

**A Review on Association of Mothers' Single Nucleotide
Polymorphism (BsmI) of Vitamin D Receptor Gene with Risk
of Preterm Birth and Low Birth Weight**

By

Hasan Shahriyer Tonmoy
Student ID: 19146019

A thesis submitted to the School of Pharmacy in partial fulfillment of the requirements
for the degree of
Bachelor of Pharmacy (Hons.)

School of Pharmacy
Brac University
September 2023

© 2023. Brac University
All rights reserved.

Declaration

It is hereby declared that

1. The thesis submitted is my own original work while completing degree at Brac University.
2. The thesis does not contain material previously published or written by a third party, except where this is appropriately cited through full and accurate referencing.
3. The thesis does not contain material which has been accepted, or submitted, for any other degree or diploma at a university or other institution.
4. I have acknowledged all main sources of help.

Student's Full Name & Signature:



Hasan Shahriyer Tonmoy
19146019

Approval

The thesis titled “A Review on Association of Mothers’ Single Nucleotide Polymorphism (BsmI) of Vitamin D Receptor Gene with Risk of Preterm Birth and Low Birth Weight” submitted by Hasan Shahriyer Tonmoy (19146019), of Summer 2022 has been accepted as satisfactory in partial fulfillment of the requirement for the degree of Bachelor of Pharmacy (Hons.) on September 2023.

Supervised By:

Dr. Raushanara Akter
Professor
School of Pharmacy
BRAC University

Approved By:

Program Director:

Professor Dr. Hasina Yasmin
Program Director and Assistant Dean
School of Pharmacy
BRAC University

Dean:

Professor Dr. Eva Rahman Kabir
Dean
School of Pharmacy
BRAC University

Ethics Statement

This study does not involve any animal or human trial.

Abstract

Preterm birth and low birth weight are linked to increased neonatal morbidity and mortality and can harm a child's health and development. An expanding corpus of research confirms that a number of genetic and immunological variables contribute to premature delivery and low birth weight. Vitamin D can have substantial impact on pregnancy and birth outcomes by influencing calcium balance, immunity, and cell proliferation. Its deficiency in pregnant women can lead to adverse results. The vitamin D receptor (VDR) is crucial to the endocrine system, regulating gene expression alongside encoding the VDR protein. This review explored the impact of the maternal vitamin D receptor gene's single nucleotide polymorphism, BsmI on VDR activity and their associations with preterm delivery and birth weight. The findings of this review revealed a correlation between the VDR gene polymorphism BsmI and preterm birth, whereas, no connection was observed for low birth weight.

Keywords: pregnancy; vitamin D; vitamin D receptor; BsmI polymorphism; gestational duration; preterm birth; birth weight.

Dedication

This work is dedicated to my parents, whose unrelenting encouragement and assistance have been the propelling force behind my academic trajectory. Their trust in me and my abilities has inspired me to pursue my passions and chase my dreams. This research paper is a humble demonstration of my appreciation for their affection, guidance, and sacrifices.

Acknowledgement

I want to start by expressing my sincere appreciation to Allah Ta'ala for giving me the fortitude, insight, and direction necessary to complete this paper.

I am thankful to the Almighty for granting me the good fortune of having a kindhearted and supportive personality like Dr. Raushanara Akter (Professor of the School of Pharmacy, Brac University) as my project supervisor. I would like to extend my sincere appreciation to her for her invaluable direction, mentoring, and unwavering support during this transformative journey. Her commitment and expertise have significantly shaped my understanding and approach to research. I would also like to thank Professor Dr. Eva Rahman Kabir (Dean of the School of Pharmacy, Brac University) and Professor Dr. Hasina Yasmin (Program Director and Assistant Dean of the School of Pharmacy, Brac University) for allowing to work on this project along with their constant support and guidance throughout the process.

My profound gratitude goes out to all of my teachers, who have influenced me to follow my aspirations and taught me a lot of valuable things. In addition, it is essential to express gratitude for my colleagues and acquaintances, whose unshakable presence has served as a continual wellspring of motivation and assistance.

Last but not least, I would want to convey my utmost appreciation to my parents, whose unflinching presence and unwavering support have always accompanied me throughout my many pursuits. Their encouragement, sacrifices, and love have made this journey possible.

The project's successful completion would not have been feasible without the irreplaceable support and guidance provided by each of these people. I express my enduring gratitude for their significant contributions.

Table of Contents

Content	Page no.
Declaration.....	ii
Approval	iii
Ethics Statement.....	iv
Abstract.....	v
Dedication	vi
Acknowledgement	vii
Table of Contents	viii
List of Tables	xi
List of Figures.....	xii
List of Acronyms	xiii
Chapter 1: Introduction	1
1.1 An Overview on Preterm Birth and Low Birth Weight.....	1
1.2 Prevalence of Preterm Birth and Low Birth Weight.....	2
1.3 Vitamin D and its Biological Functions.....	3
1.4 Sources of Vitamin D.....	7
1.5 Synthesis and Metabolism of Vitamin D	8
1.6 Vitamin D Deficiency and its Complications	9
1.7 Rationale of the Study.....	12
1.8 Aim and Objectives of the Study	12

Chapter 2: Methodology.....	13
Chapter 3: Vitamin D and Pregnancy	14
3.1 Significance of Vitamin D for Maternal and Child Health.....	14
3.2 Vitamin D Requirements for Pregnant Women.....	16
3.3 Prevalence of Vitamin D Deficiency among Pregnant Women	16
3.4 Unfavorable Pregnancy Outcomes linked to Vitamin D deficiency.....	16
Chapter 4: Pathophysiology of Preterm Birth and Low Birth Weight.....	21
4.1 Risk Factors for Preterm Birth.....	21
4.2 Complications of Preterm Birth	23
4.3 Risk Factors for Low Birth Weight	24
4.4 Complications of Low Birth Weight.....	25
Chapter 5: Vitamin D Receptor Gene Polymorphisms	27
5.1 Vitamin D Receptor and its Location	27
5.2 How does Vitamin D Receptor Work?	27
5.3 Signaling by VDRs during Preterm Birth.....	28
5.4 Gene Polymorphism and their Types.....	29
5.5 Single Nucleotide Polymorphism of VDR Genes	30
5.6 VDR BsmI Gene Polymorphism	31
5.7 VDR Gene Polymorphisms and Adverse Pregnancy outcomes	31
Chapter 6: Findings on Association of Single Nucleotide VDR Gene Polymorphism BsmI with Preterm Birth & Low Birth Weight	33
6.1 Interrelation between VDR BsmI SNP & Preterm Birth.....	33

6.2 Interrelation between VDR BsmI SNP & Low Birth Weight.....	37
6.3 Limitations of the Findings.....	38
Chapter 7: Conclusion and Future Recommendations	39
Conclusion	39
Future Recommendations	40
References	41

List of Tables

Table 1: Common Dietary Sources of Vitamin D	7
Table 2: Major Complications in Infants with Extremely Low Birth Weight.....	25
Table 3: Studies on VDR gene variations and preterm birth risk	35

List of Figures

Figure 1: The 10 countries with the highest total number of PTBs in 2010.....	2
Figure 2: Top 10 countries with the greatest PTB per 100 live births rates in 2010	3
Figure 3: Structures of vitamin D analogues	4
Figure 4: Biological functions of vitamin D	5
Figure 5: Cutaneous Vitamin D synthesis and Metabolism.....	9
Figure 6: Schematic depiction of pregnancy factors and physiological alterations that promote optimal bone health in offspring	15
Figure 7: Risk factors associated with preterm birth	22
Figure 8: VDR protein's function in regulating gene expression.....	28
Figure 9: Common SNPs of the VDR gene	30

List of Acronyms

7DHC	7-dehydrocholesterol
25(OH)D	25-hydroxyvitamin D
APR	Acute-phase Response
BPD	Bronchopulmonary Dysplasia
BV	Bacterial Vaginosis
CAMP	cathelicidin
CRP	C-reactive protein
DBP	D Binding Protein
DV	Daily Value
EC	Endocrine Society
eNOS	Endothelial Nitric Oxide Synthase
FGF23	Fibroblast Growth Factor 23
FTN	Full-term Newborns
GDM	Gestational diabetes mellitus
GMCSF-2	Granulocyte Macrophage Colony Stimulating Factor-2
HCG	Human Chorionic Gonadotropin
HGVS	Human Genome Variation Society
IOM	Institute of Medicine

IU	International Unit
IVH	Intraventricular Hemorrhage
LPS	Lipopolysaccharides
NEC	Necrotizing Enterocolitis
NF- κ B	Nuclear Factor-kappa B
OR	Odds Ratio
PPROM	Premature Rupture of Membranes
PTB	Preterm Birth
PTH	Parathyroid Hormone
PTN	Preterm Neonates
LBW	Low Birth Weight
RDA	Recommended Dietary Allowance
RDS	Respiratory Distress Syndrome
RXR	Retinoid X Receptor
SGA	Small for Gestational Age
SNP	Single Nucleotide Polymorphism
SNV	Single Nucleotide Variation
T1DM	Type 1 Diabetes Mellitus
T2DM	Type 2 Diabetes Mellitus

TLR4	Toll-like Receptor 4
TNF	Tumor Necrosis Factor
UVB	Ultraviolet B
VDBP	Vitamin D Binding Protein
VDD	Vitamin D Deficiency
VDR	Vitamin D Receptor

Chapter 1: Introduction

1.1 An Overview on Preterm Birth and Low Birth Weight

Despite revolutionary advancements in healthcare technology, unintended pregnancy outcomes still pose significant challenges to the public's health (Chiavaroli et al., 2016; Goldenberg et al., 2008; Lawn et al., 2011). Preterm birth (PTB), which is characterized as birth prior to the 37th week of pregnancy, remains the leading reason for death in newborns and the secondary leading cause in under-five children (Blencowe et al., 2012; WHO, 1970). Preterm birth has immediate effects within the first month of life and lasting consequences for infants, families, and the community. Global Burden of Disease studies reveal that preterm birth contributes to 3.1% of lifelong disabilities, surpassing the combined impact of HIV and malaria (Howson et al., 2013; Murray et al., 2012). Premature birth exposes infants to risks like low blood sugar, sepsis, jaundice, respiratory issues, and long-term neurocognitive deficits, posing significant medical, social, and economic challenges (Goldenberg et al., 2008; Hall & Greenberg, 2016; S. W. Wen et al., 2004). In addition to PTB, low birth weight (LBW), labelled as a neonate weighing below 2500g, is a prevalent cause of both immediate and later childhood illness and perinatal death. Low-birth-weight babies have up to 40 times greater mortality rates and are more likely to have long-term problems (Goldenberg & Culhane, 2007). The exact causes of PTB and LBW remain complex and multifactorial, involving genetics, stress, inflammation, infections, environmental or occupational factors, along with pregnant women's nutritional status (Crider et al., 2005; Hendler et al., 2005; Kramer et al., 2009; Lawson et al., 2009; Michalowicz et al., 2009; Scholl, 2005; Tamura et al., 1992). Nowadays, a substantial amount of research is being undertaken how vitamin D metabolism affects pregnancy, specifically preterm birth (Møller et al., 2013; Qin et al., 2016).

1.2 Prevalence of Preterm Birth and Low Birth Weight

Despite consistent efforts in antenatal care, preterm birth persists as a significant global health problem. The estimated worldwide preterm birth rate is 11.1%, leading to about 15 million preterm babies in 2010 across 184 countries (Blencowe et al., 2012; Tielsch, 2015). This prevalence varies among regions, with Africa and South Asia contributing 60% of preterm births. Developed countries also grapple with this issue, showing a prevalence of roughly 6-7%, while the United States has the highest rate at 10% (Sheikh et al., 2016). Detailed data on countries with noteworthy preterm birth counts and rates is available in Figure 1 and 2.

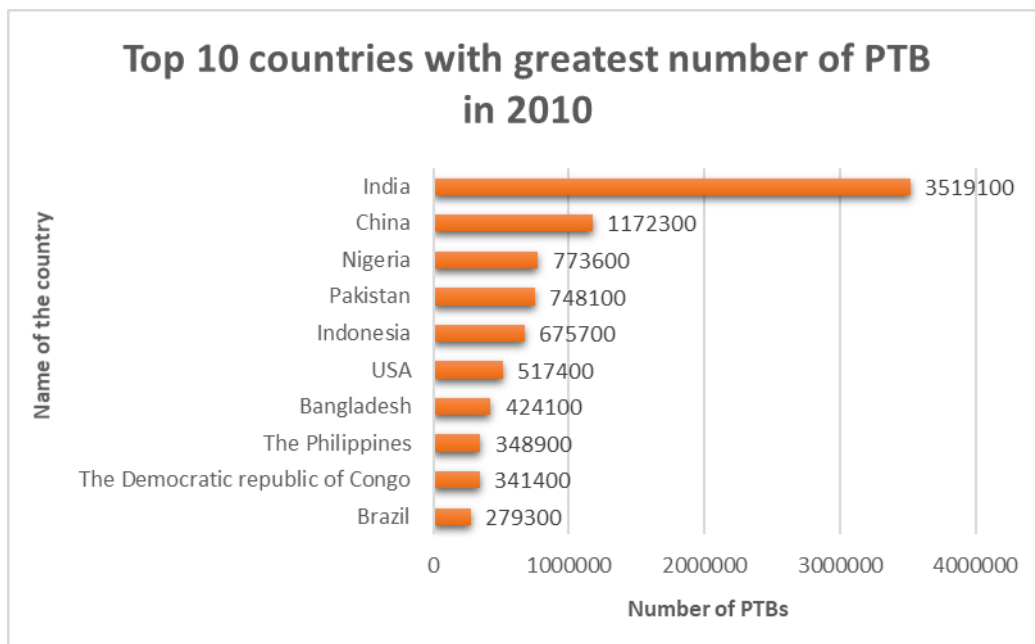


Figure 1: The 10 countries with the highest total number of PTBs in 2010 (Blencowe et al., 2012)

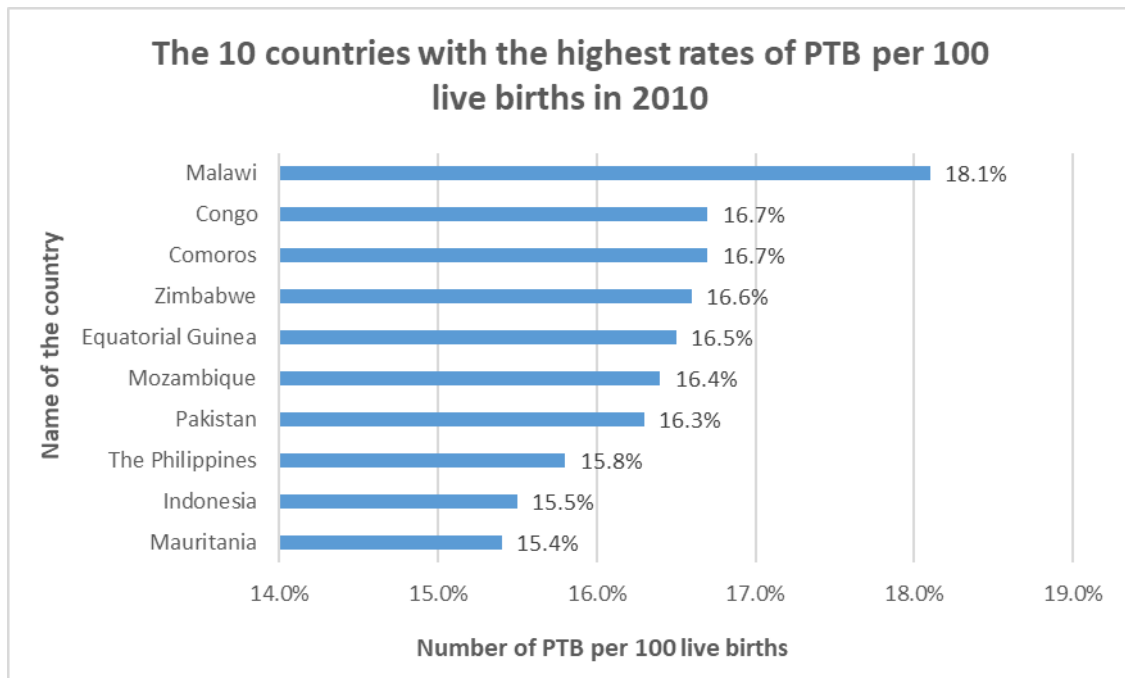
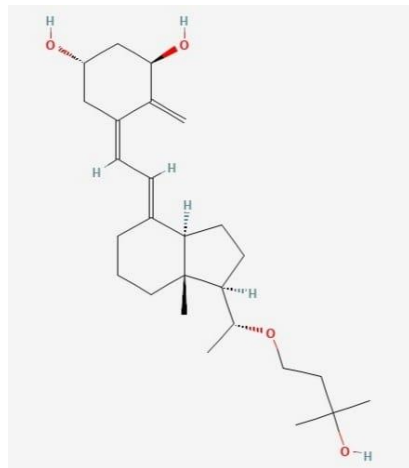


Figure 2: Top 10 countries with the greatest PTB per 100 live births rates in 2010 (Blencowe et al., 2012)

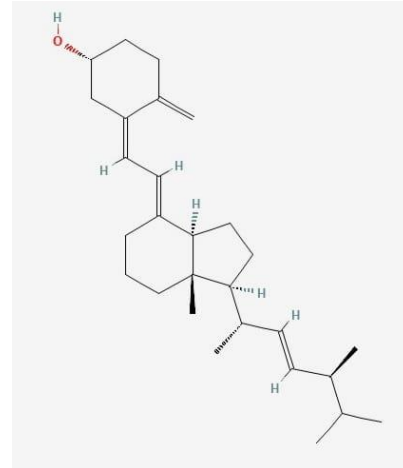
Approximately 32 million babies are born yearly with low birth weight (LBW) or small-for-gestational-age (SGA), mostly, 96.5%, in developing nations (Black, 2015). Racial disparities are apparent, with a 13.8% LBW rate for non-Hispanic Black (NHB) women in 2007 versus 7.2% for non-NHW women (Martin et al., 2001). Bangladesh shows a low birth weight prevalence of 23% to 60% (Yasmin et al., 2001).

1.3 Vitamin D and its Biological Functions

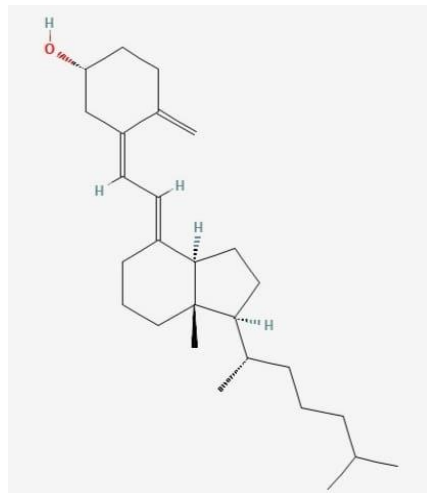
The fat-soluble prohormone group, vitamin D, was initially identified in cod liver oils for its anti-rachitic properties in the early 1900s (Zhang & Naughton, 2010). This hormone, with a history spanning 750 million years, has been generated by a range of organisms. Sunlight exposure triggers its synthesis in plants, animals, phytoplankton, and zooplankton (Holick, 2003). Vitamin D has two primary precursors: plant-based vitamin D₂ (Ergocalciferol) and skin-produced vitamin D₃ (Cholecalciferol) in response to ultraviolet B (UVB) radiation exposure (Holick, 2003; Zhang & Naughton, 2010).



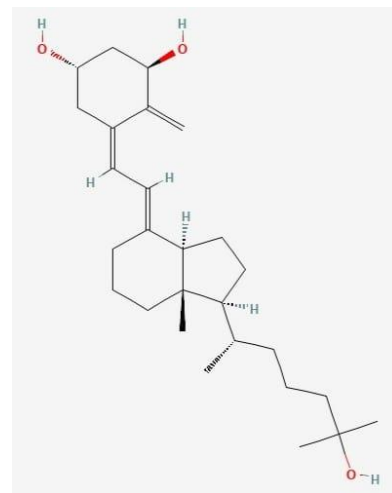
Maxacalcitol



Ergocalciferol



Cholecalciferol



Calcitriol

Figure 3: Structures of vitamin D analogues (PubChem, 2019, 2023b, 2023c, 2023a)

The vitamin D hormone is critical for sustaining proper plasma calcium levels, preventing conditions like rickets and osteomalacia. It achieves this through 3 key mechanisms:

- Firstly, it stimulates calcium absorption proteins in the intestines (Underwood & DeLuca, 1984).
- Calcium deficiency triggers the production of Receptor activator of nuclear factor kappa-B ligand (RANKL) by osteoblasts, consequently promoting the development of osteoclasts and

loss of bone mass through parathyroid hormone (PTH) and vitamin D (Garabedian et al., 1972; GARABEDIAN et al., 1974; Suda et al., 2003).

