

The Generating Factors of Antimicrobial Peptide's Production in Human

A project submitted

by

Jerin Mahbub

ID: 13146032

Session: Spring 2013

to

the Department of Pharmacy

in partial fulfillment of the requirements for the degree of
Bachelor of Pharmacy (Hons.)



Inspiring Excellence

Dhaka, Bangladesh

July 2017

Dedication

This work is dedicated to my parents and my sibling to whom I owe my achievements.

Certification statement

This is to certify that, this project titled ‘The generating factors of antimicrobial peptide’s production in human’ submitted for the partial fulfillment of the requirements for the degree of Bachelor of Pharmacy from the Department of Pharmacy, BRAC University constitutes my own work under the supervision of Sinayat Mahzabeen, Lecturer, Department of Pharmacy, BRAC University and that appropriate credit is given where I have used the language, ideas or writings of another.

Signed,

Countersigned by the Supervisor

Acknowledgement

I would like to begin my gratitude to Almighty Allah for the help in the completion of this research and preparation of this paper.

I would like to thank my supervisor Sinayat Mahzabeen, Lecturer, Department of Pharmacy, BRAC University for her continuous provision, guidance and patience throughout the project work. As a person, she has continuously inspired and motivated me with her skilled knowledge which made me more passionate about the project when it began. I am really indebted to our chairperson, Dr. Eva Rahman Kabir, Chairperson, Department of Pharmacy, BRAC University, for her support, encouragement and kind cooperation all through the project.

Jerin Mahbub

Abstract

Antimicrobial peptide (AMP) has been recognized as a noble antibiotic against wide range of bacteria now a days. Isolated AMPs from various sources are used to control a number of diseases in humans. To reinforce their expression, some factors can be used to work on specific parts of human body. In this study, we accumulate seven factors which assist production of cathelicidin and defensins family of human AMPs. Vitamin D3 and bile salt prompt up cathelicidin expression in various epithelial cell lines through vitamin D receptor and farnesoid X receptor as respectively. On the other side, beta defensins can be induced by epigallocatechin-3-gallate, isoleucine, sulphoraphane and platelet release growth factor in targeted areas. Lastly, phenylbutyrate works on both cathelicidin and beta defensin 1. Most of them work effectively in dose and time dependent manner. In some cases, the combination therapy of two or three of them have been turned out positively than their individual action which is also highlighted here. In spite of their usefulness, some obstacles are still lagging behind them from implementation which need further investigation. This study clearly indicates that all the elements successfully elevate antimicrobial peptide expression within the body and also improve innate immunity to fight against infections in both primary and critical stages.

Table of content

Certification statement	iii
Acknowledgement	iv
Abstract	v
List of Tables	vii
List of Figures	viii
Abbreviation	x
1. Introduction:	1
2. Classification of Antimicrobial Peptide:	2
3. Antimicrobial Peptides in Human:	4
3.1 Cathelicidin	4
3.2 Defensins:.....	5
4. Clinical Importance of AMPs:	7
4.1 Natural antibiotics:	7
4.2 Immunomodulator:.....	7
4.3 Anti-inflammatory agent:.....	8
4.4 Anti-infective agent:.....	8
4.5 Cancer biomarker:.....	9
5. Factors Generating Antimicrobial Peptide	10
5.1 Epigallocatechin-3-Gallate.....	12
5.2 Vitamin D3.....	15
5.3 Sodium Phenylbutyrate	17
5.4 Isoleucine	20
5.5 Sulphoraphane.....	22
5.6 Bile Salt	24
5.7 Platelet Release Growth Factor	26
6. Discussion:	28
7. References:	30

List of Tables

Table 3.1	Structure and sequences of human cathelicidin and defensins	6
Table 5.1	List of factors, targeted AMP, targeted site and related experiments. List of factors, targeted AMP, targeted site and related experiments.	10-11

List of Figures

Figure 2.1	Different types of antimicrobial peptides	3
Figure 5.1	Chemical structure of Epigallocatechin-3-Gallate	12
Figure 5.2	Showing result of pBD 1 & pBD 2 level after treating with EGCG	14
Figure 5.3	Chemical structure of Cholecalciferol	15
Figure 5.4	Showing cell stimulation with 4mM PBA and solvent alone	18
Figure 5.5	Showing result for expression of beta defensin 1 in VA10 cells and U937 cells	19
Figure 5.6	Chemical structure of isoleucine	20
Figure 5.7	Relative expression of epithelial beta defensin in treatment with both L & D- isoleucine	21
Figure 5.8	Proteasome inhibitors MG132 and Lactacystin block isoleucine activation of the defensin promoter	21
Figure 5.9	Dose dependent expression of sulphoraphane (1-20 μ M) beta-defensin 2 in colorectal cell lines	23
Figure 5.10	Chemical structure of chenodeoxycholic acid	24
Figure 5.11	Chemical structure of ursodeoxycholic acid	24
Figure 5.12	Showing possible effect of CDCA and UDCA along with vitamin D on cathelicidin's activity	25

