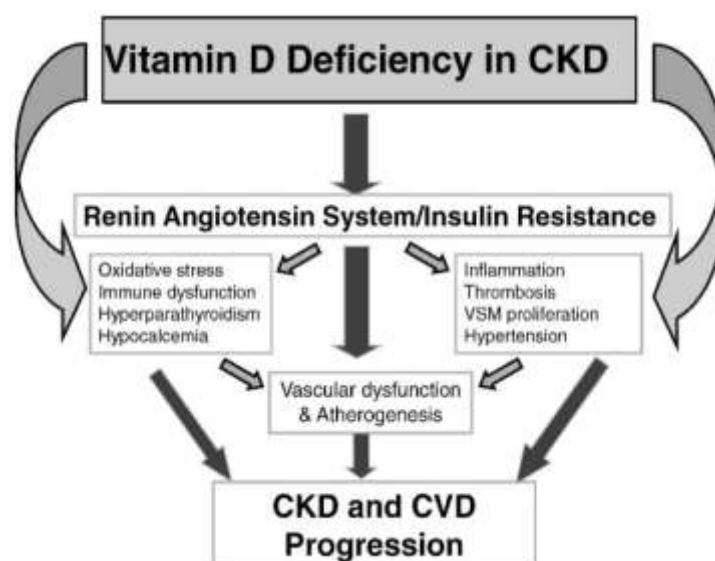


Figure 4.5: Major pathways linking vitamin D deficiency to CKD progression and complications (S. Williams et al., 2009).

4.10 Irritable Bowel Syndrome (IBS)

Irritable bowel syndrome (IBS) is a prevailing gastrointestinal disorder experienced by millions of individuals worldwide. IBS patients are often diagnosed with hypovitaminosis D (Khayyat & Attar, 2015). Data from epidemiological and clinical studies suggest that poor vitamin D levels might be accountable for IBS pathogenesis. However, there is poor consensus on the exact role of Vitamin D in IBS pathogenesis. It is generally assumed that Vitamin D improves the IBS by improving the factors involved in the development of IBS. In the gut vitamin D is known to modulate function of intestinal microbiota, regulate expression of antimicrobial peptide and exert a protective effect on epithelial walls of the intestinal mucosa (Akimbekov et al., 2020).

Some hypotheses draw a relationship between vitamin D and IBS through calcium. It has been long known that Vitamin D promotes gastrointestinal absorption of calcium and its subsequent entry into the circulation. From the circulation, calcium is carried to various tissues. In the skin tissue, calcium plays a role in decreasing the reactivity of skin to inflammatory stimuli (Klauder & Brown, 1929). It has been proposed that, calcium may have a similar role in the epithelial cells of the large intestine, where it effectively reduces the inflammatory response to gut flora and other agents in colon lumen (Garland & Garland, 1980). According to this model, people likely experience IBS due to inadequate intracellular calcium levels caused by depleted reserve of vitamin D. Another study, conducted more recently, attempted to identify potential gene expression-based biological signatures and biomarkers of IBS (Dussik et al., 2018). Their investigation was able to identify potential IBS genes, and calcitriol, the active version of vitamin D, was found to be a regulator of a subset of potential IBS genes. Such findings provide evidence to support the theory that pathological gene expression of IBS biomarkers may be initiated due to low circulating vitamin D levels. In line with this assumption, it can also be assumed that an increase in serum vitamin D status may ameliorate IBS symptoms by reversing pathological gene expression of the identified IBS biomarkers. An inverse relationship between serum vitamin D level and IBS symptom severity has also been extensively reported (C. E. Williams et al., 2018). In light of that, intervention studies have been conducted to assess the effect of vitamin D supplementation in IBS population. Most of the trials found Vitamin D supplementation to ameliorate the symptoms and quality of life in patients with IBS (Jalili et al., 2019; Abbasnezhad et al., 2016). Although the underlying mode of action of vitamin D in the pathogenesis of IBS lacks clear understanding, maintaining an ideal vitamin D status seems to have a positive effect in gut health.

4.11 Multiple Sclerosis

Multiple sclerosis (MS) is a noncommunicable neurodegenerative disorder that impacts the central nervous system (CNS). It is chronic, inflammatory, and of probable autoimmune origin. In multiple sclerosis there is a recurring or progressive demyelination and degeneration of the central nervous system. The demyelination and degeneration are believed to be caused by an erroneous immune response. Hypovitaminosis D has been identified as an environmental factor that increases susceptibility of developing MS. The role of vitamin-D in the development of MS has been the focus of intense research. A prospective nested case-control study with data from more than 7 million US military personnel records found that high serum concentrations of calcidiol confers decreased MS risk (Munger et al., 2006). Another prospective study conducted 6 years later confirmed these findings and stated that calcidiol level over 75 nmol/L lowers the risk of developing MS later in life (Salzer et al., 2012). Vitamin D intake from supplements has a protective effect and is inversely associated with the risk of MS (Munger et al., 2004). Extensive number of studies suggest that vitamin D is not only protective against the development MS, but it may also have a beneficial role in disease progression. In case of relapsing-remitting MS depletion of vitamin D increases risk of exacerbation whereas repletion may a beneficial effect on disease activity (Simpson et al., 2010; Runia et al., 2012).

Relapsing-remitting and secondary-progressive forms of multiple sclerosis are often treated with interferon- β , a cytokine in the interferon family. Vitamin D influences the prognosis of relapsing-remitting MS patients being treated with interferon- β (Simpson et al., 2010). It has been reported that, among patients undergoing interferon- β treatment, the lowest rate of new lesions was observed in patients with 25-hydroxyvitamin D levels over 100 nmol/L (Fitzgerald et al., 2015).

The involvement of Vitamin D in the pathogenesis and progression of MS does not seem to be straightforward. After the development of new genetic tools such as next-generation sequencing, and the fast growth of epigenomics, scientists have realized that the association between vitamin D and MS might be based on the widespread and typical genomic binding of the vitamin D receptor (VDR) (Lu et al., 2018).

5. Discussion and Conclusion

The unique conditions of the twenty-first century have let chronic noncommunicable diseases surpass infectious diseases as the major killers worldwide. NCDs are deemed as the most “democratic” diseases as they can affect anyone regardless of their socioeconomic or geographical background. As the burden of NCDs increases, it is important to identify and address any risk factor that contributes to the growing number. For more than a century, academics have been reporting an inverse correlation between latitude and many chronic diseases like diabetes, hypertension, common cancers, and numerous autoimmune diseases. By now, it is clear that vitamin D deficiency is a shared risk factor among many chronic non-communicable diseases. In this review, several lines of evidence were presented, suggesting that hypovitaminosis D has direct and indirect actions on non-communicable disease development.

Vitamin D is unique amid many vitamins and hormones in that its production is not enzymatic but depends on photo radiation. Solar UVB is capable to transform 7-dehydrocholesterol (7-DHC) into previtamin D₃, which undergoes thermal isomerization to form cholecalciferol. Vitamin D can also be obtained to some extent from dairy food and wild fatty fish, such as salmon and mackerel. Both vitamin D produced in the skin from 7-DHC or absorbed from food or supplement, must be activated first to calcidiol and then to its active form calcitriol. Photoproduction of calcitriol is limited by environmental conditions such as season, latitude, altitude, clothing, sunscreen, pollution and physiological conditions such as skin pigmentation, aging.

Vitamin D is remarkable considering the numerous cellular activities maintained by its active form, calcitriol. When vitamin D level is inadequate, many of these activities begin to decline, thereby setting the stage for the onset of many non-communicable diseases. Calcitriol influences a variety of biological pathways in parallel. In addition, basic mechanism of action that leads to the development of most NCDs is still not clear. Therefore, calcitriol’s involvement in their molecular pathogenesis pathways is likely difficult to prove.

Most cells in the body, including most cancerous cells, express Vitamin D receptor. Most tissues have not only VDRs, but also the hydroxylase enzyme that converts calcidiol to the active form, calcitriol. Calcitriol is a key calciotropic hormone. It’s calciotropic activities have immense significance in maintaining skeletal health. Poor vitamin D levels and subsequent

poor calcium and phosphate levels pave the way for non-communicable bone diseases such as osteoporosis, osteomalacia and in children, rickets.

Current data demonstrate that calcitriol activities extend beyond maintaining skeletal health. The effects of calcitriol extend to tissues that are not necessarily involved in calcium and phosphate homeostasis. Receptors for vitamin D have been detected in respiratory, renal, cardiovascular, gastrointestinal, reproductive, nervous and immune system. Thereby many chronic NCDs such as COPD, asthma, diabetes, cancer, CVD, hypertension, MS, chronic kidney disease, chronic liver disease have been linked to vitamin D deficiency.