- Additionally, it assists in the final 1% calcium reabsorption in the renal tubule, working with PTH (Yamamoto et al., 1984).

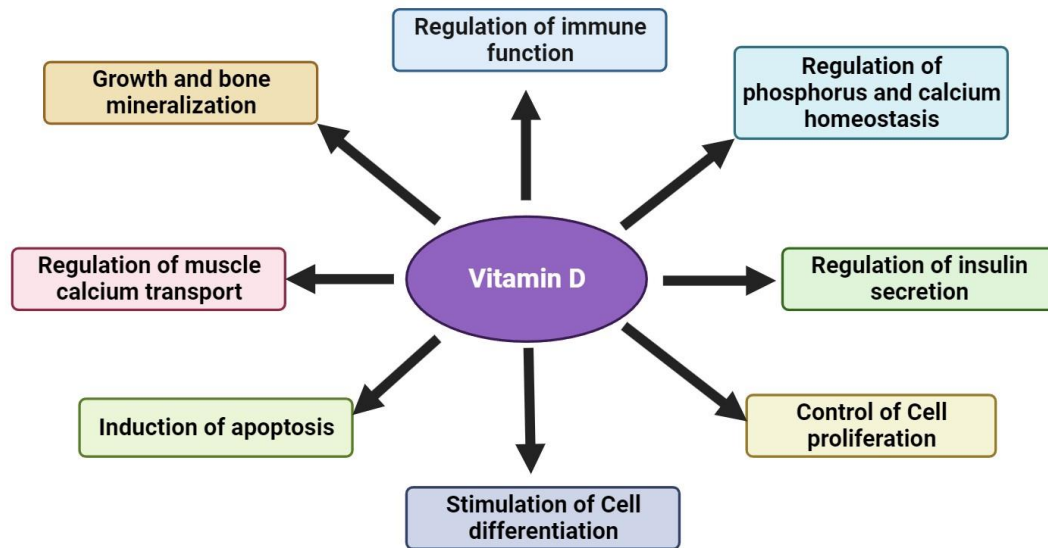


Figure 4: Biological functions of vitamin D (Layana et al., 2017; Scientific Image and Illustration Software, 2023)

Calcium absorption and increasing uptake by enterocytes rely heavily on PTH along with the vitamin D hormone. Stable plasma calcium levels trigger PTH secretion, enhancing vitamin D hormone release, which mobilizes bone calcium and is influenced by the parathyroid gland's calcium sensing (Brenza & DeLuca, 2000; Brown et al., 1993; Tanaka & DeLuca, 1984). This dynamic process improves calcium absorption, bone mobilization, and kidney reabsorption. Excess calcium prompts the C cells of thyroid gland to produce calcitonin, an inhibitor of bone calcium release (JONES et al., 1998), stimulating renal 1α -hydroxylase and generating non-calcemic vitamin D. The vitamin D system prioritizes normal serum calcium via dietary sources (Shinki et al., 1999). In calcium deficiency, it intensifies bone mobilization, kidney

reabsorption, and may contribute to osteoporosis. The vitamin D hormone primarily supports skeletal mineralization, not direct bone anabolism (DeLuca, 2004).

Vitamin D expands its functions beyond bone by aiding the promyelocytes differentiate into monocytes, the progenitors of giant osteoclasts. Through the RANKL system, this essential process helps vitamin D trigger osteoclast formation. Upon differentiation into a functioning cell line, cell growth remarkably halts, and this mechanism is unrelated to calcium or phosphorus (Suda et al., 2003).

Parathyroid gland's vitamin D receptor is crucial for renal osteodystrophy treatment using vitamin D hormone and analogs. It regulates gene production and inhibits cell growth, aiding parathyroid health in healthy individuals and dialysis patients (Darwish & DeLuca, 1999; DeLuca, 2004; JONES et al., 1998; Slatopolsky et al., 2003).

Vitamin D modulates the renin-angiotensin system (RAS), reducing hypertension as well as heart disease risk (Akter et al., 2022; Wu-Wong et al., 2007).

Vitamin D positively impacts pancreatic beta cell function. Experiments on deficient mice showed reduced insulin secretion, improved by vitamin D supplementation. Vitamin D also guards beta cells from cytokine-induced apoptosis (Bornstedt et al., 2019; Gysemans et al., 2005; Wolden-Kirk et al., 2014).

Regarding immunology, Hypovitaminosis D impacts T cell-mediated immunity, while excess suppresses immune functions. Immune-mediated illnesses including multiple sclerosis, type 1 diabetes, and systemic lupus are being researched (Cantorna et al., 1998; Lemire et al., 1992; Zella & DeLuca, 2003). Vitamin D therapy reduces inflammation, improves immunity (Griffin et al., 2001), and aids transplant rejection reduction (Hullett et al., 1998). More investigation is required to comprehend vitamin D analogs' mechanisms against autoimmune conditions.

1.4 Sources of Vitamin D

Vitamin D, a vital fat-soluble nutrient which functions as a hormone, comprises vitamin D₂ (Ergocalciferol) from plants and fungi, including Mushrooms with UV exposure, and vitamin D₃ (cholecalciferol), naturally present in foods like fatty fish and egg yolks. It's additionally added to fortified foods and supplements in many countries (J. Liu, 2012).

Table 1: Common Dietary Sources of Vitamin D (Uriu-Adams et al., 2013)

Food	IUs/serving	% DV
1 tablespoon of cod liver oil	1360	340
3 ounces cooked swordfish	566	142
3 ounces cooked salmon (sockeye)	447	112
3 ounces canned tuna in water	154	39
1 cup vitamin D-fortified orange juice	137	34
1 cup vitamin D-fortified nonfat, reduced fat, and whole milk	115-124	29-31
Yoghurt fortified with 20% of the DV for vitamin D, 6 oz.	80	20
1 tablespoon fortified margarine	60	15
2 sardines, canned in oil, drained	46	12
3 ounces cooked beef liver	42	11
1 big egg (the yolk contains vitamin D)	41	10
10% DV of vitamin D added to 0.75–1 cups of ready-to-eat cereal	40	10
1 ounce Swiss cheese	6	2

Table 1 presents vitamin D content in different dietary sources, including unfortified Western diets offering about 100 IU or 2.5 mg/day of vitamin D (Institute of Medicine, 1997). US retail milk and infant formulas are often fortified with varying levels, and traditional multivitamins contain 200-400 IU (5-10 mg) of vitamin D, with higher doses emerging (Hathcock et al., 2007; Patterson et al., 2010).

Outdoor workers' sun exposure can elevate serum 25(OH)D similar to oral doses of 2800-5000 IU (70-125 mg) per day. Sunlight converts 7-dehydrocholesterol to provitamin D, more available in the past. Certain professions have 25(OH)D levels around 122 nmol/L, while healthy sunbathers reach over 200 nmol/L (Barger-Lux & Heaney, 2002). However, levels decrease to 74 nmol/L (40% reduction) in winter, indicating intense sun exposure doesn't guarantee higher winter concentrations (Vieth, 1999).

1.5 Synthesis and Metabolism of Vitamin D

Vitamin D is distinct based on its availability from food sources and skin synthesis. Vitamin D₃ is produced by ultraviolet B radiation acting on 7-dehydrocholesterol (7DHC), enabling its acquisition through diet and skin (Provitamin D₃). The precursor, provitamin D₃, transforms to vitamin D₃ via heat isomerization in the skin, also converting to inert forms or reverting to 7DHC (MacLaughlin et al., 1982; Webb, 2006). Vitamin D gets into the bloodstream through a binding protein (WEBB et al., 1989). Figure 5 depicts the synthesis and metabolism of cutaneous vitamin D.

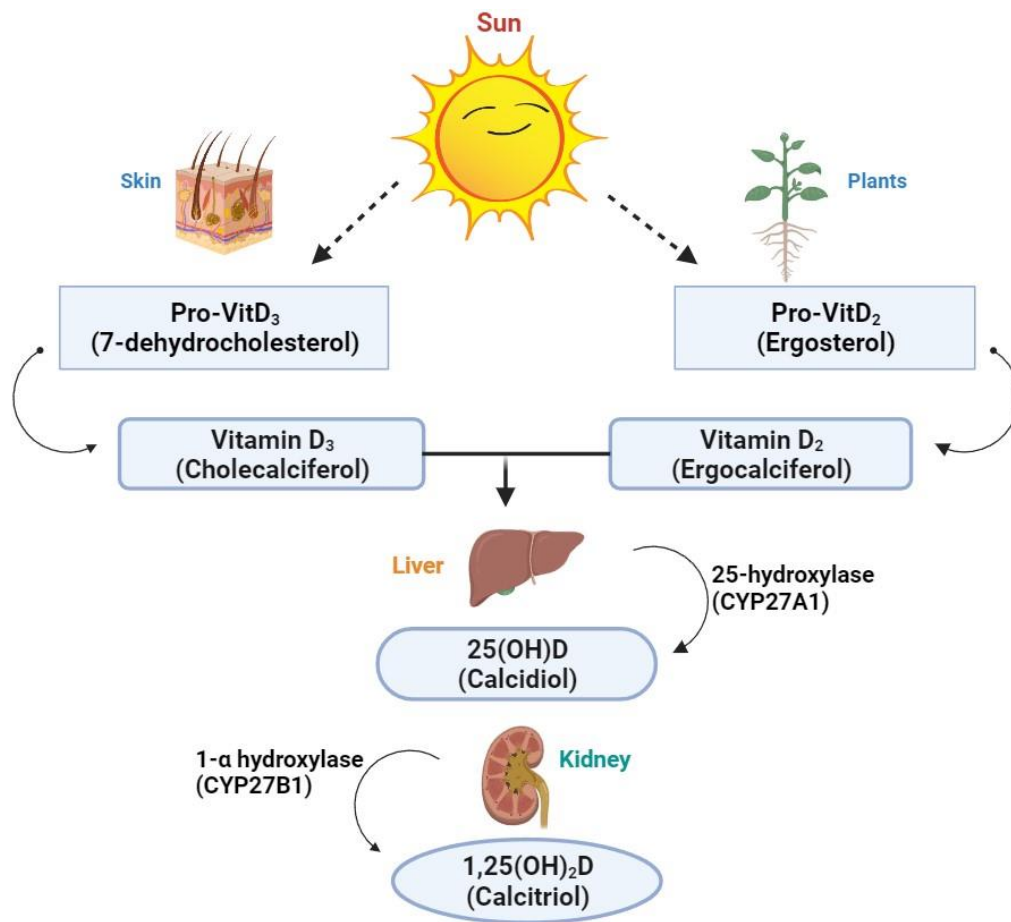


Figure 5: Cutaneous Vitamin D synthesis and Metabolism (Scientific Image and Illustration Software, 2023; Vuolo et al., 2012)

In the kidney, $1\alpha,25(\text{OH})_2\text{D}$ (calcitriol), the active hormone, results from the conversion of vitamin D_3 by 1α -hydroxylase (CYP27B1). Before this, the liver's enzyme 25-hydroxylase (CYP27A1) converts it to 25-hydroxyvitamin D ($25(\text{OH})\text{D}$) (Uriu-Adams et al., 2013).

1.6 Vitamin D Deficiency and its Complications

Vitamin D deficiency's definition is debated. Experts generally see levels below 50 nmol/L (20 ng/mL) as deficient, 51–74 nmol/L as insufficient, above 30 ng/mL as sufficient, and exceeding 375 nmol/L as toxic (150 ng/mL) (Chapuy et al., 1996; Dawson-Hughes et al., 2005; Holick, 2007; Holick et al., 2005; Malabanan et al., 1998; Thomas et al., 1998). Deficiency can have

both musculoskeletal and non-skeletal consequences (Heaney et al., 2003; Holick, 2007; Vieth, 1999).

Musculoskeletal Consequences

Inadequate vitamin D levels can result in rickets among children, accompanied by related signs, symptoms, and growth complications. In adults, insufficient vitamin D can exacerbate conditions like osteopenia and osteoporosis, increasing the likelihood of fractures (Bakhtiyarova et al., 2006; Chapuy et al., 1996; Holick, 2007; Huldschinsky, 1919; Nesby-O'Dell et al., 2002). Long-standing evidence associates musculoskeletal weakness with a lack of this nutrient, resulting in muscle weakness, body instability, and a heightened risk of falling (Bischoff-Ferrari, Dietrich, et al., 2004; Broe et al., 2007; Simpson et al., 1985; Visser et al., 2003). Insufficiency of vitamin D in adults may cause issues in bone mineralization, resulting in osteomalacia characterized by thickened osteoid layers on bones, potentially contributing to osteoporosis, both of which can elevate the likelihood of fractures, especially fractures in the hip (Lips, 2001). The interaction between PTH and serum 25(OH)D levels was explored in older adults, revealing that adequate 25(OH)D stabilizes status of PTH and that vitamin D supplements enhance bone mineral density, an effect subjected to the genotype of the vitamin D receptor (Lips, 2001).

Non-skeletal Consequences

Due to decreased synthesis of vitamin D₃, living in higher latitudes in the US is linked to an elevated incidence of common malignancies like prostate, colon, and breast cancer (Holick & Chen, 2008), with adults having below 50 nmol/L 25(OH)D status experiencing 30–50% enhanced likelihood of these cancers (Ahonen et al., 2000; Garland et al., 2006; Giovannucci et al., 2006; Grant, 2002; Grant & Garland, 2006).

Higher latitudes are additionally linked to elevated risks of multiple sclerosis, type 1 diabetes, and hypertension (Embry et al., 2000; Ponsonby et al., 2002; Rostand, 1997; Stene et al., 2000). Children with vitamin D intake of 2000 IU daily in their first year have a 78% lower type 1 diabetes risk, likewise women consuming over 400 IU of vitamin D daily possess a 40% lower chance of rheumatoid arthritis and multiple sclerosis (Hyppönen et al., 2001; Munger et al., 2004).

Moreover, higher latitudes and hypovitaminosis D elevate the likelihood of developing schizophrenia (McGrath, 2002) and depression (Gloth et al., 1999), while extreme vitamin D deficiency in African Americans raises tuberculosis risk and severity (Chan, 2000).

1.7 Rationale of the Study

The study investigates the link between maternal vitamin D Receptor gene's single nucleotide polymorphism, BsmI and the incidence of preterm birth and low birth weight. VDR BsmI SNPs impact vitamin D receptor protein and vitamin D metabolism, influencing bone wellness, immune function, and cellular growth. These SNPs reflect individual responses to vitamin D and disease susceptibility. The VDR gene is vital for fetal development and can potentially serve as a genetic predictor for assessing the risk adverse pregnancy outcomes. This review examines vitamin D's impact on pregnancy outcomes, extensively explores the correlation between BsmI SNP within the VDR gene and preterm birth & low birth weight, identifies additional genetic predictors, proposes prevention strategies, and provides future research recommendations.

1.8 Aim and Objectives of the Study

Aim

The study aims to examine the link between mothers' single nucleotide polymorphism of the vitamin D receptor gene (BsmI) and the risk of preterm birth and low birth weight among neonates.

Objectives

Specific objectives of the study are to:

- Demonstrate the need for mothers' vitamin D level for optimal pregnancy outcomes.
- Examine if maternal vitamin D intake, sun exposure, and age impact the relationship between mothers' VDR SNPs (specifically BsmI) and preterm birth & low birth weight.
- Examine the biological mechanisms underlying the association between maternal VDR SNPs, particularly BsmI and PTB & LBW, including changes in gene expression and protein function.

Chapter 2: Methodology

This review is based on credible scholarly articles from reputable sources. Relevant search terms and keywords were identified, including "pregnancy," "vitamin D," "vitamin D receptor," "BsmI polymorphism," "gestational duration," "preterm birth," and "birth weight." Multiple databases, such as PubMed, Google Scholar, Elsevier, ScienceDirect, and Wiley Online Library, were searched using advanced options to narrow results. Keywords were combined for better relevance, and reference lists were checked for additional sources. Initially, around 300 articles were found, with irrelevant ones excluded after title and abstract screening. Over 100 articles were extensively studied, notes were taken, and essential data were highlighted. Proper citations in "APA 7th Edition" style were ensured throughout the paper, with all sources well organized for easy access during writing. The gathered information was critically analyzed and used to compose the project paper.

Chapter 3: Vitamin D and Pregnancy

3.1 Significance of Vitamin D for Maternal and Child Health

Insufficient vitamin D levels at several life phases, including conception, fetal development, childhood, and adulthood, raise concerns about potential impacts. The diet of expectant and lactating mothers can affect fetal and newborn growth, potentially leading to lasting consequences like physical changes and health issues including type 2 diabetes, heart disease, obesity, and hypertension (Momentti et al., 2018). Adequate levels of this vitamin are imperative to both fetal and maternal well-being, especially for proper bone growth, dental health, and overall development during pregnancy (Fiscaletti et al., 2017).

Expectant mothers with ample vitamin D levels and increased calcium transfer to the fetus support proper bone development, reducing the risk of low blood calcium in newborns and the chances of both innate and developmental rickets in children. Mother's 25(OH)D may cross the placenta, potentially transmitting vitamin D deficiency (VDD) onto the fetus and newborn, with both immediate and prolonged effects (Fiscaletti et al., 2017).

In the course of pregnancy, calcitriol, the vitamin D's active form, substantially rises throughout the first trimester and doubles by the third. Beyond its role in providing fetal calcium, this rise is vital for immune adjustments essential for a healthy pregnancy. Vitamin D influences dendritic cell maturation and T-cell response balance, favoring Th2 dominance to prevent fetal rejection. This shift toward Th2 dominance critically supports a healthy pregnancy (Hayes et al., 2003; Raghupathy, 2001; Spilianakis et al., 2005).

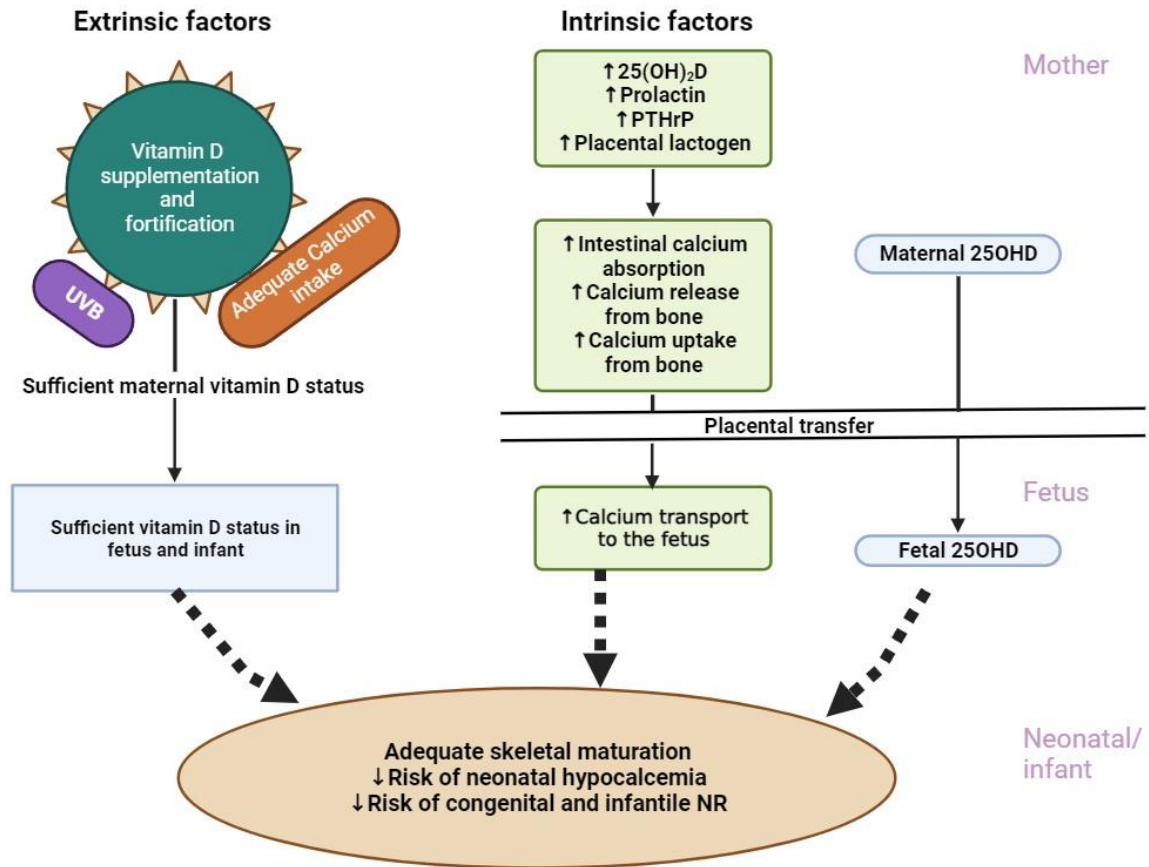


Figure 6: Schematic depiction of pregnancy factors and physiological alterations that promote optimal bone health in offspring (Fiscaletti et al., 2017; Scientific Image and Illustration Software, 2023)

Overall, vitamin D is essential for immune regulation during pregnancy, contributing to the maintenance of a healthy pregnancy and possibly protecting against pregnancy-related disorders such as preeclampsia. Nonetheless, further study is necessary to fully understand the mechanisms and determine the ideal vitamin D levels or supplementation for pregnant women (Evans et al., 2004).