Figure 5.13	Showing HBD-2 expression with PRGF in different and same dose	26
Figure 5.14	Showing the level of HBD-2 gene expression after treatment with Vivostat PRF®	27
Figure 5.15	Showing HBD-2 gene expression and protein level upon activation of EGFR with cetuximab	27

Abbreviation

AMP	-	Antimicrobial peptide
HBD	-	Human beta defensin
EGCG	-	Epigallocatechin-3-gallate
PBA	-	Phenylbutyrate
SFN	-	Sulphoraphane
HDAC	-	Histone Deactylase
VDR	-	Vitamin D Receptor
PRGF	-	Platelet Release Growth Factor

1. Introduction:

Antimicrobial peptides (AMPs) are the part of first line innate immune system that defend the host from various pathogenic attacks. Besides virus, fungi and cancerous cells, AMPs are highly effective against both gram positive and gram negative bacteria. They have been isolated not only from human but also from single celled organisms, fish, bird, insects, invertebrates and other mammals. Basically, their expression within the body increases from infectious stimuli. These AMPs or broad spectrum antibiotics were first discovered in 1939 by Dubos when he extracted an antimicrobial agent from a soil bacillus strain to use it for protection of mice from pneumococci infection (R.J, 1939). AMPs, this small peptides generally consist of 12 to 50 amino acids. Their net charges vary from -3 to +20. Variability in their structures is one of the benefits of them to use as an antimicrobial agent. Antibiotic resistance, the burning issue in the medical sector, can be minimized or controlled with the help of antimicrobial peptides. Moreover, they possess other functions such as, immunomodulator, wound healing and apoptosis etc. Recently, they have been used as a marker in order to detect many cancerous cells which also neutralized by their apoptosis activity.

From the very earlier, an extent research work has been done on AMPs activity. The structure, characteristics, classification and their mechanism of action have been identified. The effective activity concentration has been examined against bacteria, fungi, virus and other pathogens. These peptides play greater role in healing and preventing many fatal diseases. In spite of, they are still lacking behind for clinical implication in some areas due to low bioactivity over available drugs. However, some synthetic peptides which are prepared from these natural ones significantly available in treating diseases. The factors responsible for inducing their production in human body also have been identified. They are highly potent to induce antimicrobial peptides which can be taken either as food or nutrients. So, the proper use of them can lead to induce AMPs activity as well as innate immunity in order to keep us away from being infected by any diseases easily. The aim of this review is to accumulate these factors with their potentiality and also further improvement could be done in these area.

2. Classification of Antimicrobial Peptide:

The majority of AMPs are cationic in nature due to presence of arginine and/or lysine but anionic AMPs are minor in number. Till now four classes of AMPs have been proposed based on their structure and they are α -helical, β -sheet, loop and extended peptides (Hancock RE, 1998).

- The α -helical peptides family is most common and cationic in nature. This class contains 250 peptides which composed of less than 40 amino acids other than cysteine (KA., 2005).
- The β -sheet peptides are more stabilized because of the disulfide bridges (KA., 2005). These bridges and cyclic structures have greater impact on their action and also essential for antibacterial activity (Matsuzaki K, 1997) (AG., 1999).
- The extended peptides are linear in shape but have an unusual composition where over expression of one or more amino acids can be found. This class contains 90 peptides and very flexible in solution.

According to a study, four minimum possible reasons had been found for this structural diversity among AMPs (Hancock RE, 2000).

1. All classes of antimicrobial peptides cannot be active against every pathogen. So, diversity in structure can help them to kill specific microbes.
2. Different structures of AMPs can work together with synergism.
3. Those structures are helping them to do other non-antibacterial activities such as, chemotactic or pro-inflammatory activities.
4. Different AMPs are produced by different cell types which is another cause to be different in structure.