When the levels are adequate, calcitriol acts to reduce inflammation, and inflammation is the first step in the pathway of pathogenesis of many NCDs. Poor vitamin D levels not only increases inflammation but also oxidative stress, and expression of immune cells, i.e., monocytes and macrophages. Vitamin D also maintains normal mitochondrial activity by supporting respiratory chain's physiological electron flow, thus preventing the formation of ROS. Additional molecular mechanisms that are directly or indirectly regulated by calcitriol are the critical elements that coherently modulate the cell cycle progression. In fact, calcitriol was found to have the ability to inhibit proliferation and induce terminal differentiation of tumor cells and various other cells. The antiproliferative effects of calcitriol have been repeatedly confirmed in many laboratories.

For most NCDs discussed in this paper, vitamin D supplementation in deficient population was found to have beneficial effect on disease progression and in some cases even potentially prevent disease incidence. In some cases, the effects have been proven in vitro or in animal trials in vivo but not in humans. For example, numerous animal studies have showed that calcitriol or its analogs can reduce cancer growth in vivo, yet clinical trials in humans are still missing.

Although a multitude of epidemiological studies suggest an inverse relationship between vitamin D status and a large number of NCDs, such relations have not been universally confirmed. Conclusive data from clinical and interventional studies are missing for many of these disease entities. Study limitations limit the extent to which inverse relations can be attributed to vitamin D, and additional studies are needed to further understand the nature of this association. The long-term outcome of cholecalciferol therapy in patients with various chronic NCDs needs further exploration through long-term intervention studies.

6. References

1. Abbasnezhad, A., Amani, R., Hajiani, E., Alavinejad, P., Cheraghian, B., & Ghadiri, A. (2016). Effect of vitamin D on gastrointestinal symptoms and health-related quality of life in irritable bowel syndrome patients: a randomized double-blind clinical trial. *Neurogastroenterology & Motility*, 28(10), 1533–1544. <https://doi.org/10.1111/nmo.12851>
2. Abe, E., Miyaura, C., Sakagami, H., Takeda, M., Konno, K., Yamazaki, T., Yoshiki, S., & Suda, T. (1981). Differentiation of mouse myeloid leukemia cells induced by 1 α ,25-dihydroxyvitamin D₃. *Proceedings of the National Academy of Sciences of the United States of America*, 78(8 I), 4990–4994. <https://doi.org/10.1073/pnas.78.8.4990>
3. Abe, J., Moriya, Y., Saito, M., Sugawara, Y., Suda, T., & Nishii, Y. (1986). Modulation of Cell Growth, Differentiation, and Production of Interleukin-3 by 1 α ,25-Dihydroxyvitamin D₃ in the Murine Myelomonocytic Leukemia Cell Line WEHI-3. *Cancer Research*, 46(12_Part_1), 6316–6321.
4. Akimbekov, N. S., Digel, I., Sherelkhan, D. K., Lutfor, A. B., & Razzaque, M. S. (2020). Vitamin d and the host-gut microbiome: A brief overview. *Acta Histochemica et Cytochemica*, 53(3), 33–42. <https://doi.org/10.1267/ahc.20011>
5. Anderson, P. H., & Atkins, G. J. (2008). The skeleton as an intracrine organ for vitamin D metabolism. In *Molecular Aspects of Medicine* (Vol. 29, Issue 6, pp. 397–406). Pergamon. <https://doi.org/10.1016/j.mam.2008.05.003>
6. Arteh, J., Narra, S., & Nair, S. (2010). Prevalence of vitamin D deficiency in chronic liver disease. *Digestive Diseases and Sciences*, 55(9), 2624–2628. <https://doi.org/10.1007/s10620-009-1069-9>
7. Atkins, G. J., Anderson, P. H., Findlay, D. M., Welldon, K. J., Vincent, C., Zannettino, A. C. W., O'Loughlin, P. D., & Morris, H. A. (2007). Metabolism of vitamin D₃ in human osteoblasts: Evidence for autocrine and paracrine activities of 1 α ,25-dihydroxyvitamin D₃. *Bone*, 40(6), 1517–1528. <https://doi.org/10.1016/j.bone.2007.02.024>
8. Bahar-Shany, K., Ravid, A., & Koren, R. (2010). Upregulation of MMP-9 production by TNF α in keratinocytes and its attenuation by vitamin D. *Journal of Cellular Physiology*, 222(3), 729–737. <https://doi.org/10.1002/jcp.22004>
9. Baynes, K. C. R., Boucher, B. J., Feskens, E. J. M., & Kromhout, D. (1997). Vitamin D, glucose tolerance anal insulinaemia in elderly men. *Diabetologia*, 40(3), 344–347. <https://doi.org/10.1007/s001250050685>

10. Berridge, M. J. (2015). Vitamin D: A custodian of cell signalling stability in health and disease. In *Biochemical Society Transactions* (Vol. 43, Issue 3, pp. 349–358). Portland Press Ltd. <https://doi.org/10.1042/BST20140279>
11. Berridge, M. J. (2017). Vitamin D deficiency and diabetes. In *Biochemical Journal* (Vol. 474, Issue 8, pp. 1321–1332). Portland Press Ltd. <https://doi.org/10.1042/BCJ20170042>
12. Bikle, D. D. (2014). Vitamin D metabolism, mechanism of action, and clinical applications. In *Chemistry and Biology* (Vol. 21, Issue 3, pp. 319–329). Cell Press. <https://doi.org/10.1016/j.chembiol.2013.12.016>
13. Bills, C. E. (1927). ANTIRICKETIC SUBSTANCES. *Journal of Biological Chemistry*, 72(2), 751–758. [https://doi.org/10.1016/s0021-9258\(18\)84347-2](https://doi.org/10.1016/s0021-9258(18)84347-2)
14. Bischoff-Ferrari, H. A., Dawson-Hughes, B., Staehelin, H. B., Orav, J. E., Stuck, A. E., Theiler, R., Wong, J. B., Egli, A., Kiel, D. P., & Henschkowski, J. (2009). Fall prevention with supplemental and active forms of vitamin D: A meta-analysis of randomised controlled trials. *BMJ (Online)*, 339(7725), 843. <https://doi.org/10.1136/bmj.b3692>
15. Bjelakovic, G., Nikolova, D., Bjelakovic, M., & Gluud, C. (2017). Vitamin D supplementation for chronic liver diseases in adults. In *Cochrane Database of Systematic Reviews* (Vol. 2017, Issue 11). John Wiley and Sons Ltd. <https://doi.org/10.1002/14651858.CD011564.pub2>
16. Black, P. N., & Scragg, R. (2005). Relationship between serum 25-hydroxyvitamin D and pulmonary function in the Third National Health and Nutrition Examination Survey. *Chest*, 128(6), 3792–3798. <https://doi.org/10.1378/chest.128.6.3792>
17. Boucher, B. J. (1998). Inadequate vitamin D status: does it contribute to the disorders comprising syndrome ‘X’? *British Journal of Nutrition*, 79(4), 315–327. <https://doi.org/10.1079/bjn19980055>
18. Boucher, B. J., Mannan, N., Noonan, K., Hales, C. N., & Evans, S. J. W. (1995). Glucose intolerance and impairment of insulin secretion in relation to vitamin D deficiency in East London Asians. *Diabetologia*, 38(10), 1239–1245. <https://doi.org/10.1007/BF00422375>
19. Brehm, J. M., Celedón, J. C., Soto-Quiros, M. E., Avila, L., Hunninghake, G. M., Forno, E., Laskey, D., Sylvia, J. S., Hollis, B. W., Weiss, S. T., & Litonjua, A. A. (2009). Serum vitamin D levels and markers of severity of childhood asthma in Costa Rica. *American Journal of Respiratory and Critical Care Medicine*, 179(9), 765–771. <https://doi.org/10.1164/rccm.200808-1361OC>