3.2 Vitamin D Requirements for Pregnant Women

Pregnant women don't get enough vitamin D from the recommended 200 IU (5 g) per day, as specified by the Institute of Medicine, is 200 IU (5 g). In order to fulfill the nutritional requirements, it is recommended that pregnant women include a supplement comprising 1,000 IU of vitamin D with their prenatal nutrient containing 400 IU, resulting in a daily intake ranging from 1,500 to 2,000 IU. The maximum safe dosage may be extended up to 10,000 IU/day (Food and Nutrition Board Institute of Medicine, 1997; Holick et al., 2011).

3.3 Prevalence of Vitamin D Deficiency among Pregnant Women

Despite prenatal nutrient recommendations, alarmingly high rates of vitamin D shortage are seen during conception: 4% to 60% in mothers and 3% to 86% in newborns across countries (Palacios & Gonzalez, 2014; Prentice, 2008). In a recent North West England study, 27% of expectant mothers had insufficient vitamin D levels (50 nmol/L), which increased to 48% after delivery. In addition, 7% were deficient in vitamin D (25 nmol/L), and 11% remained deficient postpartum. 24% of four-month-old infants had unsatisfactory (25-50 nmol/L) vitamin D status (Emmerson et al., 2018). A UK study on pregnant teenagers raised concerns as 30% had below 25 nmol/L 25(OH)D readings (Baker et al., 2009). 84% of expecting women in India showed vitamin D concentrations below 22.5 ng/mL (Sachan et al., 2005). A rural Bangladesh research showed 47.2% had insufficient amount (30–50 nmol/L) of vitamin D, while 17.3% were deficient (less than 30.0 nmol/L) (Ahmed et al., 2021).

3.4 Unfavorable Pregnancy Outcomes linked to Vitamin D deficiency

Inadequate vitamin D levels throughout conception remain a rising concern globally. Studies report deficiencies in 30–80% of expectant females (Ginde et al., 2010). Low vitamin D levels result in undesirable consequences like gestational diabetes, pre-eclampsia, and preterm birth

(Aghajafari et al., 2013; J. Wen et al., 2017). It affects fetal size, bone formation, and neonatal health (Eckhardt et al., 2015; Miliku et al., 2016).

Fetal Repercussions

The fetus relies on maternal vitamin D and calcium stores, raising deficiency concerns (Walsh et al., 2013). Deficiency of vitamin D in newborns (cord blood at 11 ng/mL) occurs in 46% of supplemented mothers' babies (Basile et al., 2007). Maternal hypovitaminosis D is correlated with adverse neonatal outcomes, including small-for-gestational-age births, premature delivery, impaired tooth and bone growth, and higher infection risk (Karras et al., 2014).

Short for Gestational Age and Low Birth Weight

Burris and colleagues discovered that inadequate (below 10 ng/mL) 25(OH)D serum status tripled the incidence⁶⁵ of Short for Gestational Age (SGA) in the second trimester in one research finding (Burris et al., 2012). Another research involving more than 3,000 pregnancies discovered that low serum 25(OH)D at 13 weeks was connected to low weight at birth and increased SGA risk (Leffelaar et al., 2010). Adequate vitamin D (>15 ng/mL) prior to 26 weeks is related to a higher weight at birth and reduced SGA risk (Gernand et al., 2013). Despite mixed results, a 2013 meta-analysis confirmed the connection of low vitamin D with SGA prevalence (Aghajafari et al., 2013).

Bone Health

Gestational deficiency of vitamin D can affect long-term bone development and mineralization in childhood, since vitamin D is linked to fetal femoral growth (Ioannou et al., 2012; Morley et al., 2006; Walsh et al., 2013; Weiler et al., 2005). While several researches show shorter

long bones in babies of mothers with hypovitaminosis D (Morley et al., 2006), others contradict this (Gale et al., 2008; Weiler et al., 2005), prompting further research (Lawlor et al., 2013).

Respiratory Illness

Newborns are more likely to get respiratory syncytial virus infection and other respiratory illnesses if their mothers have vitamin D insufficiency (Belderbos et al., 2011; Camargo et al., 2011). Recurrent wheezing risk might be lower in kids with higher cord blood vitamin D (Camargo et al., 2007, 2011). The link to asthma remains unclear (Camargo et al., 2011; Gale et al., 2008), warranting more investigation.

Type 1 Diabetes

Children of moms with vitamin D insufficiency are potentially in threat of acquiring type 1 diabetes. A nested case-control research revealed a twofold greater chance in the offspring of pregnant women with inadequate vitamin D (Sorensen et al., 2012), while certain Finnish research showed no differences (Miettinen et al., 2012).

Atopic Symptoms

Atopic symptoms, including eczema and food-related allergies, might have a correlation with the mother's vitamin D level (Gale et al., 2008; Weisse et al., 2013). However, a study of 231 newborns over the span of their first year found that vitamin D levels below 20 ng per mill in cord blood corresponded to an increased prevalence of eczema (Jones et al., 2012).

Psychomotor development and neurological disease

Higher vitamin D levels corresponded with superior cognitive and psychomotor growth at 14 months in a Spanish sample (Morales et al., 2012), suggesting its role in pregnancy. Limited

sunlight exposure during birth month linked to multiple sclerosis risk (Torkildsen et al., 2012), while maternal milk intake and proper vitamin D could lower child multiple sclerosis risk (Mirzaei et al., 2011)

Maternal Repercussions

Pre-eclampsia

Low vitamin D has been attributed to an elevated prevalence of pregnancy issues, including pre-eclampsia. Studies associate pre-eclampsia with low vitamin D, especially in high-risk groups (Azar et al., 2011; Fernandez-Alonso et al., 2012; Powe et al., 2010; Shand et al., 2010). While a 2013 meta-analysis hints at a link, more research is needed for clarity, especially in high-risk women (Aghajafari et al., 2013).

Type 2 Diabetes

Gestational diabetes mellitus (GDM) is caused, or at least exacerbated, by a lack of vitamin D, which disrupts glucose regulation during conception (Aghajafari et al., 2013; Senti et al., 2012). Early in pregnancy, hypovitaminosis D enhance GDM risk, without regard for age, race, or weight. Higher vitamin D status lowers maternal hyperglycemia risk, notably for smokers (Tomedi et al., 2013), suggesting improved levels might reduce GDM risk.

Cesarean delivery

Research on vitamin D and caesarean delivery varies. Some studies suggest low vitamin D could raise caesarean likelihood, while some dispute this impact. Clarifying this connection is vital due to caesarean-related complications (Merewood et al., 2009; Scholl et al., 2012).

Depression

Initial findings hint at a relationship between low early pregnancy vitamin D and higher postpartum depression (Brandenbarg et al., 2012). African-American women show a similar association (Cassidy-Bushrow et al., 2012). Establishing causality is complex due to timing. More research is required to understand depression's possible link to vitamin D deficiency.

Chapter 4: Pathophysiology of Preterm Birth and Low Birth Weight

Approximately 70% of preterm births are spontaneous, with 45% due to early labor, while 25–30% from preterm premature rupture of membranes (PPROM). Additionally, roughly 30% result from obstetricians inducing early delivery due to an unfavorable uterine environment (Goldenberg et al., 2008). Genetic influences significantly impact maternal and fetal genomes and pregnancy duration. Recent studies encompass factors like the in utero environment, familial context, mother's genetics (13-25%), fetal genetic variation (11–35%), collectively shaping pregnancy duration (York et al., 2014). Exploring pregnancy complications, especially preterm birth (PTB), reveals disruptions in maternal immune responses, including the escalation of infection-related cytokines triggering premature labor, akin to preterm labor (Romero et al., 2006). The intricate interplay of fetal genotype effects and maternal genotype-influenced in utero environment further complicates our understanding (Alkhuriji et al., 2013; Crider et al., 2005; Grisaru-Granovsky et al., 2007; Rai, 2014; Siddesh et al., 2014). Given these complexities, it's clear our grasp of preterm birth mechanisms is limited, underscoring the need for intensified research and effective preventive strategies.

4.1 Risk Factors for Preterm Birth

Preterm birth (PTB) risk is heightened by numerous factors, as examined extensively in various reports (Alleman, 2014; Anum et al., 2009; Goldenberg et al., 2008; Richard E Behrman, 2007), with Figure 7 illustrating key risk factor categories (Sheikh et al., 2016).

Behavior & Socio-demographics

This category includes maternal demographic risk variables like socioeconomic position, age, race, marital status, and education, along with behavior such as consuming tobacco, alcohol, or drugs that increase susceptibility. Additional risk factors comprise vitamin D, folic acid, iron deficiencies, low prenatal weight, obesity, anxiety, depression, stressful work environments, and abuse, all implicated in PTB. Recent reviews delve into behavioral and sociodemographic aspects (Sheikh et al., 2016).

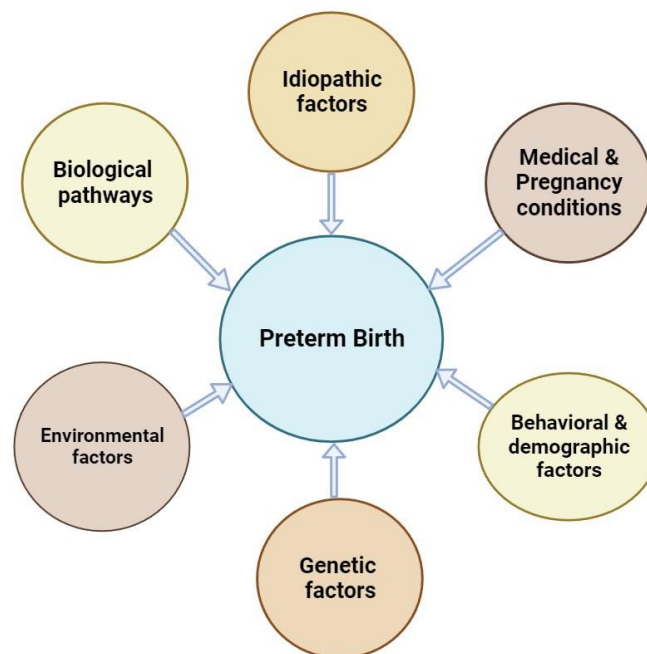


Figure 7: Risk factors associated with preterm birth (Scientific Image and Illustration Software, 2023; Sheikh et al., 2016)

Medical & Pregnancy Conditions

Factors such as multiple pregnancies, prior PTB history, short interpregnancy intervals, uterine distension, vaginal bleeding, hypertension, diabetes, thyroid disorders, cervical conditions, and uterine over-distension further heighten PTB risk, with medical and pregnancy conditions thoroughly explored in recent reviews (Sheikh et al., 2016).

Genetic Influence

Genetic factors significantly influence PTB, accounting for 25% to 40% of cases, as indicated by familial aggregation, genetic correlations, twin studies, and SNP-based investigations (Sheikh et al., 2016).

Environment

Additionally, environmental pollutants like Bisphenol A, air pollution, agricultural factors, and electronic waste contribute to PTB, which is under ongoing investigation (Richard E Behrman, 2007; Sheikh et al., 2016).

Biological Pathways

Biological pathways, including infections, inflammation, adrenal activation, decidua defects, uterine distension, and microbial infections, play critical roles in PTB, offering insights into its complexities (Sheikh et al., 2016).

In total, PTB, being a complex condition, stems from diverse factors, with about half having identifiable causes and the rest remaining idiopathic (Sheikh et al., 2016).

4.2 Complications of Preterm Birth

Premature delivery carries significant consequences, including higher newborn mortality, increased infection susceptibility, and greater risks of future health problems like neurodevelopmental, cardio-metabolic, and inflammatory disorders in premature infants (Arpino et al., 2010; Goedicke-Fritz et al., 2017; Markopoulou et al., 2019; Moster et al., 2008; Sonnenschein-van der Voort et al., 2014).. Inadequate bone mass worsens these effects, leading to post-birth growth issues (Finken et al., 2016) and frequent intensive care stays, driving

healthcare costs to over \$26 billion in the US and contributing to about 35% of infant deaths (IOM, 2007). Even late preterm births (34–36 weeks) show raised mortality risks up to age 36 due to various health concerns (Crump et al., 2011).

Despite better survival rates, concerns remain about learning, psychological challenges, and adulthood issues. Decreasing gestational age is linked to more medical and social problems in adulthood, posing unique challenges in motor skills, cognition, behavior, psychology, and social functioning during early education for very premature births (Bhutta et al., 2002; Hack et al., 2005; Hille et al., 2001; Litt et al., 2005; Marlow et al., 2005; Saigal et al., 2003). Recent studies link premature toddlers to autism spectrum disorder (Limperopoulos et al., 2008). Earlier research also suggests lower academic performance and reduced likelihood of parenthood among very preterm individuals, perpetuating a cycle of health challenges that extends to women born prematurely, who face higher preterm birth risks (Swamy et al., 2008).

4.3 Risk Factors for Low Birth Weight

A research performed by Yadav and colleagues, investigated variables that may lead to low birth weight (LBW) in 258 mothers, with 80% from rural areas. It found that the majority of LBW infants were born to moms aged 19 to 30, while normal birth weight babies had mothers aged 20 to 29, consistent with similar research in developing countries (Karim et al., 2011; Nahar et al., 1998).

Residence didn't significantly impact birth weight, implying equal access to maternal and child health services. Illiteracy was prevalent (61% of mothers), with 26% having LBW babies; educated mothers had lower LBW rates (15%), possibly due to greater health awareness. Household head education influenced birth weight, but parental education didn't. Religion and birth weight were not significantly linked (Yadav et al., 2011). Higher family income is correlated with fewer LBW cases (Yadav et al., 2011).

The birth-to-conception interval had minimal impact, indicating nutrition as a more influential factor. Previous pregnancy history (abortion, stillbirth, and neonatal death) had no significant link to birth weight (Yadav et al., 2011), aligning with prior studies (Khatun & Rahman, 1970).

4.4 Complications of Low Birth Weight

Babies born with very low weight have an 85% chance of surviving hospitalization, while premature birth carries a 2-5% risk of medical complications leading to death within 2 years (Fanaroff et al., 2007). Table 2 outlines major short and long-term issues tied to low birth weight (Eichenwald & Stark, 2008).

Table 2: Major Complications in Infants with Extremely Low Birth Weight (Eichenwald & Stark, 2008)

Affected Organ or System	Acute/Immediate Problems	Long-Term Problems
Pulmonary	Neonatal apnea, bronchopulmonary dysplasia, RDS, air leak	Reactive pulmonary illness, asthma, BPD
Gastrointestinal	food intolerance, necrotizing enterocolitis, growth failure, hyperbilirubinemia	Short bowel syndrome, cholestasis, growth failure
Central nervous system	Periventricular white-matter degeneration, intraventricular haemorrhage, hydrocephalus	hearing loss, hydrocephalus, cerebral shrinkage, neurodevelopmental delay
Immunologic	Immune deficits, perinatal and hospital-acquired infections	Bronchitis, respiratory syncytial virus infection

Ophthalmologic	Premature retinopathy	Myopia, strabismus, retinal detachment, blindness
Cardiovascular	pulmonary hypertension, patent ductus arteriosus, hypotension	pulmonary hypertension, adult-onset hypertension
Renal	Acid-base disturbances, electrolyte and water imbalances	Adulthood hypertension
Hematologic	Iatrogenic anemia, the requirement for frequent transfusions, preterm anemia	N/A
Endocrine	Hypoglycemia, momentary low thyroxine levels, a lack of cortisol	Impaired control of blood sugar, elevated insulin resistance

Chapter 5: Vitamin D Receptor Gene Polymorphisms

5.1 Vitamin D Receptor and its Location

A pivotal steroid hormone receptor, the vitamin D receptor (VDR), often called the calcitriol receptor, oversees diverse biological effects attributed to vitamin D. Encoded by the VDR gene (gene Identifier: 7421 and Molecular Identification Number: 601769), it actively manages gene transcription, mediating vitamin D's impact (Whitfield et al., 1995), including nuclear transcription factor activity prompted by ligands (Kato, 2000). Calcitriol, the primary ligand, notably binds to VDR in the nucleus (Knabl et al., 2017).

Recent research highlights elevated VDR gene expression across various cell types, including the immune system, thymus, type-II alveolar cells, osteoblasts, chondrocytes, skin epithelium, and placental components like villous trophoblasts, decidua, and placental vessel smooth muscle cells (Barrera et al., 2008; Knabl et al., 2017; Pike & Meyer, 2012; Pospeschova et al., 2009; Shahbazi et al., 2011). However, a distinct pattern emerges, with fewer vitamin D receptors present in erythrocytes, fully developed striated muscle cells, and differentiated cerebral cortex neural cells (Bischoff-Ferrari, Borchers, et al., 2004).

5.2 How does Vitamin D Receptor Work?

Compelling evidence strongly highlights the significant repercussions of vitamin D level imbalances on human wellness, leading to diseases (Zhang & Naughton, 2010). The vitamin D receptor in the nucleus plays an integral part in how the body reacts to this nutrient. Vitamin D's active form, calcitriol, attaches to the blood's vitamin D binding protein (VDBP). It then works with VDR in the cytoplasm of cells to form a molecular ensemble. Accompanied by the retinoic acid X receptor (RXR), the combination departs for the nucleus, forming a heterodimer (Barsony & Prufer, 2002; Bikle, 2014). Together, they enable gene transcription relying on vitamin D, which governs a significant portion of the human genome (3–10%), particularly in

mineral metabolism. This regulatory mechanism extends to metabolic pathways and is intertwined with embryological and immunological responses (Nair & Maseeh, 2012; Rosen et al., 2012; Zhu et al., 2013)..

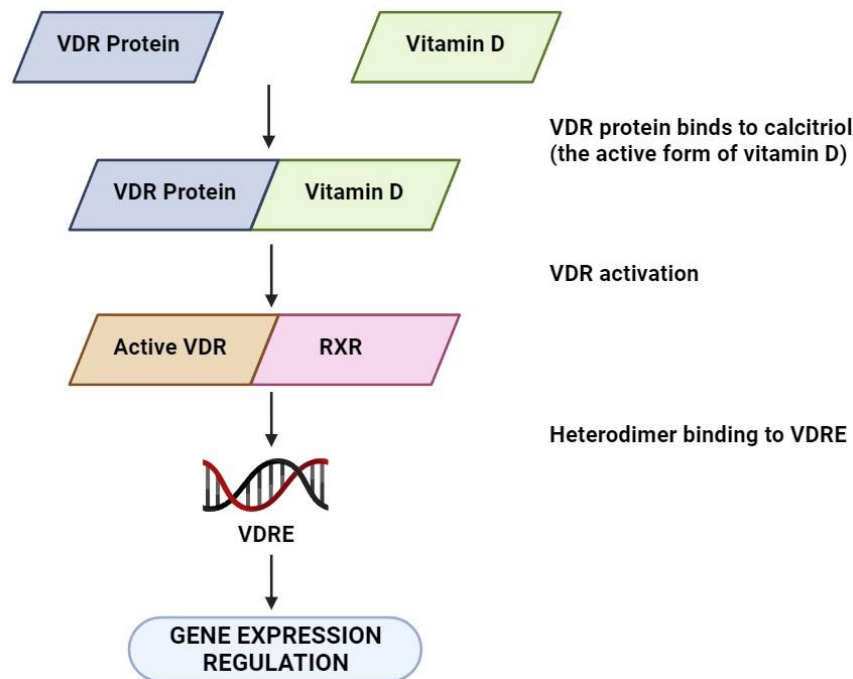


Figure 8: VDR protein's function in regulating gene expression (Agliardi et al., 2023; Scientific Image and Illustration Software, 2023)

5.3 Signaling by VDRs during Preterm Birth

In their research, scientists have found significant links between maternal vitamin D levels and placenta's defense against infectious microbes via toll-like receptors (Evans et al., 2006; N. Liu et al., 2009). Vitamin D stimulates cathelicidin production in decidua and cytotrophoblasts, crucially countering bacterial infections. Insufficient vitamin D limits toll-like receptor activation in systemic macrophages, decreasing cathelicidin levels and compromising microbial defense (Hewison, 2011; N. Q. Liu & Hewison, 2012). Moreover, vitamin D's active form, calcitriol, suppresses key immune response cytokines (IL-6, TNF- α , & GM-CSF-2).

During pregnancy, myometrial smooth muscle cells and tissue display heightened inflammatory responses to lipopolysaccharides (LPS) and IL-1, which are associated with premature birth in mouse models (Helmer et al., 2002; Sehringer et al., 2000). An important discovery is that vitamin D inhibits protein production induced by LPS and IL-1 in UtSM cells, reducing levels of inflammation-associated contractility proteins (Buhimschi et al., 2003; Romero et al., 1991; Thota et al., 2013). This finding underscores the significance of mitigating premature birth risk and highlights vitamin D's protective role in regulating placental and uterine responses to infections and inflammation (N. Q. Liu et al., 2011).

5.4 Gene Polymorphism and their Types

The concept of genetic polymorphism pertains to the inheritance of an attribute that is controlled by a solitary genetic locus containing 2 alleles, where the allele with the lower prevalence has an abundance of 1% or above. It includes random processes or exogenous influences like viruses or radiation causing DNA sequence changes in individuals, groups, or populations. Mutations are DNA sequence differences linked to diseases. External perturbations are termed "mutations," not "polymorphisms," even if they create polymorphisms. Mutations in nucleotides cause polymorphisms, which are inherited from parents (Ismail & Essawi, 2012).

It's important to note that polymorphisms in genes come in a variety of forms:

- Tandem repeat polymorphisms
- Copy-number variations
- Single Nucleotide polymorphism

5.5 Single Nucleotide Polymorphism of VDR Genes

Located on chromosome 12 (12q13-14) over 75 kilobases, the VDR gene has nine exons (Suksawatamnuay et al., 2020) and allelic variations akin to other receptor genes on the same chromosome (Taymans et al., 1999). Extensive research has honed in on the genetic polymorphism of VDR, notably BsmI (rs1544410), ApaI (rs797532), TaqI (rs731236), and FokI (rs2228570), in relation to clinical outcomes (Agliardi et al., 2023). Figure 5.1 visually represents VDR activation and associated polymorphisms.

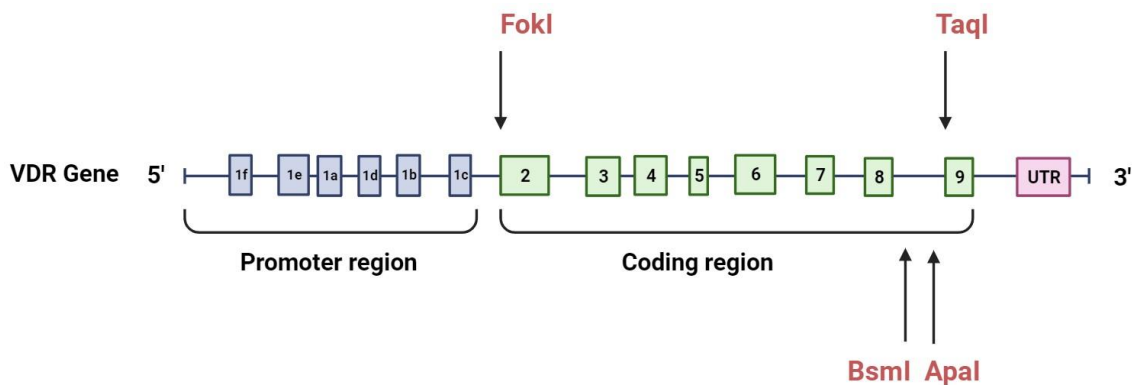


Figure 9: Common SNPs of the VDR gene (Agliardi et al., 2023; Scientific Image and Illustration Software, 2023)

The exact mechanisms behind genetic variations within the VDR gene impacting observed traits in association studies are uncertain. Specific SNP locations, like TaqI at exon-9; FokI at exon-2; ApaI and BsmI at intron-8, are of interest. Such SNPs involve distinct nucleotide changes, such as A to G for TaqI, G to A for FokI, G to A for BsmI, and A to C for ApaI (Kosik et al., 2020). TaqI alters protein function, while BsmI and ApaI change mRNA stability and expression, impacting VDR synthesis, vitamin D levels, calcium balance, and non-classical functions (Swamy et al., 2011).

5.6 VDR BsmI Gene Polymorphism

The VDR BsmI gene polymorphism (rs1544410), found within an intron between exons 8 and 9, is believed to impact the translational activity of VDR and the stability of its mRNA. because of significant linkage disequilibrium with a polyadenosine microsatellite repeat in the 3' untranslated region (Mangin et al., 2014; Mory et al., 2009), frequently co-occurring with the A allele (ApaI) and the T allele (TaqI). This relationship is linked to enhanced serum 1,25(OH)₂D₃ levels (Marco et al., 1999; Morrison et al., 1994), while some studies indicate a link between the B allele and reduced VDR activity (Morrison et al., 1994; Uitterlinden et al., 2004), suggesting a negative feedback mechanism where the VDR ligand causes CYP27B1 inhibition, limiting excessive 1,25(OH)₂D₃ production (Takeyama et al., 1997). This interaction may protect heterozygous individuals by maintaining intermediate 1,25(OH)₂D₃ concentration in the serum and activity of VDR (Carless et al., 2008). According to recent studies, the VDR gene polymorphism BsmI is linked to enhanced intestinal calcium absorption in children & women (Ames et al., 1999; Gennari et al., 1997), and heightened renal phosphate absorption in men, shedding insight on its influence on calcium and phosphate metabolism (Ferrari et al., 1999).

5.7 VDR Gene Polymorphisms and Adverse Pregnancy outcomes

The vitamin D Receptor gene variations have been linked to a range of health conditions, including metabolic disorders, immune challenges, and different cancers (Valdivielso & Fernandez, 2006). These variations have prompted investigations into their potential roles in predicting complications during pregnancy and impacting the health of both mothers and newborns (Baczyńska-Strzecha & Kalinka, 2016; Barchitta et al., 2018; Randis, 2008).

Regarding neonatal repercussions, premature infants are vulnerable to conditions like respiratory distress syndrome (RDS) due to their underdeveloped lungs (Course &

Chakraborty, 2020). Vitamin D has shown potential for reducing RDS risk, particularly at elevated levels (Ataseven et al., 2014). Genetic variations within the VDR gene, such as the TaqI polymorphism, appear to be correlated with an elevated RDS risk (Ustun et al., 2020). Similarly, intraventricular hemorrhage (IVH), which involves genetic and environmental factors, might be influenced by VDR gene variations affecting vascular endothelial cell function (Ni et al., 2014; Szpecht et al., 2017). The likelihood of bronchopulmonary dysplasia (BPD), a condition tied to insufficient lung maturation, has been linked to the VDR gene polymorphism ApaI (Kosik et al., 2020). Additionally, the Toll-like receptor 4 (TLR4), involved in NEC development, is influenced by the VDR gene polymorphism ApaI, indicating a potential role for this vitamin in TLR4 signaling (Kosik et al., 2020).

Maternal repercussions include pre-eclampsia, where VDR gene polymorphisms like BsmI and FokI have been connected to hypertension beyond pregnancy (Bodnar et al., 2007). The FokI polymorphism, for instance, may lower hypertension risk through its effect on plasma renin activity (Vaidya et al., 2011). Gestational Diabetes Mellitus (GDM), influenced by genetic along with environmental factors, has ties to VDR gene variations, potentially predisposing people to type-1 and type-2 diabetes (Motohashi et al., 2003; Tizaoui et al., 2014). Certain variants like BsmIBB, BsmIBb, and TaqItt predispose to T1DM (Sahin et al., 2017), while VDR FokI, TaqI, and ApaI SNPs show associations with GDM in diverse populations (Aslani et al., 2011; Rahmannedhad et al., 2016). A possible osteoporosis risk, particularly in postmenopausal women, has been explored in relation to VDR gene haplotypes, with varying effects observed in different cell lines (Gennari et al., 1997).

Chapter 6: Findings on Association of Single Nucleotide VDR Gene Polymorphism BsmI with Preterm Birth & Low Birth Weight

6.1 Interrelation between VDR BsmI SNP & Preterm Birth

Manzon and colleagues studied ApaI, TaqI, FokI and BsmI polymorphisms among a population residing in Israel to determine their impacts on preterm delivery. The research found substantial differences between preterm deliveries and controls, notably in mothers' FokI and TaqI VDR alleles. Interestingly, the maternal FokI variation had an OR of 3.317%, suggesting a higher risk of preterm delivery. ApaI, TaqI, and BsmI genetic variants did not significantly affect preterm birth, with odds ratios of 0.41, 0.232, and 1.13, respectively (Manzon et al., 2014).

In another study, Baczyńska-Strzecha and her colleague examined 100 preterm and 99 full-term Polish moms in 2016. Researchers found three genetic variations in participants: ApaI, TaqI, and BsmI. The research found no substantial genetic variation differences between preterm and full-term groups, despite these variances. Intriguingly, the preterm cohort displayed a higher frequency of BsmI/BB-ApaI/aa-TaqI/tt & BsmI/bb-ApaI/AA-TaqI/TT variants. Conversely, the preterm delivery incidence was also lowered by two genotype combinations: BsmI/BB-ApaI/Aa-TaqI/tt & BsmI/Bb-ApaI/AA-TaqI/Tt. This shows that certain genetic differences may raise the chance of preterm birth, while others may reduce it (Baczyńska-Strzecha & Kalinka, 2016).

Rosenfeld and his colleagues found a substantial correlation among maternal BsmI polymorphism and likelihood of PTB in a separate investigation, comprising 146 Israeli-Jewish preterm and 229 full-term women. Remarkably, the research found a negative connection between maternal BsmI polymorphism A allele count and PTB risk. Even after controlling for other covariates, this link remained strong, indicating that a greater A allele count continuously

lowered PTB risk. The research also found that mothers with spontaneous miscarriages had a higher PTB risk, especially if their babies had non-mutated (homozygous BB or bb) BsmI gene variations rather than the mutated (heterozygous Bb) genotype. These data show that the maternal BsmI polymorphism may affect PTB risk in women who have had previous instances of miscarriage (Rosenfeld et al., 2017).

In their initial 2018 Mamma and Bambino Cohort research, Barchitta and colleagues found no association between the VDR BsmI SNP and preterm birth. Integrating their data with Rosenfeld and Baczyńska-Strzecha's findings and using the Q-test and I^2 -test for heterogeneity evaluation, they found no significant differences. However, their meta-analysis of many genetic models found a negative correlation between mothers' BsmI polymorphism and premature birth. The recessive variant of the rs1544410 (BsmI) gene (AA vs. GG + AG) was linked to a lower incidence of PTB, suggesting a protective effect. The dominating model (AA+AG vs. GG) didn't show a meaningful correlation. The meta-analysis confirms that BsmI protects against preterm birth, with the extent varying by genetic model (Baczyńska-Strzecha & Kalinka, 2016; Barchitta et al., 2018; Rosenfeld et al., 2017).

Dutra accompanied by his colleagues found numerous interesting results in another study of 40 moms with preterm neonates (PTNs) as cases and 92 moms with full-term newborns (FTN) as controls. The maternal BsmI/CT single nucleotide variation (SNV) was more prevalent in the FTN group than in the PTN group, indicating a genetic composition linked to a lower chance of preterm delivery. Conversely, the BsmI/TT genotype elevated preterm birth risk by 2.36 times when paired with a 25 (OH)D deficit. This shows that the TT genotype and low vitamin D levels increase preterm birth risk. Preterm newborns had more BsmI/TT genotypes, whereas FTN neonates had more CT genotypes of the BsmI SNV (Dutra et al., 2020). Another case-control study conducted in the same year in Slovenia didn't reveal any substantial association for VDR BsmI SNP with preterm birth (Gašparović Krpina et al., 2020)

A study Wang and colleagues at Zhoushan Maternal and Child Health Hospital, China, Wang et al. investigated the link between genetic variation BsmI (rs1544410) and gestational week from August 2011 to May 2018. However, their findings showed an absence of correlation between BsmI and the gestational week, indicating limited impact on gestation length (Wang et al., 2021).

Table 3: Studies on VDR gene variations and preterm birth risk

Population Origin	Participants	VDR SNPs	Findings	Reference
Israel (Jewish)	33 Caucasian moms with PTNs (between 24 and 35 weeks), and 98 with FTNs	TaqI BsmI ApaI FokI	Preterm mothers were more likely to have the FokI/C gene. However, no link was seen between the BsmI genotype and preterm birth.	(Manzon et al., 2014)
Poland	100 Caucasian moms with PTNs (22-36.6 weeks) and 99 moms with FTNs	TaqI BsmI ApaI	Despite no differences in individual genotype frequencies, preterm birth mothers were more likely to carry the genotypic combinations: BsmI/BB-ApaI/aa-TaqI/tt & BsmI/bb-ApaI/AA-TaqI/TT	(Baczyńska-Strzecha & Kalinka, 2016)
Israel (Jewish)	146 white moms with their PTNs (at	TaqI BsmI	The genotype ApaI/AA appeared to be linked to an	(Rosenfeld et al., 2017)

	24–36 weeks), 229 women with their FTNs.	<p>ApaI</p> <p>FokI</p>	elevated preterm birth incidence. If their neonates received the non-mutated BsmI gene variant (BB or bb), mothers who had natural miscarriages were more likely to deliver prematurely.	
Italy	17 moms & their PTNs (< 37 weeks gestation), 187 moms & their FTNs	<p>BsmI</p> <p>ApaI</p> <p>FokI</p> <p>TaqI</p>	FokI polymorphism genotypes enhanced preterm birth risk in mothers. However, recessive BsmI (AA vs. GG + AG) reduced preterm birth risk.	(Barchitta et al., 2018)
Slovenia	118 spontaneously induced PTB following natural conception cases and 119 term singleton controls	<p>ApaI</p> <p>TaqI</p> <p>BsmI</p> <p>FokI</p> <p>Cdx2</p>	No association for BsmI SNP was discovered.	(Gašparović Krpina et al., 2020)
Brazil (Southeast)	40 moms and their PTNs (between 23 and 32 weeks), and 92 moms with FTNs	<p>BsmI</p> <p>ApaI</p> <p>FokI</p> <p>TaqI</p>	Prematurity risk was increased by the BsmI/TT as well as ApaI/AA genotypes, irrespective of Insufficiency in vitamin D.	(Dutra et al., 2020)

China	A total of 3465 pregnant women, with 202 of them being PTB	BsmI	No substantial association between BsmI and gestational week was discovered	(Wang et al., 2021)
-------	--	-------------	---	---------------------

6.2 Interrelation between VDR BsmI SNP & Low Birth Weight

Limited research has addressed genetic factors affecting birth outcomes related to vitamin D metabolism and overlooked 'maternal' VDR gene variations in newborn birth weight. However, potential ethnicity-specific responses to maternal VDR gene polymorphisms regarding birth weight hint at the need for further investigation. This emphasizes the necessity of comprehensively understanding how genetics and vitamin D contribute to birth outcomes across diverse ethnic groups (Swamy et al., 2011).

Lorentzon and colleagues found a connection between the VDR BsmI SNP & birth height, implying its influence on fetal bone development. In contrast, birth weight showed no significant correlation (Lorentzon et al., 2000). Within a primarily Caucasian cohort, insufficient maternal serum 25(OH)D status was linked to reduced birth weight in higher-functioning VDR FokI genotypes. However, no noteworthy correlation emerged between the BsmI SNP and birth weight (Morley et al., 2009).

Silvano and his team explored SGA neonates who didn't grow much after birth among prepubertal children. Intriguingly, no significant variations were detected in BsmI genotype distributions across diverse birth weight categories (Silvano et al., 2011). Similarly, A meta-analysis conducted on data from the "Mamma & Bambino" Cohort failed to reveal substantial variations in gestational length or birth weight among different BsmI genotypes (Barchitta et al., 2018).

Based on findings from a recent case-control research, it was shown that the FokI/TT variant of VDR exhibited an association with reduced birth weight in newborns of European descent who had spontaneous preterm birth (SPTB). There were no observed associations between VDR BsmI SNP and any further medical characteristics of SPTB mothers and their babies (Gašparović Krpina et al., 2020).

Ultimately, available evidence suggests that BsmI genotypes may not substantially impact newborn weight.

6.3 Limitations of the Findings

Conflicting VDR polymorphism research arises from variable study factors, notably sample sizes impacting reliability and generalization. Small studies, like Baczyńska-Strzecha's, hold limited relevance (Baczyńska-Strzecha & Kalinka, 2016). Ethnicity profoundly affects SNV distribution, particularly in mixed populations such as Asians and Caucasians, resulting in distinct VDR polymorphism prevalence and associations (Manzon et al., 2014). Racial origins contribute to inconsistent findings (Dutra et al., 2020). Unmeasured variables, like maternal practices and vitamin D levels, might influence associations (Barchitta et al., 2018). Genetic naming variations and analysis methods further complicate interpretation (den Dunnen et al., 2016). Recognizing ethnicity, unmeasured variables, and genetic nomenclature is vital for accurate VDR polymorphism research interpretation. Addressing these complexities in future studies is essential (den Dunnen et al., 2016; Dutra et al., 2020).

Chapter 7: Conclusion and Future Recommendations

Conclusion

Both preterm birth and low birth weight are serious worldwide healthcare concerns with complex causes. Maternal vitamin D deficiency is shown to cause preterm birth, and the VDR gene, responsible for the encoding of vitamin D receptor, takes part in regulating the levels of this vitamin through genetic variations (SNPs). Several researches have delved into the correlation between VDR BsmI SNP and the likelihood of preterm birth, with some providing evidence for the association while others failing to reach a conclusive outcome. However, almost no evidence of a BsmI SNP connection with low birth weight has yet been reported. In essence, the current findings are inconclusive, highlighting the need for more research to comprehensively comprehend how VDR gene SNPs relate to adverse pregnancy outcomes. The mother's health, lifestyle, surroundings, infections, race, ethnicity, and smoking all influence birth weight and the premature birth risk. Additionally, the variation in a particular gene is not the only genetic component that influences adverse pregnancy outcomes. Nonetheless, ensuring sufficient vitamin D intake and closely monitoring pregnant women and fetuses are essential for achieving optimal outcomes.

Future Recommendations

Future research recommendations for this subject include:

- Increasing sample sizes for stronger statistical power by studying larger populations;
- Including diverse ethnic groups to grasp VDR polymorphisms' impact on PTB and LBW;
- Analyzing functional aspects of VDR polymorphisms for their link to PTB and LBW;
- Considering maternal stress, nutrition, and pollutant exposure in future studies to better understand PTB and LBW risk;
- Conducting longitudinal studies to observe PTB and LBW progression while factoring in genetics and the environment; and
- Exploring epigenetic changes for a deeper understanding of VDR polymorphisms' connection to PTB and LBW.

Health organizations and governments should increase awareness of PTB and LBW risks, targeting pregnant women and the public through educational campaigns, materials, and community outreach. Pregnant women should prioritize a healthy lifestyle, infection prevention, sufficient vitamin D intake, monitoring, safe supplementation, and healthcare provider guidance.