20. Challoumas, D. (2014). Vitamin D supplementation and lipid profile: What does the best available evidence show? In *Atherosclerosis* (Vol. 235, Issue 1, pp. 130–139). Elsevier Ireland Ltd. <https://doi.org/10.1016/j.atherosclerosis.2014.04.024>
21. Charoenngam, N., Shirvani, A., & Holick, M. F. (2019). Vitamin D for skeletal and non-skeletal health: What we should know. In *Journal of Clinical Orthopaedics and Trauma* (Vol. 10, Issue 6, pp. 1082–1093). Elsevier B.V. <https://doi.org/10.1016/j.jcot.2019.07.004>
22. Chen, S., Law, C. S., & Gardner, D. G. (2010). Vitamin D-dependent suppression of endothelin-induced vascular smooth muscle cell proliferation through inhibition of CDK2 activity. *Journal of Steroid Biochemistry and Molecular Biology*, 118(3), 135–141. <https://doi.org/10.1016/j.jsbmb.2009.11.002>
23. Chen, T. C., Chimeh, F., Lu, Z., Mathieu, J., Person, K. S., Zhang, A., Kohn, N., Martinello, S., Berkowitz, R., & Holick, M. F. (2007). Factors that influence the cutaneous synthesis and dietary sources of vitamin D. *Archives of Biochemistry and Biophysics*, 460(2), 213–217. <https://doi.org/10.1016/j.abb.2006.12.017>
24. Cheol Whee Park, Yong Seog Oh, Young Shin Shin, Kim, C. M., Kim, Y. S., Suk Young Kim, Eui Jin Choi, Yoon Sik Chang, & Byung Kee Bang. (1999). Intravenous calcitriol regresses myocardial hypertrophy in hemodialysis patients with secondary hyperparathyroidism. *American Journal of Kidney Diseases*, 33(1), 73–81. [https://doi.org/10.1016/s0272-6386\(99\)70260-x](https://doi.org/10.1016/s0272-6386(99)70260-x)
25. Collier, J. D., Ninkovic, M., & Compston, J. E. (2002). Guidelines on the management of osteoporosis associated with chronic liver disease. In *Gut* (Vol. 50, Issue SUPPL. 1, p. i1). BMJ Publishing Group. https://doi.org/10.1136/gut.50.suppl_1.i1
26. Colston, K., Colston, M. J., & Feldman, D. (1981). 1, 25-dihydroxyvitamin D3 and malignant melanoma: The presence of receptors and inhibition of cell growth in culture. *Endocrinology*, 108(3), 1083–1086. <https://doi.org/10.1210/endo-108-3-1083>
27. Cross, H. S. (2007). Extrarenal Vitamin D Hydroxylase Expression and Activity in Normal and Malignant Cells: Modification of Expression by Epigenetic Mechanisms and Dietary Substances. *Nutrition Reviews*, 65(SUPPL.2). <https://doi.org/10.1111/j.1753-4887.2007.tb00334.x>
28. Culpitt, S. v., Rogers, D. F., Traves, S. L., Barnes, P. J., & Donnelly, L. E. (2005). Sputum matrix metalloproteases: Comparison between chronic obstructive pulmonary disease and asthma. *Respiratory Medicine*, 99(6), 703–710. <https://doi.org/10.1016/j.rmed.2004.10.022>

29. Deeb, K. K., Trump, D. L., & Johnson, C. S. (2007). Vitamin D signalling pathways in cancer: Potential for anticancer therapeutics. In *Nature Reviews Cancer* (Vol. 7, Issue 9, pp. 684–700). Nat Rev Cancer. <https://doi.org/10.1038/nrc2196>
30. Driel, M., Koedam, M., Buurman, C. J., Hewison, M., Chiba, H., Uitterlinden, A. G., Pols, H. A. P., M. van Leeuwen, J. P. T., Driel, M., Koedam, M., Buurman, C. J., Hewison, M., Chiba, H., Uitterlinden, A. G., Pols, H. A. P., & M. van Leeuwen, J. P. T. (2006). Evidence for auto/paracrine actions of vitamin D in bone: 1 α -hydroxylase expression and activity in human bone cells. *The FASEB Journal*, 20(13), 2417–2419. <https://doi.org/10.1096/fj.06-6374fje>
31. Dussik, C. M., Hockley, M., Grozic, A., Kaneko, I., Zhang, L., Sabir, M. S., Park, J., Wang, J., Nickerson, C. A., Yale, S. H., Rall, C. J., Foxx-Orenstein, A. E., Borrer, C. M., Sandrin, T. R., & Jurutka, P. W. (2018). Gene expression profiling and assessment of vitamin D and serotonin pathway variations in patients with irritable bowel syndrome. *Journal of Neurogastroenterology and Motility*, 24(1), 96–106. <https://doi.org/10.5056/jnm17021>
32. Dziedzic, E. A., Smyk, W., Sowińska, I., Dąbrowski, M., & Jankowski, P. (2021). Serum level of vitamin d is associated with severity of coronary atherosclerosis in postmenopausal women. *Biology*, 10(11). <https://doi.org/10.3390/biology10111139>
33. El-Sonbaty, M. R., & Abdul-Ghaffar, N. U. (1996). Vitamin D deficiency in veiled Kuwaiti women. *European Journal of Clinical Nutrition*, 50(5), 315–318. <https://europepmc.org/article/med/8735313>
34. Eyles, D. W., Smith, S., Kinobe, R., Hewison, M., & McGrath, J. J. (2005). Distribution of the Vitamin D receptor and 1 α -hydroxylase in human brain. *Journal of Chemical Neuroanatomy*, 29(1), 21–30. <https://doi.org/10.1016/j.jchemneu.2004.08.006>
35. Feldman, D., Pike, J., & Adams, J. (2011). *Vitamin D: Two-Volume Set*. Academic Press.
36. Fitzgerald, K. C., Munger, K. L., Köchert, K., Arnason, B. G. W., Comi, G., Cook, S., Goodin, D. S., Filippi, M., Hartung, H. P., Jeffery, D. R., O'Connor, P., Suarez, G., Sandbrink, R., Kappos, L., Pohl, C., & Ascherio, A. (2015). Association of Vitamin D levels with multiple sclerosis activity and progression in patients receiving interferon Beta-1b. *JAMA Neurology*, 72(12), 1458–1465. <https://doi.org/10.1001/jamaneurol.2015.2742>
37. Førli, L., Halse, J., Haug, E., Bjørtuft, Vatn, M., Kofstad, J., & Boe, J. (2004). Vitamin D deficiency, bone mineral density and weight in patients with advanced pulmonary disease.

- Journal of Internal Medicine*, 256(1), 56–62. <https://doi.org/10.1111/j.1365-2796.2004.01337.x>
38. Förstermann, U., Xia, N., & Li, H. (2017). Roles of vascular oxidative stress and nitric oxide in the pathogenesis of atherosclerosis. In *Circulation Research* (Vol. 120, Issue 4, pp. 713–735). Lippincott Williams and Wilkins. <https://doi.org/10.1161/CIRCRESAHA.116.309326>
39. Fort, J. M., Azpiroz, F., Casellas, F., Andreu, J., & Malagelada, J. R. (1996). Bowel habit after cholecystectomy: Physiological changes and clinical implications. *Gastroenterology*, 111(3), 617–622. <https://doi.org/10.1053/gast.1996.v111.pm8780565>
40. GARLAND, C. F., & GARLAND, F. C. (1980). Do Sunlight and Vitamin D Reduce the Likelihood of Colon Cancer? *International Journal of Epidemiology*, 9(3), 227–231. <https://doi.org/10.1093/ije/9.3.227>
41. Garland, C. F., Gorham, E. D., Mohr, S. B., & Garland, F. C. (2009). Vitamin D for Cancer Prevention: Global Perspective. In *Annals of Epidemiology* (Vol. 19, Issue 7, pp. 468–483). <https://doi.org/10.1016/j.annepidem.2009.03.021>
42. Garland, C. F., Grant, W. B., Mohr, S. B., Gorham, E. D., & Garland, F. C. (2007). What is the Dose-Response Relationship between Vitamin D and Cancer Risk? *Nutrition Reviews*, 65(SUPPL.2). <https://doi.org/10.1111/j.1753-4887.2007.tb00349.x>
43. Getzenberg, R. H., Light, B. W., Lapco, P. E., Konety, B. R., Nangia, A. K., Acierno, J. S., Dhir, R., Shurin, Z., Day, R. S., Trump, D. L., & Johnson, C. S. (1997). Vitamin D inhibition of prostate adenocarcinoma growth and metastasis in the dunning rat prostate model system. *Urology*, 50(6), 999–1006. [https://doi.org/10.1016/S0090-4295\(97\)00408-1](https://doi.org/10.1016/S0090-4295(97)00408-1)
44. Giovannucci, E. (2009). Vitamin D and Cancer Incidence in the Harvard Cohorts. In *Annals of Epidemiology* (Vol. 19, Issue 2, pp. 84–88). *Ann Epidemiol.* <https://doi.org/10.1016/j.annepidem.2007.12.002>
45. Giovannucci, E., Liu, Y., Hollis, B. W., & Rimm, E. B. (2008). 25-Hydroxyvitamin D and risk of myocardial infarction in men: A prospective study. *Archives of Internal Medicine*, 168(11), 1174–1180. <https://doi.org/10.1001/archinte.168.11.1174>
46. Global Burden of Disease: GBD cause and risk summaries. (2020). *Lancet*, 396(10258), 1204–1222. <https://www.thelancet.com/gbd/summaries>
47. Gómez Alonso, C., Naves Díaz, M., Rodríguez García, M., Fernández Martín, J., & Cannata Andía, J. (2003). Review of the concept of vitamin D “sufficiency and insufficiency.” *Nefrología*, 23(2), 73–77. <https://pubmed.ncbi.nlm.nih.gov/12778859/>