References

- Aghajafari, F., Nagulesapillai, T., Ronksley, P. E., Tough, S. C., O'Beirne, M., & Rabi, D. M. (2013). Association between maternal serum 25-hydroxyvitamin D level and pregnancy and neonatal outcomes: systematic review and meta-analysis of observational studies. *BMJ*, *346*(mar26 4), f1169–f1169. <https://doi.org/10.1136/bmj.f1169>
- Agliardi, C., Guerini, F. R., Bolognesi, E., Zanzottera, M., & Clerici, M. (2023). VDR Gene Single Nucleotide Polymorphisms and Autoimmunity: A Narrative Review. *Biology*, *12*(7), 916. <https://doi.org/10.3390/biology12070916>
- Ahmed, F., Khosravi-Boroujeni, H., Khan, M. R., Roy, A. K., & Raqib, R. (2021). Prevalence and Predictors of Vitamin D Deficiency and Insufficiency among Pregnant Rural Women in Bangladesh. *Nutrients*, *13*(2), 449. <https://doi.org/10.3390/nu13020449>
- Ahonen, M. H., Tenkanen, L., Teppo, L., Hakama, M., & Tuohimaa, P. (2000). Prostate cancer risk and prediagnostic serum 25-hydroxyvitamin D levels (Finland). *Cancer Causes & Control : CCC*, *11*(9), 847–852. <https://doi.org/10.1023/a:1008923802001>
- Akter, R., Afrose, A., Sharmin, S., Rezwan, R., Rahman, M. R., & Neelotpol, S. (2022). A comprehensive look into the association of vitamin D levels and vitamin D receptor gene polymorphism with obesity in children. *Biomedicine & Pharmacotherapy*, *153*, 113285. <https://doi.org/10.1016/j.biopha.2022.113285>
- Alkhuriji, A. F., Alhimaidi, A. R., Babay, Z. A., & Wary, A. S. (2013). The relationship between cytokine gene polymorphism and unexplained recurrent spontaneous abortion in Saudi females. *Saudi Medical Journal*, *34*(5), 484–489. <http://www.ncbi.nlm.nih.gov/pubmed/23677264>
- Alleman, B. W. (2014). *PRETERM BIRTH: PREDICTION, PREVENTION, CARE*.

- Ames, S. K., Ellis, K. J., Gunn, S. K., Copeland, K. C., & Abrams, S. A. (1999). Vitamin D Receptor Gene FokI Polymorphism Predicts Calcium Absorption and Bone Mineral Density in Children. *Journal of Bone and Mineral Research*, *14*(5), 740–746. <https://doi.org/10.1359/jbmr.1999.14.5.740>
- Anum, E. A., Springel, E. H., Shriver, M. D., & Strauss, J. F. (2009). Genetic Contributions to Disparities in Preterm Birth. *Pediatric Research*, *65*(1), 1–9. <https://doi.org/10.1203/PDR.0b013e31818912e7>
- Arpino, C., Compagnone, E., Montanaro, M. L., Cacciatore, D., De Luca, A., Cerulli, A., Di Girolamo, S., & Curatolo, P. (2010). Preterm birth and neurodevelopmental outcome: a review. *Child's Nervous System*, *26*(9), 1139–1149. <https://doi.org/10.1007/s00381-010-1125-y>
- Aslani, S., Hossein-Nezhad, A., Mirzaei, K., Maghbooli, Z., Afshar, A. N., & Karimi, F. (2011). VDR FokI polymorphism and its potential role in the pathogenesis of gestational diabetes mellitus and its complications. *Gynecological Endocrinology*, *27*(12), 1055–1060. <https://doi.org/10.3109/09513590.2011.569786>
- Ataseven, F., Aygün, C., Okuyucu, A., Bedir, A., Küçük, Y., & Küçüködük, Ş. (2014). Is vitamin D deficiency a risk factor for respiratory distress syndrome? *International Journal for Vitamin and Nutrition Research*, *83*(4), 232–237. <https://doi.org/10.1024/0300-9831/a000165>
- Azar, M., Basu, A., & Jenkins, A. J. (2011). Serum carotenoids and fat-soluble vitamins in women with type 1 diabetes and preeclampsia: A longitudinal study. *Diabetes Care*, *34*. <https://doi.org/10.2337/dc10-2145>
- Baczyńska-Strzecha, M., & Kalinka, J. (2016). Influence of Apa1 (rs7975232), Taq1 (rs731236) and Bsm1 (rs154410) polymorphisms of vitamin D receptor on preterm birth

- risk in the Polish population. *Ginekologia Polska*, 87(11), 763–768.
<https://doi.org/10.5603/GP.2016.0084>
- Baker, P. N., Wheeler, S. J., Sanders, T. A., Thomas, J. E., Hutchinson, C. J., Clarke, K., Berry, J. L., Jones, R. L., Seed, P. T., & Poston, L. (2009). A prospective study of micronutrient status in adolescent pregnancy. *The American Journal of Clinical Nutrition*, 89(4), 1114–1124. <https://doi.org/10.3945/ajcn.2008.27097>
- Bakhtiyarova, S., Lesnyak, O., Kyznesova, N., Blankenstein, M. A., & Lips, P. (2006). Vitamin D status among patients with hip fracture and elderly control subjects in Yekaterinburg, Russia. *Osteoporosis International*, 17(3), 441–446. <https://doi.org/10.1007/s00198-005-0006-9>
- Barchitta, M., Maugeri, A., La Rosa, M. C., Lio, R. M. S., Favara, G., Panella, M., Cianci, A., & Agodi, A. (2018). Single nucleotide polymorphisms in vitamin D receptor gene affect birth weight and the risk of preterm birth: Results from the “mamma & bambino” cohort and a meta-analysis. *Nutrients*, 10(9), 1172. <https://doi.org/10.3390/nu10091172>
- Barger-Lux, M. J., & Heaney, R. P. (2002). Effects of Above Average Summer Sun Exposure on Serum 25-Hydroxyvitamin D and Calcium Absorption. *The Journal of Clinical Endocrinology & Metabolism*, 87(11), 4952–4956. <https://doi.org/10.1210/jc.2002-020636>
- Barrera, D., Avila, E., Hernández, G., Méndez, I., González, L., Halhali, A., Larrea, F., Morales, A., & Díaz, L. (2008). Calcitriol affects hCG gene transcription in cultured human syncytiotrophoblasts. *Reproductive Biology and Endocrinology: RB&E*, 6, 3. <https://doi.org/10.1186/1477-7827-6-3>
- Barsony, J., & Prufer, K. (2002). Vitamin D receptor and retinoid X receptor interactions in motion. *Vitamins and Hormones*, 65, 345–376. <https://doi.org/10.1016/s0083->

- Basile, L. A., Taylor, S. N., Wagner, C. L., Quinones, L., & Hollis, B. W. (2007). Neonatal vitamin D status at birth at latitude 32°72': evidence of deficiency. *Journal of Perinatology*, 27(9), 568–571. <https://doi.org/10.1038/sj.jp.7211796>
- Belderbos, M. E., Houben, M. L., & Wilbrink, B. (2011). Cord blood vitamin D deficiency is associated with respiratory syncytial virus bronchiolitis. *Pediatrics*, 127. <https://doi.org/10.1542/peds.2010-3054>
- Bhutta, A. T., Cleves, M. A., Casey, P. H., Cradock, M. M., & Anand, K. J. S. (2002). Cognitive and Behavioral Outcomes of School-Aged Children Who Were Born Preterm. *JAMA*, 288(6), 728. <https://doi.org/10.1001/jama.288.6.728>
- Bikle, D. D. (2014). Vitamin D metabolism, mechanism of action, and clinical applications. *Chemistry and Biology*, 21(3), 319–329. <https://doi.org/10.1016/j.chembiol.2013.12.016>
- Bischoff-Ferrari, H. A., Borchers, M., Gudat, F., Dürmüller, U., Stähelin, H. B., & Dick, W. (2004). Vitamin D Receptor Expression in Human Muscle Tissue Decreases with Age. *Journal of Bone and Mineral Research*, 19(2), 265–269. <https://doi.org/10.1359/jbmr.2004.19.2.265>
- Bischoff-Ferrari, H. A., Dietrich, T., Orav, E. J., Hu, F. B., Zhang, Y., Karlson, E. W., & Dawson-Hughes, B. (2004). Higher 25-hydroxyvitamin D concentrations are associated with better lower-extremity function in both active and inactive persons aged ≥ 60 y. *The American Journal of Clinical Nutrition*, 80(3), 752–758. <https://doi.org/10.1093/ajcn/80.3.752>
- Black, R. E. (2015). Global Prevalence of Small for Gestational Age Births. In *Nestle Nutrition Institute Workshop Series* (Vol. 81, pp. 1–7). S. Karger AG.

<https://doi.org/10.1159/000365790>

Blencowe, H., Cousens, S., Oestergaard, M. Z., Chou, D., Moller, A.-B., Narwal, R., Adler, A., Vera Garcia, C., Rohde, S., Say, L., & Lawn, J. E. (2012). National, regional, and worldwide estimates of preterm birth rates in the year 2010 with time trends since 1990 for selected countries: a systematic analysis and implications. *The Lancet*, 379(9832), 2162–2172. [https://doi.org/10.1016/S0140-6736\(12\)60820-4](https://doi.org/10.1016/S0140-6736(12)60820-4)

Bodnar, L. M., Catov, J. M., Simhan, H. N., Holick, M. F., Powers, R. W., & Roberts, J. M. (2007). Maternal Vitamin D Deficiency Increases the Risk of Preeclampsia. *The Journal of Clinical Endocrinology & Metabolism*, 92(9), 3517–3522. <https://doi.org/10.1210/jc.2007-0718>

Bornstedt, M. E., Gjerlaugsen, N., Pepaj, M., Bredahl, M. K. L., & Thorsby, P. M. (2019). Vitamin D Increases Glucose Stimulated Insulin Secretion from Insulin Producing Beta Cells (INS1E). *International Journal of Endocrinology and Metabolism, In Press*(In Press). <https://doi.org/10.5812/ijem.74255>

Brandenburg, J., Vrijkotte, T. G., & Goedhart, G. (2012). Maternal early-pregnancy vitamin D status is associated with maternal depressive symptoms in the Amsterdam Born Children and their Development cohort. *Psychosomatic Medicine*, 74. <https://doi.org/10.1097/PSY.0b013e3182639fdb>

Brenza, H. L., & DeLuca, H. F. (2000). Regulation of 25-Hydroxyvitamin D3 1 α -Hydroxylase Gene Expression by Parathyroid Hormone and 1,25-Dihydroxyvitamin D3. *Archives of Biochemistry and Biophysics*, 381(1), 143–152. <https://doi.org/10.1006/abbi.2000.1970>

Broe, K. E., Chen, T. C., Weinberg, J., Bischoff-Ferrari, H. A., Holick, M. F., & Kiel, D. P. (2007). A Higher Dose of Vitamin D Reduces the Risk of Falls in Nursing Home Residents: A Randomized, Multiple-Dose Study. *Journal of the American Geriatrics*

Society, 55(2), 234–239. <https://doi.org/10.1111/j.1532-5415.2007.01048.x>

Brown, E. M., Gamba, G., Riccardi, D., Lombardi, M., Butters, R., Kifor, O., Sun, A., Hediger, M. A., Lytton, J., & Hebert, S. C. (1993). Cloning and characterization of an extracellular Ca²⁺-sensing receptor from bovine parathyroid. *Nature*, 366(6455), 575–580. <https://doi.org/10.1038/366575a0>

Buhimschi, I. A., Buhimschi, C. S., & Weiner, C. P. (2003). Protective effect of N-acetylcysteine against fetal death and preterm labor induced by maternal inflammation. *American Journal of Obstetrics and Gynecology*, 188(1), 203–208. <https://doi.org/10.1067/mob.2003.112>

Burris, H. H., Rifas-Shiman, S. L., Camargo, C. A., Litonjua, A. A., Huh, S. Y., Rich-Edwards, J. W., & Gillman, M. W. (2012). Plasma 25-hydroxyvitamin D during pregnancy and small-for-gestational age in black and white infants. *Annals of Epidemiology*, 22(8), 581–586. <https://doi.org/10.1016/j.annepidem.2012.04.015>

Camargo, C. A., Ingham, T., & Wickens, K. (2011). Cord-blood 25-hydroxyvitamin D levels and risk of respiratory infection, wheezing, and asthma. *Pediatrics*, 127. <https://doi.org/10.1542/peds.2010-0442>

Camargo, C. A., Rifas-Shiman, S. L., Litonjua, A. A., Rich-Edwards, J. W., Weiss, S. T., Gold, D. R., Kleinman, K., & Gillman, M. W. (2007). Maternal intake of vitamin D during pregnancy and risk of recurrent wheeze in children at 3 y of age. *American Journal of Clinical Nutrition*, 85(3), 788–795. <https://doi.org/10.1093/ajcn/85.3.788>

Cantorna, M. T., Hayes, C. E., & DeLuca, H. F. (1998). 1,25-Dihydroxycholecalciferol Inhibits the Progression of Arthritis in Murine Models of Human Arthritis. *The Journal of Nutrition*, 128(1), 68–72. <https://doi.org/10.1093/jn/128.1.68>

- Carless, M. A., Kraska, T., Lintell, N., Neale, R. E., Green, A. C., & Griffiths, L. R. (2008). Polymorphisms of the VDR gene are associated with presence of solar keratoses on the skin. *British Journal of Dermatology*, *159*(4), 804–810. <https://doi.org/10.1111/j.1365-2133.2008.08745.x>
- Cassidy-Bushrow, A. E., Peters, R. M., & Johnson, D. A. (2012). Vitamin D nutritional status and antenatal depressive symptoms in African American women. *Journal of Women's Health*, *21*. <https://doi.org/10.1089/jwh.2012.3528>
- Chan, T. Y. K. (2000). Vitamin D Deficiency and Susceptibility to Tuberculosis. *Calcified Tissue International*, *66*(6), 476–478. <https://doi.org/10.1007/s002230010095>
- Chapuy, M. C., Schott, A. M., Garnero, P., Hans, D., Delmas, P. D., & Meunier, P. J. (1996). Healthy elderly French women living at home have secondary hyperparathyroidism and high bone turnover in winter. EPIDOS Study Group. *The Journal of Clinical Endocrinology & Metabolism*, *81*(3), 1129–1133. <https://doi.org/10.1210/jcem.81.3.8772587>
- Chiavaroli, V., Castorani, V., Guidone, P., Derraik, J. G. B., Liberati, M., Chiarelli, F., & Mohn, A. (2016). Incidence of infants born small- and large-for-gestational-age in an Italian cohort over a 20-year period and associated risk factors. *Italian Journal of Pediatrics*, *42*(1), 42. <https://doi.org/10.1186/s13052-016-0254-7>
- Course, C., & Chakraborty, M. (2020). Management of Respiratory Distress Syndrome in Preterm Infants In Wales: A Full Audit Cycle of a Quality Improvement Project. *Scientific Reports*, *10*(1), 3536. <https://doi.org/10.1038/s41598-020-60091-6>
- Crider, K. S., Whitehead, N., & Buus, R. M. (2005). Genetic variation associated with preterm birth: A HuGE review. *Genetics in Medicine*, *7*(9), 593–604. <https://doi.org/10.1097/01.gim.0000187223.69947.db>

- Crump, C., Sundquist, K., Sundquist, J., & Winkleby, M. A. (2011). Gestational age at birth and mortality in young adulthood. *JAMA*, *306*(11), 1233–1240. <https://doi.org/10.1001/jama.2011.1331>
- Darwish, H. M., & DeLuca, H. F. (1999). Identification of a Transcription Factor That Binds to the Promoter Region of the Human Parathyroid Hormone Gene. *Archives of Biochemistry and Biophysics*, *365*(1), 123–130. <https://doi.org/10.1006/abbi.1999.1160>
- Dawson-Hughes, B., Heaney, R. P., Holick, M. F., Lips, P., Meunier, P. J., & Vieth, R. (2005). Estimates of optimal vitamin D status. *Osteoporosis International*, *16*(7), 713–716. <https://doi.org/10.1007/s00198-005-1867-7>
- DeLuca, H. F. (2004). Overview of general physiologic features and functions of vitamin D. *The American Journal of Clinical Nutrition*, *80*(6), 1689S–1696S. <https://doi.org/10.1093/ajcn/80.6.1689S>
- den Dunnen, J. T., Dalgleish, R., Maglott, D. R., Hart, R. K., Greenblatt, M. S., McGowan-Jordan, J., Roux, A.-F., Smith, T., Antonarakis, S. E., & Taschner, P. E. M. (2016). HGVS Recommendations for the Description of Sequence Variants: 2016 Update. *Human Mutation*, *37*(6), 564–569. <https://doi.org/10.1002/humu.22981>
- Dutra, L. V., Affonso-Kaufman, F. A., Cafeo, F. R., Kassai, M. S., Barbosa, C. P., Santos Figueiredo, F. W., Suano-Souza, F. I., & Bianco, B. (2020). Association between vitamin D plasma concentrations and VDR gene variants and the risk of premature birth. *BMC Pregnancy and Childbirth*, *20*(1), 3. <https://doi.org/10.1186/s12884-019-2671-2>
- Eckhardt, C. L., Gernand, A. D., Roth, D. E., & Bodnar, L. M. (2015). Maternal vitamin D status and infant anthropometry in a US multi-centre cohort study. *Annals of Human Biology*, *42*(3), 217–224. <https://doi.org/10.3109/03014460.2014.954616>

- Eichenwald, E. C., & Stark, A. R. (2008). Management and Outcomes of Very Low Birth Weight. *New England Journal of Medicine*, 358(16), 1700–1711. <https://doi.org/10.1056/NEJMra0707601>
- Embry, A. F., Snowdon, L. R., & Vieth, R. (2000). Vitamin D and seasonal fluctuations of gadolinium-enhancing magnetic resonance imaging lesions in multiple sclerosis. *Annals of Neurology*, 48(2), 271–272. <http://www.ncbi.nlm.nih.gov/pubmed/10939587>
- Emmerson, A. J. B., Dockery, K. E., Mughal, M. Z., Roberts, S. A., Tower, C. L., & Berry, J. L. (2018). Vitamin D status of White pregnant women and infants at birth and 4 months in North West England: A cohort study. *Maternal & Child Nutrition*, 14(1), e12453. <https://doi.org/10.1111/mcn.12453>
- Evans, K. N., Bulmer, J. N., Kilby, M. D., & Hewison, M. (2004). Vitamin D and placental-decidual function. *Journal of the Society for Gynecologic Investigation*, 11(5), 263–271. <https://doi.org/10.1016/j.jsgi.2004.02.002>
- Evans, K. N., Nguyen, L., Chan, J., Innes, B. A., Bulmer, J. N., Kilby, M. D., & Hewison, M. (2006). Effects of 25-hydroxyvitamin D3 and 1,25-dihydroxyvitamin D3 on cytokine production by human decidual cells. *Biology of Reproduction*, 75(6), 816–822. <https://doi.org/10.1095/biolreprod.106.054056>
- Fanaroff, A. A., Stoll, B. J., Wright, L. L., Carlo, W. A., Ehrenkranz, R. A., Stark, A. R., Bauer, C. R., Donovan, E. F., Korones, S. B., Laptook, A. R., Lemons, J. A., Oh, W., Papile, L.-A., Shankaran, S., Stevenson, D. K., Tyson, J. E., & Poole, W. K. (2007). Trends in neonatal morbidity and mortality for very low birthweight infants. *American Journal of Obstetrics and Gynecology*, 196(2), 147.e1-147.e8. <https://doi.org/10.1016/j.ajog.2006.09.014>
- Fernandez-Alonso, A. M., Dionis-Sanchez, E. C., & Chedraui, P. (2012). First-trimester

maternal serum 25-hydroxyvitamin D(3) status and pregnancy outcome. *International Journal of Gynaecology and Obstetrics: The Official Organ of the International Federation of Gynaecology and Obstetrics*, 116. <https://doi.org/10.1016/j.ijgo.2011.07.029>

Ferrari, S., Manen, D., Bonjour, J.-P., Slosman, D., & Rizzoli, R. (1999). Bone Mineral Mass and Calcium and Phosphate Metabolism in Young Men: Relationships with Vitamin D Receptor Allelic Polymorphisms¹. *The Journal of Clinical Endocrinology & Metabolism*, 84(6), 2043–2048. <https://doi.org/10.1210/jcem.84.6.5790>

Finken, M. J. J., Schrevel, M., Houwing-Duistermaat, J. J., Kharagjitsingh, A. V., Dekker, F. W., Koeleman, B. P., Roep, B. O., & Wit, J. M. (2016). Vitamin D receptor polymorphisms and growth until adulthood after very premature birth. *Journal of Bone and Mineral Metabolism*, 34(5), 564–570. <https://doi.org/10.1007/s00774-015-0697-8>

Fiscaletti, M., Stewart, P., & Munns, C. F. (2017). The importance of vitamin D in maternal and child health: a global perspective. *Public Health Reviews*, 38, 19. <https://doi.org/10.1186/s40985-017-0066-3>

Food and Nutrition Board Institute of Medicine. (1997). *Dietary Reference Intakes for Calcium, Phosphorus, Magnesium, Vitamin D, and Fluoride*. National Academies Press. <https://doi.org/10.17226/5776>