48. González-Sancho, J. M., Aguilera, O., García, J. M., Pendás-Franco, N., Peña, C., Cal, S., de Herreros, A. G., Bonilla, F., & Muñoz, A. (2005). The Wnt antagonist DICKKOPF-1 gene is a downstream target of β -catenin/TCF and is downregulated in human colon cancer. *Oncogene*, *24*(6), 1098–1103. <https://doi.org/10.1038/sj.onc.1208303>
49. Hansdottir, S., Monick, M. M., Hinde, S. L., Lovan, N., Look, D. C., & Hunninghake, G. W. (2008). Respiratory Epithelial Cells Convert Inactive Vitamin D to Its Active Form: Potential Effects on Host Defense. *The Journal of Immunology*, *181*(10), 7090–7099. <https://doi.org/10.4049/jimmunol.181.10.7090>
50. Harris, M. I., Flegal, K. M., Cowie, C. C., Eberhardt, M. S., Goldstein, D. E., Little, R. R., Wiedmeyer, H. M., & Byrd-Holt, D. D. (1998). Prevalence of diabetes, impaired fasting glucose, and impaired glucose tolerance in U.S. adults: The Third National Health and Nutrition Examination Survey, 1988-1994. *Diabetes Care*, *21*(4), 518–524. <https://doi.org/10.2337/diacare.21.4.518>
51. Hatun, S., Islam, Ö., Cizmecioglu, F., Kara, B., Babaoglu, K., Berk, F., & Gökalp, A. S. (2005). Subclinical vitamin D deficiency is increased in adolescent girls who wear concealing clothing. *Journal of Nutrition*, *135*(2), 218–222. <https://doi.org/10.1093/jn/135.2.218>
52. Haussler, M. R., Haussler, C. A., Whitfield, G. K., Hsieh, J. C., Thompson, P. D., Barthel, T. K., Bartik, L., Egan, J. B., Wu, Y., Kubicek, J. L., Lowmiller, C. L., Moffet, E. W., Forster, R. E., & Jurutka, P. W. (2010). The nuclear vitamin D receptor controls the expression of genes encoding factors which feed the “Fountain of Youth” to mediate healthful aging. *Journal of Steroid Biochemistry and Molecular Biology*, *121*(1–2), 88–97. <https://doi.org/10.1016/j.jsbmb.2010.03.019>
53. Haussler, M. R., Whitfield, G. K., Haussler, C. A., Hsieh, J. C., Thompson, P. D., Selznick, S. H., Dominguez, C. E., & Jurutka, P. W. (1998). The nuclear vitamin D receptor: Biological and molecular regulatory properties revealed. In *Journal of Bone and Mineral Research* (Vol. 13, Issue 3, pp. 325–349). J Bone Miner Res. <https://doi.org/10.1359/jbmr.1998.13.3.325>
54. Heaney, R. P. (2004). Functional indices of vitamin D status and ramifications of vitamin D deficiency. In *The American journal of clinical nutrition* (Vol. 80, Issue 6 Suppl, pp. 1706S-1709S). Oxford Academic. <https://doi.org/10.1093/ajcn/80.6.1706s>
55. Hoffman, F. L. (1915). The mortality from cancer throughout the world. *Prudential Press*.

56. Holick, M. F. (1987). Photosynthesis of vitamin D in the skin: effect of environmental and life-style variables. *Fed Proc*, 46(5), 1876–1882. <https://pubmed.ncbi.nlm.nih.gov/3030826/>
57. Holick, M. F. (1994). McCollum award lecture, 1994: Vitamin D - New horizons for the 21st century. *American Journal of Clinical Nutrition*, 60(4), 619–630. <https://doi.org/10.1093/ajcn/60.4.619>
58. Holick, M. F. (2004). Vitamin D: Importance in the prevention of cancers, type 1 diabetes, heart disease, and osteoporosis. *American Journal of Clinical Nutrition*, 79(3), 362–371. <https://doi.org/10.1093/ajcn/79.3.362>
59. Holick, M. F. (2005). The vitamin D epidemic and its health consequences. *Journal of Nutrition*, 135(11). <https://doi.org/10.1093/jn/135.11.2739s>
60. Holick, M. F. (2007). Vitamin D Deficiency. *New England Journal of Medicine*, 357(3), 266–281. <https://doi.org/10.1056/NEJMra070553>
61. Holick, M. F., Matsuoka, L. Y., & Wortsman, J. (1989). AGE, VITAMIN D, AND SOLAR ULTRAVIOLET. In *The Lancet* (Vol. 334, Issue 8671, pp. 1104–1105). Lancet. [https://doi.org/10.1016/S0140-6736\(89\)91124-0](https://doi.org/10.1016/S0140-6736(89)91124-0)
62. Hosseinpanah, F., Pour, S. H., Heibatollahi, M., Moghbel, N., Asefzade, S., & Azizi, F. (2010). The effects of air pollution on vitamin D status in healthy women: A cross sectional study. *BMC Public Health*, 10(1), 519. <https://doi.org/10.1186/1471-2458-10-519>
63. Hyppönen, E., & Power, C. (2007). Hypovitaminosis D in British adults at age 45 y: Nationwide cohort study of dietary and lifestyle predictors. *American Journal of Clinical Nutrition*, 85(3), 860–868. <https://doi.org/10.1093/ajcn/85.3.860>
64. Jablonski, K. L., Jovanovich, A., Holmen, J., Targher, G., McFann, K., Kendrick, J., & Chonchol, M. (2013). Low 25-hydroxyvitamin D level is independently associated with non-alcoholic fatty liver disease. *Nutrition, Metabolism and Cardiovascular Diseases*, 23(8), 792–798. <https://doi.org/10.1016/j.numecd.2012.12.006>
65. Jablonski, N. G., & Chaplin, G. (2000). The evolution of human skin coloration. *Journal of Human Evolution*, 39(1), 57–106. <https://doi.org/10.1006/jhev.2000.0403>
66. Jalili, M., Vahedi, H., Poustchi, H., & Hekmatdoost, A. (2019). Effects of Vitamin D supplementation in patients with irritable bowel syndrome: A randomized, double-blind, placebo-controlled clinical trial. *International Journal of Preventive Medicine*, 10(1). https://doi.org/10.4103/ijpvm.IJPVM_512_17

67. Janssens, W., Bouillon, R., Claes, B., Carremans, C., Lehouck, A., Buyschaert, I., Coolen, J., Mathieu, C., Decramer, M., & Lambrechts, D. (2010). Vitamin D deficiency is highly prevalent in COPD and correlates with variants in the vitamin D-binding gene. *Thorax*, *65*(3), 215–220. <https://doi.org/10.1136/thx.2009.120659>
68. Jensen, S. S., Madsen, M. W., Lukas, J., Binderup, L., & Bartek, J. (2001). Inhibitory Effects of 1 α ,25-Dihydroxyvitamin D₃ on the G1–S Phase-Controlling Machinery. *Molecular Endocrinology*, *15*(8), 1370–1380. <https://doi.org/10.1210/mend.15.8.0673>
69. Jilka, R. L. (2003). Biology of the basic multicellular unit and the pathophysiology of osteoporosis. *Medical and Pediatric Oncology*, *41*(3), 182–185. <https://doi.org/10.1002/mpo.10334>
70. Johansen, C., Iversen, L., Ryborg, A., & Kragballe, K. (2000). 1 α ,25-dihydroxyvitamin D₃ induced differentiation of cultured human keratinocytes is accompanied by a PKC-independent regulation of AP-1 DNA binding activity. *Journal of Investigative Dermatology*, *114*(6), 1174–1179. <https://doi.org/10.1046/j.1523-1747.2000.00003.x>
71. Johnson, J. A., Grande, J. P., Roche, P. C., & Kumar, R. (1994). Immunohistochemical localization of the 1,25(OH)₂D₃ receptor and calbindin D(28k) in human and rat pancreas. *American Journal of Physiology - Endocrinology and Metabolism*, *267*(3 30-3). <https://doi.org/10.1152/ajpendo.1994.267.3.e356>
72. Jones, G. (2007). Expanding role for vitamin D in chronic kidney disease: Importance of blood 25-OH-D levels and extra-renal 1 α -Hydroxylase in the Classical and Nonclassical Actions of 1 α ,25-Dihydroxyvitamin D₃. *Seminars in Dialysis*, *20*(4), 316–324. <https://doi.org/10.1111/j.1525-139X.2007.00302.x>
73. Jorde, R., Svartberg, J., & Sundsfjord, J. (2005). Serum parathyroid hormone as a predictor of increase in systolic blood pressure in men. *Journal of Hypertension*, *23*(9), 1639–1644. <https://doi.org/10.1097/01.hjh.0000179764.40701.36>
74. Kanis, J. A., Johnell, O., Oden, A., Sernbo, I., Redlund-Johnell, I., Dawson, A., de Laet, C., & Jonsson, B. (2000). Long-term risk of osteoporotic fracture in Malmo. *Osteoporosis International*, *11*(8), 669–674. <https://doi.org/10.1007/s001980070064>
75. Kenny, D. E., O'Hara, T. M., Chen, T. C., Lu, Z., Tian, X., & Holick, M. F. (2004). Vitamin D content in Alaskan Arctic zooplankton, fishes, and marine mammals. *Zoo Biology*, *23*(1), 33–43. <https://doi.org/10.1002/zoo.10104>
76. Khayyat, Y., & Attar, S. (2015). Vitamin D deficiency in patients with irritable bowel syndrome: Does it exist? *Oman Medical Journal*, *30*(2), 115–118. <https://doi.org/10.5001/omj.2015.25>