Gale, C. R., Robinson, S. M., Harvey, N. C., Javaid, M. K., Jiang, B., Martyn, C. N., Godfrey, K. M., & Cooper, C. (2008). Maternal vitamin D status during pregnancy and child outcomes. *European Journal of Clinical Nutrition*, 62(1), 68–77. <https://doi.org/10.1038/sj.ejcn.1602680>

Garabedian, M., Holick, M. F., Deluca, H. F., & Boyle, I. T. (1972). Control of 25-Hydroxycholecalciferol Metabolism by Parathyroid Glands. *Proceedings of the National*

Academy of Sciences, 69(7), 1673–1676. <https://doi.org/10.1073/pnas.69.7.1673>

GARABEDIAN, M., TANAKA, Y., HOLICK, M. F., & DELUCA, H. F. (1974). Response of Intestinal Calcium Transport and Bone Calcium Mobilization to 1,25-Dihydroxyvitamin D_a in Thyroparathyroidectomized Rats 1. *Endocrinology*, 94(4), 1022–1027. <https://doi.org/10.1210/endo-94-4-1022>

Garland, C. F., Garland, F. C., Gorham, E. D., Lipkin, M., Newmark, H., Mohr, S. B., & Holick, M. F. (2006). The Role of Vitamin D in Cancer Prevention. *American Journal of Public Health*, 96(2), 252–261. <https://doi.org/10.2105/AJPH.2004.045260>

Gašparović Krpina, M., Barišić, A., Peterlin, A., Tul, N., Ostojić, S., Peterlin, B., & Pereza, N. (2020). Vitamin D receptor polymorphisms in spontaneous preterm birth: a case-control study. *Croatian Medical Journal*, 61(4), 338–345. <https://doi.org/10.3325/cmj.2020.61.338>

Gennari, L., Becherini, L., Masi, L., Gonnelli, S., Cepollaro, C., Martini, S., Mansani, R., & Brandi, M. L. (1997). Vitamin D receptor genotypes and intestinal calcium absorption in postmenopausal women. *Calcified Tissue International*, 61(6), 460–463. <https://doi.org/10.1007/s002239900368>

Gernand, A. D., Simhan, H. N., Klebanoff, M. A., & Bodnar, L. M. (2013). Maternal Serum 25-Hydroxyvitamin D and Measures of Newborn and Placental Weight in a U.S. Multicenter Cohort Study. *The Journal of Clinical Endocrinology & Metabolism*, 98(1), 398–404. <https://doi.org/10.1210/jc.2012-3275>

Ginde, A. A., Sullivan, A. F., Mansbach, J. M., & Camargo, C. A. (2010). Vitamin D insufficiency in pregnant and nonpregnant women of childbearing age in the United States. *American Journal of Obstetrics and Gynecology*, 202(5), 436.e1-436.e8. <https://doi.org/10.1016/j.ajog.2009.11.036>

- Giovannucci, E., Liu, Y., Rimm, E. B., Hollis, B. W., Fuchs, C. S., Stampfer, M. J., & Willett, W. C. (2006). Prospective Study of Predictors of Vitamin D Status and Cancer Incidence and Mortality in Men. *JNCI: Journal of the National Cancer Institute*, 98(7), 451–459. <https://doi.org/10.1093/jnci/djj101>
- Gloth, F. M., Alam, W., & Hollis, B. (1999). Vitamin D vs broad spectrum phototherapy in the treatment of seasonal affective disorder. *The Journal of Nutrition, Health & Aging*, 3(1), 5–7. <http://www.ncbi.nlm.nih.gov/pubmed/10888476>
- Goedicke-Fritz, S., Härtel, C., Krasteva-Christ, G., Kopp, M. V., Meyer, S., & Zemlin, M. (2017). Preterm Birth Affects the Risk of Developing Immune-Mediated Diseases. *Frontiers in Immunology*, 8. <https://doi.org/10.3389/fimmu.2017.01266>
- Goldenberg, R. L., & Culhane, J. F. (2007). Low birth weight in the United States. *The American Journal of Clinical Nutrition*, 85(2), 584S-590S. <https://doi.org/10.1093/ajcn/85.2.584S>
- Goldenberg, R. L., Culhane, J. F., Iams, J. D., & Romero, R. (2008). Epidemiology and causes of preterm birth. *The Lancet*, 371(9606), 75–84. [https://doi.org/10.1016/S0140-6736\(08\)60074-4](https://doi.org/10.1016/S0140-6736(08)60074-4)
- Grant, W. B. (2002). An estimate of premature cancer mortality in the U.S. due to inadequate doses of solar ultraviolet-B radiation. *Cancer*, 94(6), 1867–1875. <https://doi.org/10.1002/cncr.10427>
- Grant, W. B., & Garland, C. F. (2006). The association of solar ultraviolet B (UVB) with reducing risk of cancer: multifactorial ecologic analysis of geographic variation in age-adjusted cancer mortality rates. *Anticancer Research*, 26(4A), 2687–2699. <http://www.ncbi.nlm.nih.gov/pubmed/16886679>

- Griffin, M. D., Lutz, W., Phan, V. A., Bachman, L. A., McKean, D. J., & Kumar, R. (2001). Dendritic cell modulation by $1\alpha,25$ dihydroxyvitamin D₃ and its analogs: A vitamin D receptor-dependent pathway that promotes a persistent state of immaturity in vitro and in vivo. *Proceedings of the National Academy of Sciences*, *98*(12), 6800–6805. <https://doi.org/10.1073/pnas.121172198>
- Grisaru-Granovsky, S., Tevet, A., Bar-Shavit, R., Salah, Z., Elstein, D., Samueloff, A., & Altarescu, G. (2007). Association study of protease activated receptor 1 gene polymorphisms and adverse pregnancy outcomes: Results of a pilot study in Israel. *American Journal of Medical Genetics Part A*, *143A*(21), 2557–2563. <https://doi.org/10.1002/ajmg.a.31985>
- Gysemans, C. A., Cardozo, A. K., Callewaert, H., Giulietti, A., Hulshagen, L., Bouillon, R., Eizirik, D. L., & Mathieu, C. (2005). $1,25$ -Dihydroxyvitamin D₃ modulates expression of chemokines and cytokines in pancreatic islets: implications for prevention of diabetes in nonobese diabetic mice. *Endocrinology*, *146*(4), 1956–1964. <https://doi.org/10.1210/en.2004-1322>
- Hack, M., Taylor, H. G., Drotar, D., Schluchter, M., Cartar, L., Andreias, L., Wilson-Costello, D., & Klein, N. (2005). Chronic Conditions, Functional Limitations, and Special Health Care Needs of School-aged Children Born With Extremely Low-Birth-Weight in the 1990s. *JAMA*, *294*(3), 318. <https://doi.org/10.1001/jama.294.3.318>
- Hall, E. S., & Greenberg, J. M. (2016). Estimating community-level costs of preterm birth. *Public Health*, *141*, 222–228. <https://doi.org/10.1016/j.puhe.2016.09.033>
- Hathcock, J. N., Shao, A., Vieth, R., & Heaney, R. (2007). Risk assessment for vitamin D. *The American Journal of Clinical Nutrition*, *85*(1), 6–18. <https://doi.org/10.1093/ajcn/85.1.6>
- Hayes, C. E., Nashold, F. E., Spach, K. M., & Pedersen, L. B. (2003). The immunological

- functions of the vitamin D endocrine system. *Cellular and Molecular Biology (Noisy-Le-Grand, France)*, 49(2), 277–300. <http://www.ncbi.nlm.nih.gov/pubmed/12887108>
- Heaney, R. P., Dowell, M. S., Hale, C. A., & Bendich, A. (2003). Calcium Absorption Varies within the Reference Range for Serum 25-Hydroxyvitamin D. *Journal of the American College of Nutrition*, 22(2), 142–146. <https://doi.org/10.1080/07315724.2003.10719287>
- Helmer, H., Tretzmüller, U., Brunbauer, M., Kaider, A., Husslein, P., & Knöfler, M. (2002). Production of oxytocin receptor and cytokines in primary uterine smooth muscle cells cultivated under inflammatory conditions. *Journal of the Society for Gynecologic Investigation*, 9(1), 15–21. [https://doi.org/10.1016/s1071-5576\(01\)00142-3](https://doi.org/10.1016/s1071-5576(01)00142-3)
- Hendler, I., Goldenberg, R. L., Mercer, B. M., Iams, J. D., Meis, P. J., Moawad, A. H., MacPherson, C. A., Caritis, S. N., Miodovnik, M., Menard, K. M., Thurnau, G. R., & Sorokin, Y. (2005). The Preterm Prediction Study: association between maternal body mass index and spontaneous and indicated preterm birth. *American Journal of Obstetrics and Gynecology*, 192(3), 882–886. <https://doi.org/10.1016/j.ajog.2004.09.021>
- Hewison, M. (2011). Antibacterial effects of vitamin D. *Nature Reviews. Endocrinology*, 7(6), 337–345. <https://doi.org/10.1038/nrendo.2010.226>
- Hille, E. T., den Ouden, A. L., Saigal, S., Wolke, D., Lambert, M., Whitaker, A., Pinto-Martin, J. A., Hout, L., Meyer, R., Feldman, J. F., Verloove-Vanhorick, S. P., & Paneth, N. (2001). Behavioural problems in children who weigh 1000 g or less at birth in four countries. *The Lancet*, 357(9269), 1641–1643. [https://doi.org/10.1016/S0140-6736\(00\)04818-2](https://doi.org/10.1016/S0140-6736(00)04818-2)
- Holick, M. F. (2003). Vitamin D: A millenium perspective. *Journal of Cellular Biochemistry*, 88(2), 296–307. <https://doi.org/10.1002/jcb.10338>

- Holick, M. F. (2007). Vitamin D Deficiency. *New England Journal of Medicine*, 357(3), 266–281. <https://doi.org/10.1056/NEJMra070553>
- Holick, M. F., Binkley, N. C., Bischoff-Ferrari, H. A., Gordon, C. M., Hanley, D. A., Heaney, R. P., Murad, M. H., & Weaver, C. M. (2011). Evaluation, Treatment, and Prevention of Vitamin D Deficiency: an Endocrine Society Clinical Practice Guideline. *The Journal of Clinical Endocrinology & Metabolism*, 96(7), 1911–1930. <https://doi.org/10.1210/jc.2011-0385>
- Holick, M. F., & Chen, T. C. (2008). Vitamin D deficiency: A worldwide problem with health consequences. *American Journal of Clinical Nutrition*, 87(4), 1080–1086. <https://doi.org/10.1093/ajcn/87.4.1080s>
- Holick, M. F., Siris, E. S., Binkley, N., Beard, M. K., Khan, A., Katzer, J. T., Petruschke, R. A., Chen, E., & de Papp, A. E. (2005). Prevalence of Vitamin D Inadequacy among Postmenopausal North American Women Receiving Osteoporosis Therapy. *The Journal of Clinical Endocrinology & Metabolism*, 90(6), 3215–3224. <https://doi.org/10.1210/jc.2004-2364>
- Howson, C. P., Kinney, M. V, McDougall, L., & Lawn, J. E. (2013). Born Too Soon: Preterm birth matters. *Reproductive Health*, 10(S1), S1. <https://doi.org/10.1186/1742-4755-10-S1-S1>
- Huldschinsky, K. (1919). Heilung von Rachitis durch künstliche Höhensonne. *DMW - Deutsche Medizinische Wochenschrift*, 45(26), 712–713. <https://doi.org/10.1055/s-0028-1137830>
- Hullett, D. A., Cantorna, M. T., Redaelli, C., Humpal-Winter, J., Hayes, C. E., Sollinger, H. W., & Deluca, H. F. (1998). PROLONGATION OF ALLOGRAFT SURVIVAL BY 1,25-DIHYDROXYVITAMIN D31. *Transplantation*, 66(7), 824–828.

<https://doi.org/10.1097/00007890-199810150-00002>

Hyppönen, E., Läärä, E., Reunanen, A., Järvelin, M.-R., & Virtanen, S. M. (2001). Intake of vitamin D and risk of type 1 diabetes: a birth-cohort study. *The Lancet*, 358(9292), 1500–1503. [https://doi.org/10.1016/S0140-6736\(01\)06580-1](https://doi.org/10.1016/S0140-6736(01)06580-1)

Institute of Medicine. (1997). *Dietary reference intakes for calcium, phosphorus, magnesium, vitamin D and fluoride*. https://scholar.google.com/scholar?hl=en&as_sdt=0%2C5&q=Institute+of+Medicine.+1997.+Dietary+reference+intakes+for+calcium%2C+phosphorus%2C+magnesium%2C+vitamin+D+and+fluoride.+Washington%2C+D.C.%3A+National+Academies+Press.+p.+1-522.&btnG=

Ioannou, C., Javaid, M. K., Mahon, P., Yaqub, M. K., Harvey, N. C., Godfrey, K. M., Noble, J. A., Cooper, C., & Papageorgiou, A. T. (2012). The Effect of Maternal Vitamin D Concentration on Fetal Bone. *The Journal of Clinical Endocrinology & Metabolism*, 97(11), E2070–E2077. <https://doi.org/10.1210/jc.2012-2538>

IOM. (2007). *Preterm Birth*. National Academies Press. <https://doi.org/10.17226/11622>

Ismail, S., & Essawi, M. (2012). Genetic polymorphism studies in humans. *Middle East Journal of Medical Genetics*, 1(2), 57–63. <https://doi.org/10.1097/01.MXE.0000415225.85003.47>

Jones, A. P., Palmer, D., & Zhang, G. (2012). Cord blood 25-hydroxyvitamin D3 and allergic disease during infancy. *Pediatrics*, 130. <https://doi.org/10.1542/peds.2012-1172>

JONES, G., STRUGNELL, S. A., & DeLUCA, H. F. (1998). Current Understanding of the Molecular Actions of Vitamin D. *Physiological Reviews*, 78(4), 1193–1231. <https://doi.org/10.1152/physrev.1998.78.4.1193>

- Karim, S. A., Nusrat, U., & Aziz, S. (2011). Vitamin D deficiency in pregnant women and their newborns as seen at a tertiary-care center in Karachi, Pakistan. *International Journal of Gynecology & Obstetrics*, *112*(1), 59–62. <https://doi.org/10.1016/j.ijgo.2010.07.034>
- Karras, S. N., Anagnostis, P., Annweiler, C., Naughton, D. P., Petroczi, A., Bili, E., Harizopoulou, V., Tarlatzis, B. C., Persinaki, A., Papadopoulou, F., & Goulis, D. G. (2014). Maternal vitamin D status during pregnancy: the Mediterranean reality. *European Journal of Clinical Nutrition*, *68*(8), 864–869. <https://doi.org/10.1038/ejcn.2014.80>
- Kato, S. (2000). The function of vitamin D receptor in vitamin D action. *Journal of Biochemistry*, *127*(5), 717–722. <https://doi.org/10.1093/oxfordjournals.jbchem.a022662>
- Khatun, S., & Rahman, M. (1970). Socio-economic determinants of low birth weight in Bangladesh: A multivariate approach. *Bangladesh Medical Research Council Bulletin*, *34*(3), 81–86. <https://doi.org/10.3329/bmrcb.v34i3.1857>
- Knabl, J., Vattai, A., Ye, Y., Jueckstock, J., Hutter, S., Kainer, F., Mahner, S., & Jeschke, U. (2017). Role of placental VDR expression and function in common late pregnancy disorders. *International Journal of Molecular Sciences*, *18*(11), 2340. <https://doi.org/10.3390/ijms18112340>
- Kosik, K., Szpecht, D., Al-Saad, S. R., Karbowski, L. M., Kurzawińska, G., Szymankiewicz, M., Drews, K., Wolski, H., & Seremak-Mrozikiewicz, A. (2020). Single nucleotide vitamin D receptor polymorphisms (FokI, BsmI, ApaI, and TaqI) in the pathogenesis of prematurity complications. *Scientific Reports*, *10*(1), 21098. <https://doi.org/10.1038/s41598-020-78125-4>
- Kramer, M. S., Lydon, J., Seguin, L., Goulet, L., Kahn, S. R., McNamara, H., Genest, J., Dassa, C., Chen, M. F., Sharma, S., Meaney, M. J., Thomson, S., Van Uum, S., Koren, G., Dahhou, M., Lamoureux, J., & Platt, R. W. (2009). Stress Pathways to Spontaneous

- Preterm Birth: The Role of Stressors, Psychological Distress, and Stress Hormones. *American Journal of Epidemiology*, 169(11), 1319–1326. <https://doi.org/10.1093/aje/kwp061>
- Lawlor, D. A., Wills, A. K., Fraser, A., Sayers, A., Fraser, W. D., & Tobias, J. H. (2013). Association of maternal vitamin D status during pregnancy with bone-mineral content in offspring: a prospective cohort study. *The Lancet*, 381(9884), 2176–2183. [https://doi.org/10.1016/S0140-6736\(12\)62203-X](https://doi.org/10.1016/S0140-6736(12)62203-X)
- Lawn, J. E., Blencowe, H., Pattinson, R., Cousens, S., Kumar, R., Ibiebele, I., Gardosi, J., Day, L. T., & Stanton, C. (2011). Stillbirths: Where? When? Why? How to make the data count? *The Lancet*, 377(9775), 1448–1463. [https://doi.org/10.1016/S0140-6736\(10\)62187-3](https://doi.org/10.1016/S0140-6736(10)62187-3)
- Lawson, C. C., Whelan, E. A., Hibert, E. N., Grajewski, B., Spiegelman, D., & Rich-Edwards, J. W. (2009). Occupational factors and risk of preterm birth in nurses. *American Journal of Obstetrics and Gynecology*, 200(1), 51.e1-51.e8. <https://doi.org/10.1016/j.ajog.2008.08.006>
- Layana, A., Minnella, A., Garhöfer, G., Aslam, T., Holz, F., Leys, A., Silva, R., Delcourt, C., Souied, E., & Seddon, J. (2017). Vitamin D and Age-Related Macular Degeneration. *Nutrients*, 9(10), 1120. <https://doi.org/10.3390/nu9101120>
- Leffelaar, E. R., Vrijkotte, T. G. M., & van Eijsden, M. (2010). Maternal early pregnancy vitamin D status in relation to fetal and neonatal growth: results of the multi-ethnic Amsterdam Born Children and their Development cohort. *British Journal of Nutrition*, 104(1), 108–117. <https://doi.org/10.1017/S000711451000022X>
- Lemire, J. M., Ince, A., & Takashima, M. (1992). 1,25-Dihydroxyvitamin D₃ Attenuates of Expression of Experimental Murine Lupus of MRL/1 Mice. *Autoimmunity*, 12(2), 143–

148. <https://doi.org/10.3109/08916939209150321>

Limperopoulos, C., Bassan, H., Sullivan, N. R., Soul, J. S., Robertson, R. L., Moore, M., Ringer, S. A., Volpe, J. J., & du Plessis, A. J. (2008). Positive Screening for Autism in Ex-preterm Infants: Prevalence and Risk Factors. *Pediatrics*, *121*(4), 758–765. <https://doi.org/10.1542/peds.2007-2158>

Lips, P. (2001). Vitamin D Deficiency and Secondary Hyperparathyroidism in the Elderly: Consequences for Bone Loss and Fractures and Therapeutic Implications. *Endocrine Reviews*, *22*(4), 477–501. <https://doi.org/10.1210/edrv.22.4.0437>

Litt, J., Taylor, H. G., Klein, N., & Hack, M. (2005). Learning disabilities in children with very low birthweight: prevalence, neuropsychological correlates, and educational interventions. *Journal of Learning Disabilities*, *38*(2), 130–141. <https://doi.org/10.1177/00222194050380020301>

Liu, J. (2012). Vitamin D content of food and its contribution to vitamin D status: a brief overview and Australian focus. *Photochemical & Photobiological Sciences*, *11*(12), 1802–1807. <https://doi.org/10.1039/c2pp25150g>

Liu, N., Kaplan, A. T., Low, J., Nguyen, L., Liu, G. Y., Equils, O., & Hewison, M. (2009). Vitamin D Induces Innate Antibacterial Responses in Human Trophoblasts via an Intracrine Pathway¹. *Biology of Reproduction*, *80*(3), 398–406. <https://doi.org/10.1095/biolreprod.108.073577>

Liu, N. Q., & Hewison, M. (2012). Vitamin D, the placenta and pregnancy. *Archives of Biochemistry and Biophysics*, *523*(1), 37–47. <https://doi.org/10.1016/j.abb.2011.11.018>

Liu, N. Q., Kaplan, A. T., Lagishetty, V., Ouyang, Y. B., Ouyang, Y., Simmons, C. F., Equils, O., & Hewison, M. (2011). Vitamin D and the regulation of placental inflammation.

Journal of Immunology (Baltimore, Md. : 1950), 186(10), 5968–5974.
<https://doi.org/10.4049/jimmunol.1003332>

Lorentzon, M., Lorentzon, R., & Nordström, P. (2000). Vitamin D receptor gene polymorphism is associated with birth height, growth to adolescence, and adult stature in healthy caucasian men: a cross-sectional and longitudinal study. *The Journal of Clinical Endocrinology and Metabolism*, 85(4), 1666–1670.
<https://doi.org/10.1210/jcem.85.4.6566>

MacLaughlin, J. A., Anderson, R. R., & Holick, M. F. (1982). Spectral Character of Sunlight Modulates Photosynthesis of Previtamin D₃ and Its Photoisomers in Human Skin. *Science*, 216(4549), 1001–1003. <https://doi.org/10.1126/science.6281884>

Malabanan, A., Veronikis, I., & Holick, M. (1998). Redefining vitamin D insufficiency. *The Lancet*, 351(9105), 805–806. [https://doi.org/10.1016/S0140-6736\(05\)78933-9](https://doi.org/10.1016/S0140-6736(05)78933-9)

Mangin, M., Sinha, R., & Fincher, K. (2014). Inflammation and vitamin D: the infection connection. *Inflammation Research: Official Journal of the European Histamine Research Society ... [et Al.]*, 63(10), 803–819. <https://doi.org/10.1007/s00011-014-0755-z>

Manzon, L., Altarescu, G., Tevet, A., Schimmel, M. S., Elstein, D., Samueloff, A., & Grisaru-Granovsky, S. (2014). Vitamin D receptor polymorphism FokI is associated with spontaneous idiopathic preterm birth in an Israeli population. *European Journal of Obstetrics & Gynecology and Reproductive Biology*, 177, 84–88.
<https://doi.org/10.1016/j.ejogrb.2014.03.008>

Marco, M. P., Martínez, I., Amoedo, M. L., Borràs, M., Saracho, R., Almirall, J., Fibla, J., & Fernández, E. (1999). Vitamin D receptor genotype influences parathyroid hormone and calcitriol levels in predialysis patients. *Kidney International*, 56(4), 1349–1353.

<https://doi.org/10.1046/j.1523-1755.1999.00678.x>

- Markopoulou, P., Papanikolaou, E., Analytis, A., Zoumakis, E., & Siahaidou, T. (2019). Preterm Birth as a Risk Factor for Metabolic Syndrome and Cardiovascular Disease in Adult Life: A Systematic Review and Meta-Analysis. *The Journal of Pediatrics*, *210*, 69-80.e5. <https://doi.org/10.1016/j.jpeds.2019.02.041>
- Marlow, N., Wolke, D., Bracewell, M. A., & Samara, M. (2005). Neurologic and Developmental Disability at Six Years of Age after Extremely Preterm Birth. *New England Journal of Medicine*, *352*(1), 9–19. <https://doi.org/10.1056/NEJMoa041367>
- Martin, J. A., Hamilton, B. E., & Ventura, S. J. (2001). Births: preliminary data for 2000. *National Vital Statistics Reports : From the Centers for Disease Control and Prevention, National Center for Health Statistics, National Vital Statistics System*, *49*(5), 1–20. <http://www.ncbi.nlm.nih.gov/pubmed/11499334>
- McGrath, J. (2002). Long-term trends in sunshine duration and its association with schizophrenia birth rates and age at first registration — data from Australia and the Netherlands. *Schizophrenia Research*, *54*(3), 199–212. [https://doi.org/10.1016/S0920-9964\(01\)00259-6](https://doi.org/10.1016/S0920-9964(01)00259-6)
- Merewood, A., Mehta, S. D., Chen, T. C., Bauchner, H., & Holick, M. F. (2009). Association between Vitamin D Deficiency and Primary Cesarean Section. *The Journal of Clinical Endocrinology & Metabolism*, *94*(3), 940–945. <https://doi.org/10.1210/jc.2008-1217>
- Michalowicz, B. S., Novak, M. J., Hodges, J. S., DiAngelis, A., Buchanan, W., Papapanou, P. N., Mitchell, D. A., Ferguson, J. E., Lupo, V., Bofill, J., Matseoane, S., Steffen, M., & Ebersole, J. L. (2009). Serum Inflammatory Mediators in Pregnancy: Changes After Periodontal Treatment and Association With Pregnancy Outcomes. *Journal of Periodontology*, *80*(11), 1731–1741. <https://doi.org/10.1902/jop.2009.090236>

- Miettinen, M. E., Reinert, L., & Kinnunen, L. (2012). Serum 25-hydroxyvitamin D level during early pregnancy and type 1 diabetes risk in the offspring. *Diabetologia*, *55*.
<https://doi.org/10.1007/s00125-012-2458-8>
- Miliku, K., Vinkhuyzen, A., Blanken, L. M., McGrath, J. J., Eyles, D. W., Burne, T. H., Hofman, A., Tiemeier, H., Steegers, E. A., Gaillard, R., & Jaddoe, V. W. (2016). Maternal vitamin D concentrations during pregnancy, fetal growth patterns, and risks of adverse birth outcomes. *The American Journal of Clinical Nutrition*, *103*(6), 1514–1522.
<https://doi.org/10.3945/ajcn.115.123752>
- Mirzaei, F., Michels, K. B., & Munger, K. (2011). Gestational vitamin D and the risk of multiple sclerosis in offspring. *Annals of Neurology*, *70*.
<https://doi.org/10.1002/ana.22456>
- Møller, U. K., Strem, S., Mosekilde, L., Heickendorff, L., Flyvbjerg, A., Frystyk, J., Jensen, L. T., & Rejnmark, L. (2013). Changes in calcitropic hormones, bone markers and insulin-like growth factor I (IGF-I) during pregnancy and postpartum: a controlled cohort study. *Osteoporosis International*, *24*(4), 1307–1320. <https://doi.org/10.1007/s00198-012-2062-2>
- Momentti, A. C., Estadella, D., & Pellegrini Pisani, L. (2018). Role of vitamin D in pregnancy and Toll-like receptor pathway. *Steroids*, *137*, 22–29.
<https://doi.org/10.1016/j.steroids.2018.07.009>
- Morales, E., Guxens, M., & Llop, S. (2012). Circulating 25-hydroxyvitamin D3 in pregnancy and infant neuropsychological development. *Pediatrics*, *130*.
<https://doi.org/10.1542/peds.2011-3289>
- Morley, R., Carlin, J. B., Pasco, J. A., & Wark, J. D. (2006). Maternal 25-Hydroxyvitamin D and Parathyroid Hormone Concentrations and Offspring Birth Size. *The Journal of*

Clinical Endocrinology & Metabolism, 91(3), 906–912. <https://doi.org/10.1210/jc.2005-1479>

Morley, R., Carlin, J. B., Pasco, J. A., Wark, J. D., & Ponsonby, A.-L. (2009). Maternal 25-hydroxyvitamin D concentration and offspring birth size: effect modification by infant VDR genotype. *European Journal of Clinical Nutrition*, 63(6), 802–804. <https://doi.org/10.1038/ejcn.2008.55>

Morrison, N. A., Qi, J. C., Tokita, A., Kelly, P. J., Crofts, L., Nguyen, T. V., Sambrook, P. N., & Eisman, J. A. (1994). Prediction of bone density from vitamin D receptor alleles. *Nature*, 367(6460), 284–287. <https://doi.org/10.1038/367284a0>

Mory, D. B., Rocco, E. R., Miranda, W. L., Kasamatsu, T., Crispim, F., & Dib, S. A. (2009). Prevalence of vitamin D receptor gene polymorphisms FokI and BsmI in Brazilian individuals with type 1 diabetes and their relation to beta-cell autoimmunity and to remaining beta-cell function. *Human Immunology*, 70(6), 447–451. <https://doi.org/10.1016/j.humimm.2009.03.007>

Moster, D., Lie, R. T., & Markestad, T. (2008). Long-Term Medical and Social Consequences of Preterm Birth. *New England Journal of Medicine*, 359(3), 262–273. <https://doi.org/10.1056/NEJMoa0706475>

Motohashi, Y., Yamada, S., Yanagawa, T., Maruyama, T., Suzuki, R., Niino, M., Fukazawa, T., Kasuga, A., Hirose, H., Matsubara, K., Shimada, A., & Saruta, T. (2003). Vitamin D receptor gene polymorphism affects onset pattern of type 1 diabetes. *The Journal of Clinical Endocrinology and Metabolism*, 88(7), 3137–3140. <https://doi.org/10.1210/jc.2002-021881>

Munger, K. L., Zhang, S. M., O'Reilly, E., Hernan, M. A., Olek, M. J., Willett, W. C., & Ascherio, A. (2004). Vitamin D intake and incidence of multiple sclerosis. *Neurology*,

62(1), 60–65. <https://doi.org/10.1212/01.WNL.0000101723.79681.38>

Murray, C. J. L., Vos, T., Lozano, R., Naghavi, M., Flaxman, A. D., Michaud, C., Ezzati, M., Shibuya, K., Salomon, J. A., Abdalla, S., Aboyans, V., Abraham, J., Ackerman, I., Aggarwal, R., Ahn, S. Y., Ali, M. K., AlMazroa, M. A., Alvarado, M., Anderson, H. R., ... Lopez, A. D. (2012). Disability-adjusted life years (DALYs) for 291 diseases and injuries in 21 regions, 1990–2010: a systematic analysis for the Global Burden of Disease Study 2010. *The Lancet*, 380(9859), 2197–2223. [https://doi.org/10.1016/S0140-6736\(12\)61689-4](https://doi.org/10.1016/S0140-6736(12)61689-4)

Nahar, N., Afroza, S., & Hossain, M. (1998). Incidence of low birth weight in three selected communities of Bangladesh. *Bangladesh Medical Research Council Bulletin*, 24(2), 49–54. <http://www.ncbi.nlm.nih.gov/pubmed/9926483>

Nair, R., & Maseeh, A. (2012). Vitamin D: The “sunshine” vitamin. *Journal of Pharmacology & Pharmacotherapeutics*, 3(2), 118–126. <https://doi.org/10.4103/0976-500X.95506>

Nesby-O’Dell, S., Scanlon, K. S., Cogswell, M. E., Gillespie, C., Hollis, B. W., Looker, A. C., Allen, C., Dougherty, C., Gunter, E. W., & Bowman, B. A. (2002). Hypovitaminosis D prevalence and determinants among African American and white women of reproductive age: third National Health and Nutrition Examination Survey, 1988–1994. *The American Journal of Clinical Nutrition*, 76(1), 187–192. <https://doi.org/10.1093/ajcn/76.1.187>

Ni, W., Watts, S. W., Ng, M., Chen, S., Glenn, D. J., & Gardner, D. G. (2014). Elimination of vitamin D receptor in vascular endothelial cells alters vascular function. *Hypertension (Dallas, Tex. : 1979)*, 64(6), 1290–1298. <https://doi.org/10.1161/HYPERTENSIONAHA.114.03971>

Palacios, C., & Gonzalez, L. (2014). Is vitamin D deficiency a major global public health problem? *The Journal of Steroid Biochemistry and Molecular Biology*, 144 Pt A, 138–

145. <https://doi.org/10.1016/j.jsbmb.2013.11.003>

Patterson, K. Y., Phillips, K. M., Horst, R. L., Byrdwell, W. C., Exler, J., Lemar, L. E., & Holden, J. M. (2010). Vitamin D content and variability in fluid milks from a US Department of Agriculture nationwide sampling to update values in the National Nutrient Database for Standard Reference. *Journal of Dairy Science*, *93*(11), 5082–5090. <https://doi.org/10.3168/jds.2010-3359>

Pike, J. W., & Meyer, M. B. (2012). The Vitamin D Receptor: New Paradigms for the Regulation of Gene Expression by 1,25-Dihydroxyvitamin D₃. *Rheumatic Disease Clinics of North America*, *38*(1), 13–27. <https://doi.org/10.1016/j.rdc.2012.03.004>

Ponsonby, A.-L., McMichael, A., & van der Mei, I. (2002). Ultraviolet radiation and autoimmune disease: insights from epidemiological research. *Toxicology*, *181–182*, 71–78. [https://doi.org/10.1016/S0300-483X\(02\)00257-3](https://doi.org/10.1016/S0300-483X(02)00257-3)

Pospechova, K., Rozehnal, V., Stejskalova, L., Vrzal, R., Pospisilova, N., Jamborova, G., May, K., Siegmund, W., Dvorak, Z., Nachtigal, P., Semecky, V., & Pavek, P. (2009). Expression and activity of vitamin D receptor in the human placenta and in choriocarcinoma BeWo and JEG-3 cell lines. *Molecular and Cellular Endocrinology*, *299*(2), 178–187. <https://doi.org/10.1016/j.mce.2008.12.003>

Powe, C. E., Seely, E. W., & Rana, S. (2010). First trimester vitamin D, vitamin D binding protein, and subsequent preeclampsia. *Hypertension*, *56*. <https://doi.org/10.1161/HYPERTENSIONAHA.110.158238>

Prentice, A. (2008). Vitamin D deficiency: a global perspective. *Nutrition Reviews*, *66*(10 Suppl 2), S153-64. <https://doi.org/10.1111/j.1753-4887.2008.00100.x>

PubChem. (2019). *Cholecalciferol* | *C27H44O* | *CID 5280795* - *PubChem*. The National

Center for Biotechnology Information.

<https://pubchem.ncbi.nlm.nih.gov/compound/Cholecalciferol>

PubChem. (2023a). *Calcitriol* | C27H44O3 | CID 5280453. The National Center for Biotechnology Information. <https://pubchem.ncbi.nlm.nih.gov/compound/Calcitriol>

PubChem. (2023b). *Ergocalciferol* | C28H44O | CID 5280793. The National Center for Biotechnology Information. <https://pubchem.ncbi.nlm.nih.gov/compound/Ergocalciferol>

PubChem. (2023c). *Maxacalcitol* | C26H42O4 | CID 6398761. The National Center for Biotechnology Information. <https://pubchem.ncbi.nlm.nih.gov/compound/Maxacalcitol>

Qin, L.-L., Lu, F.-G., Yang, S.-H., Xu, H.-L., & Luo, B.-A. (2016). Does Maternal Vitamin D Deficiency Increase the Risk of Preterm Birth: A Meta-Analysis of Observational Studies. *Nutrients*, 8(5), 301. <https://doi.org/10.3390/nu8050301>

Raghupathy, R. (2001). Pregnancy: success and failure within the Th1/Th2/Th3 paradigm. *Seminars in Immunology*, 13(4), 219–227. <https://doi.org/10.1006/smim.2001.0316>

Rahmannezhad, G., Mashayekhi, F. J., Goodarzi, M. T., Rezvanfar, M. R., & Sadeghi, A. (2016). Association between vitamin D receptor ApaI and TaqI gene polymorphisms and gestational diabetes mellitus in an Iranian pregnant women population. *Gene*, 581(1), 43–47. <https://doi.org/10.1016/j.gene.2016.01.026>

Rai, V. (2014). Methylenetetrahydrofolate reductase gene A1298C polymorphism and susceptibility to recurrent pregnancy loss: a meta-analysis. *Cellular and Molecular Biology (Noisy-Le-Grand, France)*, 60(2), 27–34. <http://www.ncbi.nlm.nih.gov/pubmed/24970119>

Randis, T. M. (2008). Complications associated with premature birth. *Virtual Mentor*, 10(10), 647–650. <https://doi.org/10.1001/virtualmentor.2008.10.10.cpr11-0810>

- Richard E Behrman, A. S. B. (2007). *Preterm Birth*. National Academies Press.
<https://doi.org/10.17226/11622>
- Romero, R., Espinoza, J., Gonçalves, L. F., Kusanovic, J. P., Friel, L. A., & Nien, J. K. (2006). Inflammation in preterm and term labour and delivery. *Seminars in Fetal and Neonatal Medicine*, *11*(5), 317–326. <https://doi.org/10.1016/j.siny.2006.05.001>
- Romero, R., Mazor, M., & Tartakovsky, B. (1991). Systemic administration of interleukin-1 induces preterm parturition in mice. *American Journal of Obstetrics and Gynecology*, *165*(4 Pt 1), 969–971. [https://doi.org/10.1016/0002-9378\(91\)90450-6](https://doi.org/10.1016/0002-9378(91)90450-6)
- Rosen, C. J., Adams, J. S., Bikle, D. D., Black, D. M., Demay, M. B., Manson, J. E., Murad, M. H., & Kovacs, C. S. (2012). The Nonskeletal Effects of Vitamin D: An Endocrine Society Scientific Statement. *Endocrine Reviews*, *33*(3), 456–492. <https://doi.org/10.1210/er.2012-1000>
- Rosenfeld, T., Salem, H., Altarescu, G., Grisaru-Granovsky, S., Tevet, A., & Birk, R. (2017). Maternal–fetal vitamin D receptor polymorphisms significantly associated with preterm birth. *Archives of Gynecology and Obstetrics*, *296*(2), 215–222. <https://doi.org/10.1007/s00404-017-4412-y>
- Rostand, S. G. (1997). Ultraviolet Light May Contribute to Geographic and Racial Blood Pressure Differences. *Hypertension*, *30*(2), 150–156. <https://doi.org/10.1161/01.HYP.30.2.150>
- Sachan, A., Gupta, R., Das, V., Agarwal, A., Awasthi, P. K., & Bhatia, V. (2005). High prevalence of vitamin D deficiency among pregnant women and their newborns in northern India. *The American Journal of Clinical Nutrition*, *81*(5), 1060–1064. <https://doi.org/10.1093/ajcn/81.5.1060>

- Sahin, O. A., Goksen, D., Ozpinar, A., Serdar, M., & Onay, H. (2017). Association of vitamin D receptor polymorphisms and type 1 diabetes susceptibility in children: a meta-analysis. *Endocrine Connections*, 6(3), 159–171. <https://doi.org/10.1530/EC-16-0110>
- Saigal, S., Ouden, L. den, Wolke, D., Hoult, L., Paneth, N., Streiner, D. L., Whitaker, A., & Pinto-Martin, J. (2003). School-Age Outcomes in Children Who Were Extremely Low Birth Weight From Four International Population-Based Cohorts. *Pediatrics*, 112(4), 943–950. <https://doi.org/10.1542/peds.112.4.943>
- Scholl, T. O. (2005). Iron status during pregnancy: setting the stage for mother and infant. *The American Journal of Clinical Nutrition*, 81(5), 1218S-1222S. <https://doi.org/10.1093/ajcn/81.5.1218>
- Scholl, T. O., Chen, X., & Stein, P. (2012). Maternal vitamin D status and delivery by cesarean. *Nutrients*, 4. <https://doi.org/10.3390/nu4040319>
- Scientific Image and Illustration Software*. (2023). BioRender. <https://www.biorender.com/>
- Sehringer, B., Schäfer, W. R., Wetzka, B., Deppert, W. R., Brunner-Spahr, R., Benedek, E., & Zahradnik, H. P. (2000). Formation of proinflammatory cytokines in human term myometrium is stimulated by lipopolysaccharide but not by corticotropin-releasing hormone. *The Journal of Clinical Endocrinology and Metabolism*, 85(12), 4859–4865. <https://doi.org/10.1210/jcem.85.12.7006>
- Senti, J., Thiele, D. K., & Anderson, C. M. (2012). Maternal vitamin D status as a critical determinant in gestational diabetes. *Journal of Obstetric, Gynecologic, and Neonatal Nursing: JOGNN/NAACOG*, 41. <https://doi.org/10.1111/j.1552-6909.2012.01366.x>
- Shahbazi, M., Jeddi-Tehrani, M., Zareie, M., Salek-Moghaddam, A., Akhondi, M. M., Bahmanpoor, M., Sadeghi, M. R., & Zarnani, A. H. (2011). Expression profiling of

vitamin D receptor in placenta, decidua and ovary of pregnant mice. *Placenta*, 32(9), 657–664. <https://doi.org/10.1016/j.placenta.2011.06.013>

Shand, A. W., Nassar, N., & Dadelszen, P. (2010). Maternal vitamin D status in pregnancy and adverse pregnancy outcomes in a group at high risk for pre-eclampsia. *BJOG: An International Journal of Obstetrics and Gynaecology*, 117. <https://doi.org/10.1111/j.1471-0528.2010.02742.x>