77. KIMURA, Y., KAWAMURA, M., OWADA, M., OSHIMA, T., MUROOKA, M., FUJIWARA, T., & HIRAMORI, K. (1999). Effectiveness of 1,25-dihydroxyvitamin D Supplementation on Blood Pressure Reduction in a Pseudohypoparathyroidism Patient with High Renin Activity. *Internal Medicine*, 38(1), 31–35. <https://doi.org/10.2169/internalmedicine.38.31>
78. Klauder, J. v., & Brown, H. (1929). Experimental studies in eczema: III. The role of the sympathetic nervous system and calcium-potassium ratio in altering cutaneous irritability in the rabbit. *Archives of Dermatology and Syphilology*, 19(1), 52–65. <https://doi.org/10.1001/archderm.1929.02380190055004>
79. Kumar, S., Davies, M., Zakaria, Y., Mawer, E. B., Gordon, C., Olukoga, A. O., & Boulton, A. J. M. (1994). Improvement in glucose tolerance and beta-cell function in a patient with vitamin D deficiency during treatment with vitamin D. *Postgraduate Medical Journal*, 70(824), 440–443. <https://doi.org/10.1136/pgmj.70.824.440>
80. LAMPRECHT, S. A., & LIPKIN, M. (2001). Cellular Mechanisms of Calcium and Vitamin D in the Inhibition of Colorectal Carcinogenesis. *Annals of the New York Academy of Sciences*, 952(1), 73–87. <https://doi.org/10.1111/j.1749-6632.2001.tb02729.x>
81. Lazo, M., & Clark, J. M. (2008). The epidemiology of nonalcoholic fatty liver disease: A global perspective. In *Seminars in Liver Disease* (Vol. 28, Issue 4, pp. 339–350). Semin Liver Dis. <https://doi.org/10.1055/s-0028-1091978>
82. Leech, J. A., Dulberg, C., Kellie, S., Pattee, L., & Gay, J. (1990). Relationship of lung function to severity of osteoporosis in women. *American Review of Respiratory Disease*, 141(1), 68–71. <https://doi.org/10.1164/ajrccm/141.1.68>
83. Lehmann, B., & Meurer, M. (2003). Extrarenal sites of calcitriol synthesis: the particular role of the skin. *Recent Results in Cancer Research. Fortschritte Der Krebsforschung. Progrès Dans Les Recherches Sur Le Cancer*, 164, 135–145. https://doi.org/10.1007/978-3-642-55580-0_9
84. Levin, A., Bakris, G. L., Molitch, M., Smulders, M., Tian, J., Williams, L. A., & Andress, D. L. (2007). Prevalence of abnormal serum vitamin D, PTH, calcium, and phosphorus in patients with chronic kidney disease: Results of the study to evaluate early kidney disease. *Kidney International*, 71(1), 31–38. <https://doi.org/10.1038/sj.ki.5002009>
85. Li, Y. C. (2003). Vitamin D regulation of the renin-angiotensin system. *Journal of Cellular Biochemistry*, 88(2), 327–331. <https://doi.org/10.1002/jcb.10343>
86. Li, Y. C. (2010). Renoprotective effects of vitamin D analogs. *Kidney International*, 78(2), 134–139. <https://doi.org/10.1038/ki.2009.175>

87. Li, Y. C., Qiao, G., Uskokovic, M., Xiang, W., Zheng, W., & Kong, J. (2004). Vitamin D: A negative endocrine regulator of the renin-angiotensin system and blood pressure. *Journal of Steroid Biochemistry and Molecular Biology*, 89–90(1–5), 387–392. <https://doi.org/10.1016/j.jsbmb.2004.03.004>
88. Lips, P. (2001). Vitamin D deficiency and secondary hyperparathyroidism in the elderly: Consequences for bone loss and fractures and therapeutic implications. In *Endocrine Reviews* (Vol. 22, Issue 4, pp. 477–501). Endocrine Society. <https://doi.org/10.1210/edrv.22.4.0437>
89. LIPS, P., HOSKING, D., LIPPUNER, K., NORQUIST, J. M., WEHREN, L., MAALOUF, G., RAGI-EIS, S., & CHANDLER, J. (2006). The prevalence of vitamin D inadequacy amongst women with osteoporosis: an international epidemiological investigation. *Journal of Internal Medicine*, 260(3), 245–254. <https://doi.org/10.1111/j.1365-2796.2006.01685.x>
90. Litonjua, A. A., & Weiss, S. T. (2007). Is vitamin D deficiency to blame for the asthma epidemic? In *Journal of Allergy and Clinical Immunology* (Vol. 120, Issue 5, pp. 1031–1035). J Allergy Clin Immunol. <https://doi.org/10.1016/j.jaci.2007.08.028>
91. Lu, M., Taylor, B. v., & Körner, H. (2018). Genomic effects of the vitamin D receptor: Potentially the link between vitamin D, immune cells, and multiple sclerosis. In *Frontiers in Immunology* (Vol. 9, Issue MAR). Frontiers Media S.A. <https://doi.org/10.3389/fimmu.2018.00477>
92. Luxwolda, M. F., Kuipers, R. S., Kema, I. P., van der Veer, E., Dijck-Brouwer, D. A. J., & Muskiet, F. A. J. (2013). Vitamin D status indicators in indigenous populations in East Africa. *European Journal of Nutrition*, 52(3), 1115–1125. <https://doi.org/10.1007/s00394-012-0421-6>
93. MacLaughlin, J., & Holick, M. F. (1985). Aging decreases the capacity of human skin to produce vitamin D₃. *Journal of Clinical Investigation*, 76(4), 1536–1538. <https://doi.org/10.1172/JCI112134>
94. Majewski, S., Skopinska, M., Marczak, M., Szmurlo, A., Bollag, W., & Jablonska, S. (1996). Vitamin D₃ is a potent inhibitor of tumor cell-induced angiogenesis. *Journal of Investigative Dermatology Symposium Proceedings*, 1(1), 97–101. <https://europepmc.org/article/med/9627702>
95. Manousopoulou, A., Al-Daghri, N. M., Garbis, S. D., & Chrousos, G. P. (2015). Vitamin D and cardiovascular risk among adults with obesity: a systematic review and meta-