Sheikh, I. A., Ahmad, E., Jamal, M. S., Rehan, M., Assidi, M., Tayubi, I. A., AlBasri, S. F., Bajouh, O. S., Turki, R. F., Abuzenadah, A. M., Damanhour, G. A., Beg, M. A., & Al-Qahtani, M. (2016). Spontaneous preterm birth and single nucleotide gene polymorphisms: a recent update. *BMC Genomics*, 17(S9), 759. <https://doi.org/10.1186/s12864-016-3089-0>

Shinki, T., Ueno, Y., DeLuca, H. F., & Suda, T. (1999). Calcitonin is a major regulator for the expression of renal 25-hydroxyvitamin D 3-1 α -hydroxylase gene in normocalcemic rats. *Proceedings of the National Academy of Sciences*, 96(14), 8253–8258. <https://doi.org/10.1073/pnas.96.14.8253>

Siddesh, A., Parveen, F., Misra, M. K., Phadke, S. R., & Agrawal, S. (2014). Platelet-specific collagen receptor glycoprotein VI gene variants affect recurrent pregnancy loss. *Fertility and Sterility*, 102(4), 1078-1084.e3. <https://doi.org/10.1016/j.fertnstert.2014.07.002>

Silvano, L., Miras, M., Pérez, A., Picotto, G., de Barboza, G. D., Muñoz, L., Martín, S., Sobrero, G., Armelini, P., Mericq, V., & de Talamoni, N. T. (2011). Comparative analysis of clinical, biochemical and genetic aspects associated with bone mineral density in small for gestational age children. *Journal of Pediatric Endocrinology and Metabolism*, 24(7–8). <https://doi.org/10.1515/jpem.2011.196>

Simpson, R. U., Thomas, G. A., & Arnold, A. J. (1985). Identification of 1,25-

- dihydroxyvitamin D3 receptors and activities in muscle. *The Journal of Biological Chemistry*, 260(15), 8882–8891. <http://www.ncbi.nlm.nih.gov/pubmed/2991224>
- Slatopolsky, E., Gonzalez, E., & Martin, K. (2003). Proceedings: Pathogenesis and Treatment of Renal Osteodystrophy. *Blood Purification*, 21(4–5), 318–326. <https://doi.org/10.1159/000072552>
- Sonnenschein-van der Voort, A. M. M., Arends, L. R., de Jongste, J. C., Annesi-Maesano, I., Arshad, S. H., Barros, H., Basterrechea, M., Bisgaard, H., Chatzi, L., Corpeleijn, E., Correia, S., Craig, L. C., Devereux, G., Dogaru, C., Dostal, M., Duchon, K., Eggesbø, M., van der Ent, C. K., Fantini, M. P., ... Duijts, L. (2014). Preterm birth, infant weight gain, and childhood asthma risk: A meta-analysis of 147,000 European children. *Journal of Allergy and Clinical Immunology*, 133(5), 1317–1329. <https://doi.org/10.1016/j.jaci.2013.12.1082>
- Sorensen, I. M., Joner, G., & Jenum, P. A. (2012). Maternal serum levels of 25-hydroxyvitamin D during pregnancy and risk of type 1 diabetes in the offspring. *Diabetes*, 61. <https://doi.org/10.2337/db11-0875>
- Spilianakis, C. G., Lee, G. R., & Flavell, R. A. (2005). Twisting the Th1/Th2 immune response via the retinoid X receptor: lessons from a genetic approach. *European Journal of Immunology*, 35(12), 3400–3404. <https://doi.org/10.1002/eji.200535588>
- Stene, L. C., Ulriksen, J., Magnus, P., & Joner, G. (2000). Use of cod liver oil during pregnancy associated with lower risk of Type I diabetes in the offspring. *Diabetologia*, 43(9), 1093–1098. <https://doi.org/10.1007/s001250051499>
- Suda, T., Ueno, Y., Fujii, K., & Shinki, T. (2003). Vitamin D and bone. *Journal of Cellular Biochemistry*, 88(2), 259–266. <https://doi.org/10.1002/jcb.10331>

- Suksawatamnuay, S., Sriphoosanaphan, S., Aumpansub, P., Aniwana, S., Thanapirom, K., Tanasanvimon, S., Thaimai, P., Wiangngoen, S., Ponauthai, Y., Sumdin, S., Angspatt, P., Rerknimitr, R., Poovorawan, Y., & Komolmit, P. (2020). Association between Vitamin D Receptor Single-Nucleotide Polymorphisms and Colorectal Cancer in the Thai Population: A Case-Control Study. *BioMed Research International*, 2020, 7562958. <https://doi.org/10.1155/2020/7562958>
- Swamy, G. K., Garrett, M. E., Miranda, M. L., & Ashley-Koch, A. E. (2011). Maternal vitamin D receptor genetic variation contributes to infant birthweight among black mothers. *American Journal of Medical Genetics Part A*, 155(6), 1264–1271. <https://doi.org/10.1002/ajmg.a.33583>
- Swamy, G. K., Østbye, T., & Skjærven, R. (2008). Association of preterm birth with long-term survival, reproduction, and next-generation preterm birth. *Jama*, 299(12), 1429–1436. <https://doi.org/10.1001/jama.299.12.1429>
- Szpecht, D., Gadzinowski, J., Seremak-Mrozikiewicz, A., Kurzawińska, G., & Szymankiewicz, M. (2017). Role of endothelial nitric oxide synthase and endothelin-1 polymorphism genes with the pathogenesis of intraventricular hemorrhage in preterm infants. *Scientific Reports*, 7, 42541. <https://doi.org/10.1038/srep42541>
- Takeyama, K., Kitanaka, S., Sato, T., Kobori, M., Yanagisawa, J., & Kato, S. (1997). 25-Hydroxyvitamin D3 1alpha-hydroxylase and vitamin D synthesis. *Science (New York, N.Y.)*, 277(5333), 1827–1830. <https://doi.org/10.1126/science.277.5333.1827>
- Tamura, T., Goldenberg, R. L., Freeberg, L. E., Cliver, S. P., Cutter, G. R., & Hoffman, H. J. (1992). Maternal serum folate and zinc concentrations and their relationships to pregnancy outcome. *The American Journal of Clinical Nutrition*, 56(2), 365–370. <https://doi.org/10.1093/ajcn/56.2.365>

- Tanaka, Y., & DeLuca, H. F. (1984). Rat renal 25-hydroxyvitamin D₃ 1- and 24-hydroxylases: their in vivo regulation. *American Journal of Physiology-Endocrinology and Metabolism*, 246(2), E168–E173. <https://doi.org/10.1152/ajpendo.1984.246.2.E168>
- Taymans, S. E., Pack, S., Pak, E., Orban, Z., Barsony, J., Zhuang, Z., & Stratakis, C. A. (1999). The human vitamin D receptor gene (VDR) is localized to region 12cen- q12 by fluorescent in situ hybridization and radiation hybrid mapping: Genetic and physical VDR map. *Journal of Bone and Mineral Research*, 14(7), 1163–1166. <https://doi.org/10.1359/jbmr.1999.14.7.1163>
- Thomas, M. K., Lloyd-Jones, D. M., Thadhani, R. I., Shaw, A. C., Deraska, D. J., Kitch, B. T., Vamvakas, E. C., Dick, I. M., Prince, R. L., & Finkelstein, J. S. (1998). Hypovitaminosis D in Medical Inpatients. *New England Journal of Medicine*, 338(12), 777–783. <https://doi.org/10.1056/NEJM199803193381201>
- Thota, C., Farmer, T., Garfield, R. E., Menon, R., & Al-Hendy, A. (2013). Vitamin D elicits anti-inflammatory response, inhibits contractile-associated proteins, and modulates Toll-like receptors in human myometrial cells. *Reproductive Sciences (Thousand Oaks, Calif.)*, 20(4), 463–475. <https://doi.org/10.1177/1933719112459225>
- Tielsch, J. M. (2015). Global Incidence of Preterm Birth. *Nestle Nutrition Institute Workshop Series*, 81, 9–15. <https://doi.org/10.1159/000365798>
- Tizaoui, K., Kaabachi, W., Hamzaoui, A., & Hamzaoui, K. (2014). Contribution of VDR polymorphisms to type 1 diabetes susceptibility: Systematic review of case-control studies and meta-analysis. *The Journal of Steroid Biochemistry and Molecular Biology*, 143, 240–249. <https://doi.org/10.1016/j.jsbmb.2014.03.011>
- Tomedi, L. E., Simhan, H. N., & Bodnar, L. M. (2013). Early-pregnancy maternal vitamin D status and maternal hyperglycaemia. *Diabetic Medicine*, 30(9), 1033–1039.

<https://doi.org/10.1111/dme.12229>

Torkildsen, O., Grytten, N., & Aarseth, J. (2012). Month of birth as a risk factor for multiple sclerosis: An update. *Acta Neurologica Scandinavica. Supplementum*, 195.

<https://doi.org/10.1111/ane.12040>

Uitterlinden, A. G., Fang, Y., van Meurs, J. B. J., Pols, H. A. P., & van Leeuwen, J. P. T. M. (2004). Genetics and biology of vitamin D receptor polymorphisms. *Gene*, 338(2), 143–

156. <https://doi.org/10.1016/j.gene.2004.05.014>

Underwood, J. L., & DeLuca, H. F. (1984). Vitamin D is not directly necessary for bone growth and mineralization. *American Journal of Physiology-Endocrinology and Metabolism*,

246(6), E493–E498. <https://doi.org/10.1152/ajpendo.1984.246.6.E493>

Uriu-Adams, J. Y., Obican, S. G., & Keen, C. L. (2013). Vitamin D and maternal and child health: Overview and implications for dietary requirements. *Birth Defects Research Part C: Embryo Today: Reviews*,

99(1), 24–44. <https://doi.org/10.1002/bdrc.21031>

Ustun, N., Eyerci, N., Karadag, N., Yesilyurt, A., Zenciroglu, A., & Okumus, N. (2020). Association of vitamin D receptor gene FokI and TaqI polymorphisms and risk of RDS.

Journal of Maternal-Fetal and Neonatal Medicine, 33(21), 3640–3646.

<https://doi.org/10.1080/14767058.2019.1582629>

Valdivielso, J. M., & Fernandez, E. (2006). Vitamin D receptor polymorphisms and diseases. *Clinica Chimica Acta; International Journal of Clinical Chemistry*, 371(1–2), 1–12.

<https://doi.org/10.1016/j.cca.2006.02.016>

Vieth, R. (1999). Vitamin D supplementation, 25-hydroxyvitamin D concentrations, and safety. *The American Journal of Clinical Nutrition*, 69(5), 842–856.

<https://doi.org/10.1093/ajcn/69.5.842>

- Visser, M., Deeg, D. J. H., & Lips, P. (2003). Low Vitamin D and High Parathyroid Hormone Levels as Determinants of Loss of Muscle Strength and Muscle Mass (Sarcopenia): The Longitudinal Aging Study Amsterdam. *Journal of Clinical Endocrinology and Metabolism*, 88(12), 5766–5772. <https://doi.org/10.1210/jc.2003-030604>
- Vuolo, L., Di Somma, C., Faggiano, A., & Colao, A. (2012). Vitamin D and Cancer. *Frontiers in Endocrinology*, 3. <https://doi.org/10.3389/fendo.2012.00058>
- Walsh, J. M., Kilbane, M., McGowan, C. A., McKenna, M. J., & McAuliffe, F. M. (2013). Pregnancy in dark winters: implications for fetal bone growth? *Fertility and Sterility*, 99(1), 206–211. <https://doi.org/10.1016/j.fertnstert.2012.09.010>
- Wang, S., Xin, X., Luo, W., Mo, M., Si, S., Shao, B., Shen, Y., Cheng, H., & Yu, Y. (2021). Association of vitamin D and gene variants in the vitamin D metabolic pathway with preterm birth. *Nutrition*, 89, 111349. <https://doi.org/10.1016/j.nut.2021.111349>
- Webb, A. R. (2006). Who, what, where and when—influences on cutaneous vitamin D synthesis. *Progress in Biophysics and Molecular Biology*, 92(1), 17–25. <https://doi.org/10.1016/j.pbiomolbio.2006.02.004>
- WEBB, A. R., DECOSTA, B. R., & HOLICK, M. F. (1989). Sunlight Regulates the Cutaneous Production of Vitamin D₃ by Causing Its Photodegradation*. *The Journal of Clinical Endocrinology & Metabolism*, 68(5), 882–887. <https://doi.org/10.1210/jcem-68-5-882>
- Weiler, H., Fitzpatrick-Wong, S., Veitch, R., Kovacs, H., Schellenberg, J., McCloy, U., & Chui, K. Y. (2005). Vitamin D deficiency and whole-body and femur bone mass relative to weight in healthy newborns. *CMAJ. Canadian Medical Association Journal*, 172(6), 757–761. <https://doi.org/10.1503/cmaj.1040508>
- Weisse, K., Winkler, S., & Hirche, F. (2013). Maternal and newborn vitamin D status and its

- impact on food allergy development in the German LINA cohort study. *Allergy*, 68.
<https://doi.org/10.1111/all.12081>
- Wen, J., Hong, Q., Zhu, L., Xu, P., Fu, Z., Cui, X., You, L., Wang, X., Wu, T., Ding, H., Dai, Y., Ji, C., & Guo, X. (2017). Association of maternal serum 25-hydroxyvitamin D concentrations in second and third trimester with risk of gestational diabetes and other pregnancy outcomes. *International Journal of Obesity (2005)*, 41(4), 489–496.
<https://doi.org/10.1038/ijo.2016.227>
- Wen, S. W., Smith, G., Yang, Q., & Walker, M. (2004). Epidemiology of preterm birth and neonatal outcome. *Seminars in Fetal and Neonatal Medicine*, 9(6), 429–435.
<https://doi.org/10.1016/j.siny.2004.04.002>
- Whitfield, G. K., Hsieh, J. C., Nakajima, S., MacDonald, P. N., Thompson, P. D., Jurutka, P. W., Haussler, C. A., & Haussler, M. R. (1995). A highly conserved region in the hormone-binding domain of the human vitamin D receptor contains residues vital for heterodimerization with retinoid X receptor and for transcriptional activation. *Molecular Endocrinology (Baltimore, Md.)*, 9(9), 1166–1179.
<https://doi.org/10.1210/mend.9.9.7491109>
- WHO. (1970). The prevention of perinatal mortality and morbidity. Report of a WHO Expert Committee. *World Health Organization Technical Report Series*, 457, 1–60.
- Wolden-Kirk, H., Rondas, D., Bugliani, M., Korf, H., Van Lommel, L., Brusgaard, K., Christesen, H. T., Schuit, F., Proost, P., Masini, M., Marchetti, P., Eizirik, D. L., Overbergh, L., & Mathieu, C. (2014). Discovery of molecular pathways mediating 1,25-dihydroxyvitamin D3 protection against cytokine-induced inflammation and damage of human and male mouse islets of Langerhans. *Endocrinology*, 155(3), 736–747.
<https://doi.org/10.1210/en.2013-1409>

- Wu-Wong, J. R., Nakane, M., & Ma, J. (2007). Vitamin D analogs modulate the expression of plasminogen activator inhibitor-1, thrombospondin-1 and thrombomodulin in human aortic smooth muscle cells. *Journal of Vascular Research*, 44(1), 11–18. <https://doi.org/10.1159/000097812>
- Yadav, D. K., Chaudhary, U., & Shrestha, N. (2011). Risk Factors Associated with Low Birth Weight. *Journal of Nepal Health Research Council*, 9(2), 159–164. <https://doi.org/10.33314/JNHRC.V0I0.266>
- Yamamoto, M., Kawanobe, Y., Takahashi, H., Shimazawa, E., Kimura, S., & Ogata, E. (1984). Vitamin D deficiency and renal calcium transport in the rat. *Journal of Clinical Investigation*, 74(2), 507–513. <https://doi.org/10.1172/JCI111448>
- Yasmin, S., Osrin, D., Paul, E., & Costello, A. (2001). *Neonatal mortality of low-birth-weight infants in Bangladesh*.
- York, T. P., Eaves, L. J., Neale, M. C., & Strauss, J. F. (2014). The contribution of genetic and environmental factors to the duration of pregnancy. *American Journal of Obstetrics and Gynecology*, 210(5), 398–405. <https://doi.org/10.1016/j.ajog.2013.10.001>
- Zella, J. B., & DeLuca, H. F. (2003). Vitamin D and autoimmune diabetes. *Journal of Cellular Biochemistry*, 88(2), 216–222. <https://doi.org/10.1002/jcb.10347>
- Zhang, R., & Naughton, D. P. (2010). Vitamin D in health and disease: Current perspectives. *Nutrition Journal*, 9(1), 65. <https://doi.org/10.1186/1475-2891-9-65>
- Zhu, H., Wang, X., Shi, H., Su, S., Harshfield, G. A., Gutin, B., Snieder, H., & Dong, Y. (2013). A Genome-Wide Methylation Study of Severe Vitamin D Deficiency in African American Adolescents. *The Journal of Pediatrics*, 162(5), 1004-1009.e1. <https://doi.org/10.1016/j.jpeds.2012.10.059>

A Review on "Association of Mothers' Single Nucleotide Polymorphism (BsmI) of Vitamin D Receptor Gene with Risk of Preterm Birth and Birth Weight"

ORIGINALITY REPORT

6%

SIMILARITY INDEX

3%

INTERNET SOURCES

4%

PUBLICATIONS

1%

STUDENT PAPERS

PRIMARY SOURCES

1	"Dietary reference values for vitamin D", EFSA Journal, 2016 Publication	1%
2	www.science.gov Internet Source	1%
3	bmcpregnancychildbirth.biomedcentral.com Internet Source	<1%
4	"Preface", Walter de Gruyter GmbH, 2010 Publication	<1%
5	Submitted to University of North Florida Student Paper	<1%
6	Recent Results in Cancer Research, 2003. Publication	<1%
7	www.mdpi.com Internet Source	<1%
8	www.supermarketsavvy.com Internet Source	<1%

9	www.omicsonline.org Internet Source	<1 %
10	Diptika Tiwari, Saswati Sanyal Choudhury, Anjuma Begum, Tina Nath, Sujoy Bose. "Maternal Vitamin D receptor gene variant genotype and its influence on preterm birth, low birth weight, and fetal death. A case-control study from Northeast India", Gene Reports, 2022 Publication	<1 %
11	hdl.handle.net Internet Source	<1 %
12	Submitted to Erasmus University of Rotterdam Student Paper	<1 %
13	Mary Leppert, Marilee C. Allen. "Risk Assessment and Neurodevelopmental Outcomes", Elsevier BV, 2012 Publication	<1 %
14	"Vitamin D and the Lung", Springer Science and Business Media LLC, 2012 Publication	<1 %
15	mdpi.com Internet Source	<1 %
16	Leppert, Mary, and Marilee C. Allen. "Risk Assessment and Neurodevelopmental	<1 %

Outcomes", Avery s Diseases of the Newborn, 2012.

Publication

17

www.businessinsider.in

Internet Source

<1 %

18

Submitted to Texas Woman's University

Student Paper

<1 %

19

wiki.alquds.edu

Internet Source

<1 %

20

www.frontiersin.org

Internet Source

<1 %

21

I González-Molero. "Vitamin D deficiency in Spain: a population-based cohort study", European Journal of Clinical Nutrition, 12/22/2010

Publication

<1 %

22

Patricia Chocano-Bedoya, Alayne G Ronnenberg. "Vitamin D and tuberculosis", Nutrition Reviews, 2009

Publication

<1 %

23

Sheryl Vondracek. "Clinical challenges in the management of osteoporosis", Clinical Interventions in Aging, 2008

Publication

<1 %

24

qnaguides.com

Internet Source

<1 %

25

Mary Norval, Anna Coussens, Robert Wilkinson, Liza Bornman, Robyn Lucas, Caradee Wright. "Vitamin D Status and Its Consequences for Health in South Africa", International Journal of Environmental Research and Public Health, 2016

Publication

<1 %

26

Anna K. Coussens. "Immunomodulatory Actions of Vitamin D Metabolites and their Potential Relevance to Human Lung Disease", Current Respiratory Medicine Reviews, 2011

Publication

<1 %

27

Patricia T. Alpert, Ulfat Shaikh. "The Effects of Vitamin D Deficiency and Insufficiency on the Endocrine and Paracrine Systems", Biological Research For Nursing, 2016

Publication

<1 %

28

Vladimira Mondockova, Veronika Kovacova, Nina Zemanova, Martina Babikova et al. "Vitamin D Receptor Gene Polymorphisms Affect Osteoporosis-Related Traits and Response to Antiresorptive Therapy", Genes, 2023

Publication

<1 %

Exclude bibliography On