- analysis. *European Journal of Clinical Investigation*, 45(10), 1113–1126. <https://doi.org/10.1111/eci.12510>
96. Mantell, D. J., Owens, P. E., Bundred, N. J., Mawer, E. B., & Canfield, A. E. (2000). 1 α ,25-Dihydroxyvitamin D3 inhibits angiogenesis in vitro and in vivo. *Circulation Research*, 87(3), 214–220. <https://doi.org/10.1161/01.RES.87.3.214>
97. Marchesini, G. (2003). Nonalcoholic fatty liver, steatohepatitis, and the metabolic syndrome. *Hepatology*, 37(4), 917–923. <https://doi.org/10.1053/jhep.2003.50161>
98. Mathiasen, I. S., Lademann, U., & Jäätelä. (1999). Apoptosis Induced by Vitamin D Compounds in Breast Cancer Cells Is Inhibited by Bcl-2 but Does Not Involve Known Caspases or p531 . *Cancer Research*, 59(19), 4848–4856.
99. Matsuoka, L. Y., Ide, L., Wortsman, J., Maclaughlin, J. A., & Holick, M. F. (1987). Sunscreens Suppress Cutaneous Vitamin D3 Synthesis. *Journal of Clinical Endocrinology and Metabolism*, 64(6), 1165–1168. <https://doi.org/10.1210/jcem-64-6-1165>
100. Matsuoka, L. Y., Wortsman, J., Dannenberg, M. J., Hollis, B. W., Lu, Z., & Holick, M. F. (1992). Clothing prevents ultraviolet-B radiation-dependent photosynthesis of vitamin D3. *The Journal of Clinical Endocrinology & Metabolism*, 75(4), 1099–1103. <https://doi.org/10.1210/jcem.75.4.1328275>
101. Mccarron, D. A., Morris, C. D., & Bukoski, R. (1987). The calcium paradox of essential hypertension. *The American Journal of Medicine*, 82(1 SUPPL. 2), 27–33. [https://doi.org/10.1016/0002-9343\(87\)90268-3](https://doi.org/10.1016/0002-9343(87)90268-3)
102. McKenna, M. J. (1992). Differences in vitamin D status between countries in young adults and the elderly. In *The American Journal of Medicine* (Vol. 93, Issue 1, pp. 69–77). Elsevier. [https://doi.org/10.1016/0002-9343\(92\)90682-2](https://doi.org/10.1016/0002-9343(92)90682-2)
103. Melton, L. J., Atkinson, E. J., O'Connor, M. K., O'Fallon, W. M., & Riggs, B. L. (1998). Bone density and fracture risk in men. *Journal of Bone and Mineral Research*, 13(12), 1915–1923. <https://doi.org/10.1359/jbmr.1998.13.12.1915>
104. Melton, L. J., Chrischilles, E. A., Cooper, C., Lane, A. W., & Riggs, B. L. (1992). Perspective how many women have osteoporosis? *Journal of Bone and Mineral Research*, 7(9), 1005–1010. <https://doi.org/10.1002/jbmr.5650070902>
105. Mizoue, T. (2004). ECOLOGICAL STUDY OF SOLAR RADIATION AND CANCER MORTALITY IN JAPAN. *Health Physics*, 87(5), 532–538. <https://doi.org/10.1097/01.HP.0000137179.03423.0b>

106. Moreno, J., Krishnan, A. v., Swami, S., Nonn, L., Peehl, D. M., & Feldman, D. (2005). Regulation of prostaglandin metabolism by calcitriol attenuates growth stimulation in prostate cancer cells. *Cancer Research*, *65*(17), 7917–7925. <https://doi.org/10.1158/0008-5472.CAN-05-1435>
107. Munger, K. L., Levin, L. I., Hollis, B. W., Howard, N. S., & Ascherio, A. (2006). Serum 25-hydroxyvitamin D levels and risk of multiple sclerosis. *Journal of the American Medical Association*, *296*(23), 2832–2838. <https://doi.org/10.1001/jama.296.23.2832>
108. Munger, K. L., Zhang, S. M., O'Reilly, E., Hernán, M. A., Olek, M. J., Willett, W. C., & Ascherio, A. (2004). Vitamin D intake and incidence of multiple sclerosis. *Neurology*, *62*(1), 60–65. <https://doi.org/10.1212/01.WNL.0000101723.79681.38>
109. Nakagawa, K. (2004). 1,25-Dihydroxyvitamin D₃ is a preventive factor in the metastasis of lung cancer. *Carcinogenesis*, *26*(2), 429–440. <https://doi.org/10.1093/carcin/bgh332>
110. Newsholme, P., Cruzat, V. F., Keane, K. N., Carlessi, R., & de Bittencourt, P. I. H. (2016). Molecular mechanisms of ROS production and oxidative stress in diabetes. In *Biochemical Journal* (Vol. 473, Issue 24, pp. 4527–4550). Portland Press Ltd. <https://doi.org/10.1042/BCJ20160503C>
111. Noto, H., & Heller, H. J. (2009). Vitamin D Deficiency as an Ignored Cause of Hypocalcemia in Acute Illness: Report of 2 Cases and Review of Literature. *The Open Endocrinology Journal*, *3*(1), 1–4. <https://doi.org/10.2174/1874216500903010001>
112. Nuti, R., Siviero, P., Maggi, S., Guglielmi, G., Caffarelli, C., Crepaldi, G., & Gonnelli, S. (2009). Vertebral fractures in patients with chronic obstructive pulmonary disease: The EOLO Study. *Osteoporosis International*, *20*(6), 989–998. <https://doi.org/10.1007/s00198-008-0770-4>
113. Nykjaer, A., Dragun, D., Walther, D., Vorum, H., Jacobsen, C., Herz, J., Melsen, F., Christensen, E. I., & Willnow, T. E. (1999). An endocytic pathway essential for renal uptake and activation of the steroid 25-(OH) vitamin D₃. *Cell*, *96*(4), 507–515. [https://doi.org/10.1016/S0092-8674\(00\)80655-8](https://doi.org/10.1016/S0092-8674(00)80655-8)
114. O'Loughlin, P. D., & Morris, H. A. (1998). Oestrogen deficiency impairs intestinal calcium absorption in the rat. *Journal of Physiology*, *511*(1), 313–322. <https://doi.org/10.1111/j.1469-7793.1998.313bi.x>
115. Ordóñez-Morán, P., Larriba, M. J., Pálmer, H. G., Valero, R. A., Barbáchano, A., Duñach, M., de Herreros, A. G., Villalobos, C., Berciano, M. T., Lafarga, M., & Muñoz, A. (2008). RhoA - ROCK and p38MAPK-MSK1 mediate vitamin D effects on gene

- expression, phenotype, and Wnt pathway in colon cancer cells. *Journal of Cell Biology*, 183(4), 697–710. <https://doi.org/10.1083/jcb.200803020>
116. Ortlepp, J. R., Lauscher, J., Hoffmann, R., Hanrath, P., & Joost, H. G. (2001). The vitamin D receptor gene variant is associated with the prevalence of Type 2 diabetes mellitus and coronary artery disease. *Diabetic Medicine*, 18(10), 842–845. <https://doi.org/10.1046/j.1464-5491.2001.00585.x>
117. Ougolkov, A. v., & Billadeau, D. D. (2006). Targeting GSK-3: A promising approach for cancer therapy? In *Future Oncology* (Vol. 2, Issue 1, pp. 91–100). Future Medicine Ltd London, UK . <https://doi.org/10.2217/14796694.2.1.91>
118. Papi, A., Luppi, F., Franco, F., & Fabbri, L. M. (2006). Pathophysiology of exacerbations of chronic obstructive pulmonary disease. *Proceedings of the American Thoracic Society*, 3(3), 245–251. <https://doi.org/10.1513/pats.200512-125SF>
119. Parfitt, A. M. (2002). Targeted and nontargeted bone remodeling: Relationship to basic multicellular unit origination and progression. *Bone*, 30(1), 5–7. [https://doi.org/10.1016/S8756-3282\(01\)00642-1](https://doi.org/10.1016/S8756-3282(01)00642-1)
120. Paternostro, R., Wagner, D., Reiberger, T., Mandorfer, M., Schwarzer, R., Ferlitsch, M., Trauner, M., Peck-Radosavljevic, M., & Ferlitsch, A. (2017). Low 25-OH-vitamin D levels reflect hepatic dysfunction and are associated with mortality in patients with liver cirrhosis. *Wiener Klinische Wochenschrift*, 129(1–2), 8–15. <https://doi.org/10.1007/s00508-016-1127-1>
121. Pendás-Franco, N., García, J. M., Peña, C., Valle, N., Pálmer, H. G., Heinäniemi, M., Carlberg, C., Jiménez, B., Bonilla, F., Muñoz, A., & González-Sancho, J. M. (2008). DICKKOPF-4 is induced by TCF/ β -catenin and upregulated in human colon cancer, promotes tumour cell invasion and angiogenesis and is repressed by 1 α ,25-dihydroxyvitamin D₃. *Oncogene*, 27(32), 4467–4477. <https://doi.org/10.1038/onc.2008.88>
122. Petta, S., Cammà, C., Scazzone, C., Tripodo, C., Marco, V. di, Bono, A., Cabibi, D., Licata, G., Porcasi, R., Marchesini, G., & Craxí, A. (2010). Low vitamin d serum level is related to severe fibrosis and low responsiveness to interferon-based therapy in genotype 1 chronic hepatitis C. *Hepatology*, 51(4), 1158–1167. <https://doi.org/10.1002/hep.23489>
123. Pietschmann, P., Schernthaner, G., & Woloszczuk, W. (1988). Serum osteocalcin levels in diabetes mellitus: analysis of the type of diabetes and microvascular complications. *Diabetologia*, 31(12), 892–895. <https://doi.org/10.1007/BF00265373>

124. Polat, H. B., & Beyazal, M. S. (2018). The effect of cholecystectomy on 25-hydroxyvitamin D levels and bone mineral density in postmenopausal women. *Archives of Osteoporosis*, *13*(1). <https://doi.org/10.1007/s11657-018-0458-0>
125. Poon, A. H., Laprise, C., Lemire, M., Montpetit, A., Sinnett, D., Schurr, E., & Hudson, T. J. (2004). Association of vitamin D receptor genetic variants with susceptibility to asthma and atopy. *American Journal of Respiratory and Critical Care Medicine*, *170*(9), 967–973. <https://doi.org/10.1164/rccm.200403-412OC>
126. Prestwood, K. M., Pannullo, A. M., Kenny, A. M., Pilbeam, C. C., & Raisz, L. G. (1996). The effect of a short course of calcium and vitamin D on bone turnover in older women. *Osteoporosis International*, *6*(4), 314–319. <https://doi.org/10.1007/BF01623391>
127. Reichel, H., Koeffler, H. P., & Norman, A. W. (1989). The Role of the Vitamin D Endocrine System in Health and Disease. *New England Journal of Medicine*, *320*(15), 980–991. <https://doi.org/10.1056/NEJM198904133201506>
128. Resnick, L. M., Muller, F. B., & Laragh, J. H. (1986). Calcium-regulating hormones in essential hypertension: Relation to plasma renin activity and sodium metabolism. *Annals of Internal Medicine*, *105*(5), 649–654. <https://doi.org/10.7326/0003-4819-105-5-649>
129. Rostand, S. G. (1997). Ultraviolet light may contribute to geographic and racial blood pressure differences. *Hypertension*, *30*(2), 150–156. <https://doi.org/10.1161/01.HYP.30.2.150>
130. Runia, T. F., Hop, W. C. J., de Rijke, Y. B., Buljevac, D., & Hintzen, R. Q. (2012). Lower serum vitamin D levels are associated with a higher relapse risk in multiple sclerosis. *Neurology*, *79*(3), 261–266. <https://doi.org/10.1212/WNL.0b013e31825fdec7>
131. Salekzamani, S., Babil, A. S., Mehralizadeh, H., Jafarabadi, M. A., Ghezeli, A., & Gargari, B. P. (2017). The effects of vitamin D supplementation on proatherogenic inflammatory markers and carotid intima media thickness in subjects with metabolic syndrome: a randomized double-blind placebo-controlled clinical trial. *Endocrine*, *57*(1), 51–59. <https://doi.org/10.1007/s12020-017-1317-2>
132. Salzer, J., Hallmans, G., Nyström, M., Stenlund, H., Wadell, G., & Sundström, P. (2012). Vitamin D as a protective factor in multiple sclerosis. *Neurology*, *79*(21), 2140–2145. <https://doi.org/10.1212/WNL.0b013e3182752ea8>
133. Schwartz, G. G., Eads, D., Rao, A., Cramer, S. D., Willingham, M. C., Chen, T. C., Jamieson, D. P., Wang, L., Burnstein, K. L., Holick, M. F., & Koumenis, C. (2004). Pancreatic cancer cells express 25-hydroxyvitamin D-1 α -hydroxylase and their

- proliferation is inhibited by the prohormone 25-hydroxyvitamin D3. *Carcinogenesis*, 25(6), 1015–1026. <https://doi.org/10.1093/carcin/bgh086>
134. Scragg, R. (1981). Seasonality of cardiovascular disease mortality and the possible protective effect of ultra-violet radiation. *International Journal of Epidemiology*, 10(4), 337–341. <https://doi.org/10.1093/ije/10.4.337>
135. Scragg, R., Holdaway, I., Singh, V., Metcalf, P., Baker, J., & Dryson, E. (1995). Serum 25-hydroxyvitamin D3 levels decreased in impaired glucose tolerance and diabetes mellitus. *Diabetes Research and Clinical Practice*, 27(3), 181–188. [https://doi.org/10.1016/0168-8227\(95\)01040-K](https://doi.org/10.1016/0168-8227(95)01040-K)
136. Sergeev, I. N. (2016). 1,25-DihydroxyVitamin D3 and type 2 diabetes: Ca²⁺-dependent molecular mechanisms and the role of Vitamin D status. In *Hormone Molecular Biology and Clinical Investigation* (Vol. 26, Issue 1, pp. 61–65). Walter de Gruyter GmbH. <https://doi.org/10.1515/hmbci-2015-0069>
137. Simpson R U, Thomas G A, & Arnold A J. (1985). Identification of 1,25-dihydroxyvitamin D3 receptors and activities in muscle - PubMed. *J Biol Chem*, 260(15), 8882–8891. <https://pubmed.ncbi.nlm.nih.gov/2991224/>
138. Simpson, S., Taylor, B., Blizzard, L., Ponsonby, A. L., Pittas, F., Tremlett, H., Dwyer, T., Gies, P., & van der Mei, I. (2010). Higher 25-hydroxyvitamin D is associated with lower relapse risk in multiple sclerosis. *Annals of Neurology*, 68(2), 193–203. <https://doi.org/10.1002/ana.22043>
139. Spina Catherine Sheridan. (2005). *Novel Vitamin D3 Analogues and Colon Cancer Growth in Mice*. Boston University.
140. Stene, L. C., Ulriksen, J., Magnus, P., & Joner, G. (2000). Use of cod liver oil during pregnancy associated with lower risk of Type I diabetes in the offspring. *Diabetologia*, 43(9), 1093–1098. <https://doi.org/10.1007/s001250051499>
141. Stoker, G. E., Buchowski, J. M., & Stoker, M. E. (2012). Prior cholecystectomy as a predictor of preoperative vitamin D deficiency in adults undergoing spine surgery. In *Archives of Surgery* (Vol. 147, Issue 6, pp. 577–578). American Medical Association. <https://doi.org/10.1001/archsurg.2012.463>
142. Sun, J., Kong, J., Duan, Y., Szeto, F. L., Liao, A., Madara, J. L., & Yan, C. L. (2006). Increased NF-κB activity in fibroblasts lacking the vitamin D receptor. *American Journal of Physiology - Endocrinology and Metabolism*, 291(2). <https://doi.org/10.1152/ajpendo.00590.2005>

143. Surdu, A. M., Pinzariu, O., Ciobanu, D. M., Negru, A. G., Căinap, S. S., Lazea, C., Iacob, D., Săraci, G., Tirinescu, D., Borda, I. M., & Cismaru, G. (2021). Vitamin D and its role in the lipid metabolism and the development of atherosclerosis. In *Biomedicines* (Vol. 9, Issue 2, pp. 1–16). MDPI AG. <https://doi.org/10.3390/biomedicines9020172>
144. Sutherland, E. R., Goleva, E., Jackson, L. P., Stevens, A. D., & Leung, D. Y. M. (2010). Vitamin D levels, lung function, and steroid response in adult asthma. *American Journal of Respiratory and Critical Care Medicine*, *181*(7), 699–704. <https://doi.org/10.1164/rccm.200911-1710OC>
145. Targher, G., Bertolini, L., Scala, L., Cigolini, M., Zenari, L., Falezza, G., & Arcaro, G. (2007). Associations between serum 25-hydroxyvitamin D3 concentrations and liver histology in patients with non-alcoholic fatty liver disease. *Nutrition, Metabolism and Cardiovascular Diseases*, *17*(7), 517–524. <https://doi.org/10.1016/j.numecd.2006.04.002>
146. The EURODIAB Substudy 2 Study Group. (1999). Vitamin D supplement in early childhood and risk for Type I (insulin-dependent) diabetes mellitus. *Diabetologia*, *42*, 51–54.
147. Thompson, W. G., & Thompson, G. R. (1969). Effect of cholestyramine on the absorption of vitamin D3 and calcium. *Gut*, *10*(9), 717–722. <https://doi.org/10.1136/gut.10.9.717>
148. Tilg, H., & Moschen, A. R. (2010). Evolution of inflammation in nonalcoholic fatty liver disease: The multiple parallel hits hypothesis. *Hepatology*, *52*(5), 1836–1846. <https://doi.org/10.1002/hep.24001>
149. Uhmann, A., Niemann, H., Lammering, B., Henkel, C., Heß, I., Nitzki, F., Fritsch, A., Prüfer, N., Rosenberger, A., Dullin, C., Schraepfer, A., Reifenberger, J., Schweyer, S., Pietsch, T., Strutz, F., Schulz-Schaeffer, W., & Hahn, H. (2011). Antitumoral effects of calcitriol in basal cell carcinomas involve inhibition of hedgehog signaling and induction of vitamin D receptor signaling and differentiation. *Molecular Cancer Therapeutics*, *10*(11), 2179–2188. <https://doi.org/10.1158/1535-7163.MCT-11-0422>
150. Underwood, J. L., & DeLuca, H. F. (1984). Vitamin D is not directly necessary for bone growth and mineralization. *American Journal of Physiology - Endocrinology and Metabolism*, *9*(6). <https://doi.org/10.1152/ajpendo.1984.246.6.e493>
151. Urashima, M., Segawa, T., Okazaki, M., Kurihara, M., Wada, Y., & Ida, H. (2010). Randomized trial of vitamin D supplementation to prevent seasonal influenza A in schoolchildren. *American Journal of Clinical Nutrition*, *91*(5), 1255–1260. <https://doi.org/10.3945/ajcn.2009.29094>

152. Uribarri, J. (2007). PHOSPHORUS METABOLISM AND MANAGEMENT IN CHRONIC KIDNEY DISEASE: Expanding Role for Vitamin D in Chronic Kidney Disease: Importance of Blood 25-OH-D Levels and Extra-Renal 1 α -Hydroxylase in the Classical and Nonclassical Actions of 1 α ,25-Dihydroxyvitamin D. *Seminars in Dialysis*.
153. Vanoirbeek, E., Krishnan, A., Eelen, G., Verlinden, L., Bouillon, R., Feldman, D., & Verstuyf, A. (2011). The anti-cancer and anti-inflammatory actions of 1,25(OH)₂D₃. In *Best Practice and Research: Clinical Endocrinology and Metabolism* (Vol. 25, Issue 4, pp. 593–604). Bailliere Tindall Ltd. <https://doi.org/10.1016/j.beem.2011.05.001>
154. Vieth, R. (2020). Vitamin D supplementation: cholecalciferol, calcifediol, and calcitriol. In *European Journal of Clinical Nutrition* (Vol. 74, Issue 11, pp. 1493–1497). Springer Nature. <https://doi.org/10.1038/s41430-020-0697-1>
155. von Hurst, P. R., Stonehouse, W., & Coad, J. (2010). Vitamin D supplementation reduces insulin resistance in South Asian women living in New Zealand who are insulin resistant and vitamin D deficient—a randomised, placebo-controlled trial. *British Journal of Nutrition*, 103(4), 549–555. <https://doi.org/10.1017/S0007114509992017>
156. Vuolo, L., di Somma, C., Faggiano, A., & Colao, A. (2012). Vitamin D and cancer. In *Frontiers in Endocrinology* (Vol. 3, Issue APR). Front Endocrinol (Lausanne). <https://doi.org/10.3389/fendo.2012.00058>
157. Wacker, M., & Holick, M. F. (2013). Sunlight and Vitamin D: A global perspective for health. In *Dermato-Endocrinology* (Vol. 5, Issue 1, pp. 51–108). Landes Bioscience. <https://doi.org/10.4161/derm.24494>
158. Webb, A. R., Kline, L., & Holick, M. F. (1988). Influence of Season and Latitude on the Cutaneous Synthesis of Vitamin D₃: Exposure to Winter Sunlight in Boston and Edmonton Will Not Promote Vitamin D₃ Synthesis in Human Skin. *Journal of Clinical Endocrinology and Metabolism*, 67(2), 373–378. <https://doi.org/10.1210/jcem-67-2-373>
159. Welsh, J. (1994). Induction of apoptosis in breast cancer cells in response to vitamin D and antiestrogens. In *Biochemistry and cell biology = Biochimie et biologie cellulaire* (Vol. 72, Issues 11–12, pp. 537–545). NRC Research Press Ottawa, Canada . <https://doi.org/10.1139/o94-072>
160. Welsh, J., VanWeelden, K., Flanagan, L., Byrne, I., Nolan, E., & Narvaez, C. J. (1998). The role of vitamin D₃ and antiestrogens in modulating apoptosis of breast cancer cells and tumors. In *Sub-cellular biochemistry* (Vol. 30, pp. 245–270). Springer, Boston, MA. https://doi.org/10.1007/978-1-4899-1789-8_11

161. Williams, C. E., Williams, E. A., & Corfe, B. M. (2018). Vitamin D status in irritable bowel syndrome and the impact of supplementation on symptoms: what do we know and what do we need to know? In *European Journal of Clinical Nutrition* (Vol. 72, Issue 10, pp. 1358–1363). Nature Publishing Group. <https://doi.org/10.1038/s41430-017-0064-z>
162. Williams, S., Malatesta, K., & Norris, K. (2009). Vitamin D and chronic kidney disease. In *Ethnicity & disease* (Vol. 19, Issue 4 Suppl 5, p. S5). NIH Public Access. [/pmc/articles/PMC2878736/](https://pubmed.ncbi.nlm.nih.gov/19122221/)
163. Wimalawansa, S. J. (2018). Associations of vitamin D with insulin resistance, obesity, type 2 diabetes, and metabolic syndrome. In *Journal of Steroid Biochemistry and Molecular Biology* (Vol. 175, pp. 177–189). Elsevier Ltd. <https://doi.org/10.1016/j.jsbmb.2016.09.017>
164. Wolf, D., & Ley, K. (2019). Immunity and Inflammation in Atherosclerosis. In *Circulation Research* (Vol. 124, Issue 2, pp. 315–327). Lippincott Williams and Wilkins. <https://doi.org/10.1161/CIRCRESAHA.118.313591>
165. World Health Organization. (2021). *Noncommunicable diseases*. <https://www.who.int/news-room/fact-sheets/detail/noncommunicable-diseases>
166. Xie, S. P., Pirianov, G., & Colston, K. W. (1999). Vitamin D analogues suppress IGF-I signalling and promote apoptosis in breast cancer cells. *European Journal of Cancer*, 35(12), 1717–1723. [https://doi.org/10.1016/S0959-8049\(99\)00200-2](https://doi.org/10.1016/S0959-8049(99)00200-2)
167. Xystrakis, E., Kusumakar, S., Boswell, S., Peek, E., Urry, Z., Richards, D. F., Adikibi, T., Pridgeon, C., Dallman, M., Loke, T. K., Robinson, D. S., Barrat, F. J., O’Garra, A., Lavender, P., Lee, T. H., Corrigan, C., & Hawrylowicz, C. M. (2006). Reversing the defective induction of IL-10-secreting regulatory T cells in glucocorticoid-resistant asthma patients. *Journal of Clinical Investigation*, 116(1), 146–155. <https://doi.org/10.1172/JCI21759>
168. Zaidi, M. (2007). Skeletal remodeling in health and disease. In *Nature Medicine* (Vol. 13, Issue 7, pp. 791–801). Nature Publishing Group. <https://doi.org/10.1038/nm1593>
169. Zhang, Y., Leung, D. Y. M., Richers, B. N., Liu, Y., Remigio, L. K., Riches, D. W., & Goleva, E. (2012). Vitamin D Inhibits Monocyte/Macrophage Proinflammatory Cytokine Production by Targeting MAPK Phosphatase-1. *The Journal of Immunology*, 188(5), 2127–2135. <https://doi.org/10.4049/jimmunol.1102412>
170. Zhernakova, N. I., Bunova, S. S., Agarkov, N. M., Lebedev, D. T., & Aksenov, V. v. (2021). Vitamin D deficiency as an independent predictor of myocardial infarction in the

elderly. *Archives of Razi Institute*, 76(4), 937–944.
<https://doi.org/10.22092/ARI.2021.356047.1766>

171. Zhou, J. Y., Norman, A. W., Chen, D. L., Sun, G. W., Uskokovic, M., & Phillip Koeffler, H. (1990). 1,25-dihydroxy-16-ene-23-yne-vitamin D₃ prolongs survival time of leukemic mice. *Proceedings of the National Academy of Sciences of the United States of America*, 87(10), 3929–3932. <https://doi.org/10.1073/pnas.87.10.3929>
172. Zittermann, A. (2006). Vitamin D and disease prevention with special reference to cardiovascular disease. In *Progress in Biophysics and Molecular Biology* (Vol. 92, Issue 1, pp. 39–48). Elsevier Ltd. <https://doi.org/10.1016/j.pbiomolbio.2006.02.001>