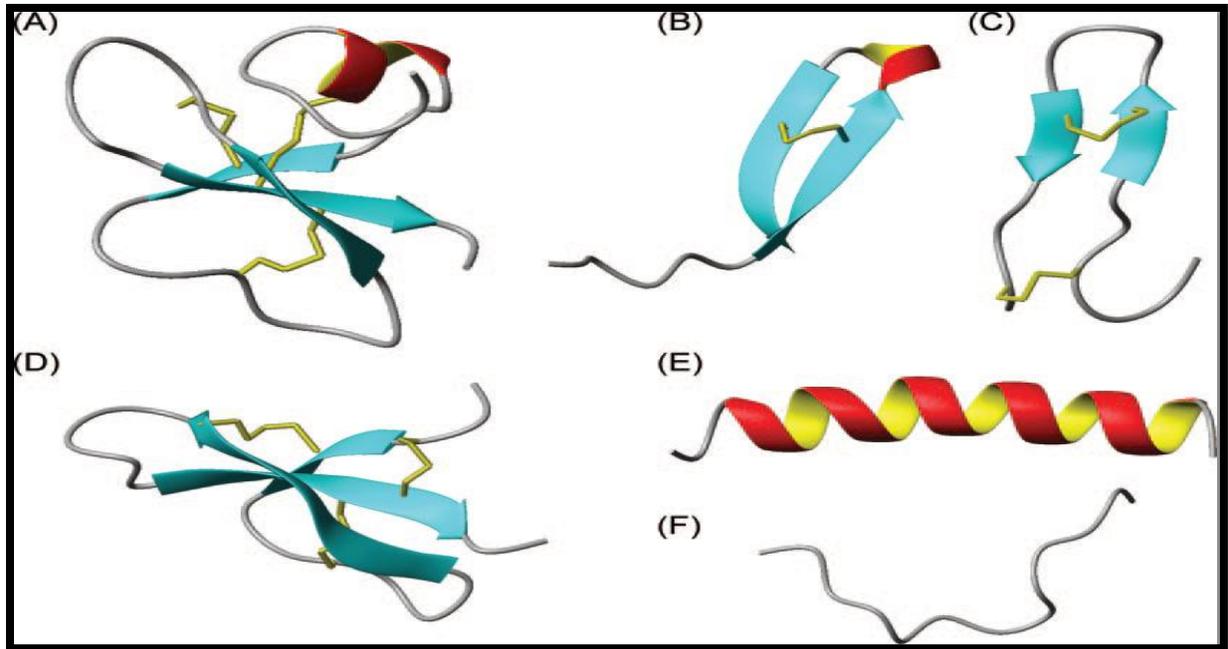


Figure 2.1: Different types of antimicrobial peptide. (Håvard J, 2006)

In the above picture, various classes of antimicrobial peptides have been shown (Håvard J, 2006). They are given below.

- A) Mixed structure of human β -defensin-2;
- (B) Looped thanatin;
- (C) β -sheeted polyphemusin;
- (D) Rabbit kidney defensin 1;
- (E) α -helical magainin-2;
- (F) Extended indolicidin.

3. Antimicrobial Peptides in Human:

Many antimicrobial peptides can be found in different species but among them a few can be isolated from the human. Among them, two are very much important and they are cathelicidins (LL-37) and defensins (alpha and beta).

3.1 Cathelicidin

Cathelicidins are varying in amino acid sequence, structure and size. They range in size from 12-80 amino acid residues and have a wide range of structure (Gennaro R, 2000). About 30 cathelicidin family members have been identified in mammalian species but LL-37 is the only one cathelicidin found in human. LL-37 which begins with two leucine residues at its N-terminus, and is 37 residues long, with a molecular weight of 18 kDa (Gudmundsson GH, 1995). LL-37 is disordered in aqueous solution but can quickly transfer into α -helix while binding with bacterial cell wall (Nijnik A, 2009).

Location: Cathelicidins are a family of polypeptides and mostly found in metropolis, epithelia of the skin, guts and lungs, monocytes, natural killer cells and mast cells.

Mechanism of action: Though the actual mechanism of action of cathelicidin is not clear, it is hypothesized that they bind to the cytoplasmic membranes of microbes and cause a disruptive effect (Lee CC, 2011).

Activity: It can work effectively as antimicrobial agent at a physiological concentration of approximately 2 μ g/ml but can increase eventually during infection (Bals R, 1999) (Schaller-Bals S, 2002). It is strongly active against various bacterial strains such as, Escherichia coli, Staphylococcus aureus, Pseudomonas aeruginosa, Neisseria gonorrhoeae etc. It prohibits the formation of P. aeruginosa bacterial biofilm which is a bacterial growth pattern related to the antibiotic resistance and it happens at 0.5 μ g/ml concentration only (Overhage J, 2008). This inhibition is done by lowering the attachment of bacterial cells, provoking quorum sensing system and stirring twitching motility which ultimately affecting the gene regulation for biofilm formation (Overhage J, 2008). This α -helical peptide also gives immune response against Mycobacterium tuberculosis. It has been observed that mycobacterial infection stimulates the production of this peptide in A549 epithelial cell, alveolar macrophages, neutrophils and Toll-like receptors (Liu PT,

2006) (Rivas-Santiago B, 2008). Moreover, it had been found that LL-37 gene expression is influenced by MEK 1/2 and p38 MAPK signaling pathways in A549 cells infected with *Mycobacterium bovis* bacillus Calmette-Guerin which is a tuberculosis vaccine mostly used worldwide (Mendez-Samperio P, 2008). As an antiviral agent, LL-37 has been found to give local protection against HIV-1 infection by epithelial expression (Bergman P, 2007). It was found that the immunological response of this peptide has been changed in infected patients with papillomavirus (Conner K, 2002). As an antifungal agent, it is also active against *Candida albicans*. The activity is depending on the culture condition whether it will effective as a superficial barrier to the invasion (den Hertog AL, 2006) (Lopez-Garcia B, 2005).

3.2 Defensins:

Defensins are also small, cysteine-rich and cationic peptide which are consisting of 18-45 amino acids with 6-8 cysteine residues and they form disulfide bridges. Due to the differences in alignment of disulfide bond and molecular structure, two subclasses can be found in human. They are alpha defensins and beta defensins. Human alpha defensins are also called human neutrophil peptides and have four subtypes such as, HNP1, HNP2, HNP3 and HNP4 which are 29-35 amino acids long. First three subtypes increase the expression of interleukin-1 and tumor necrosis factor α in human monocytes (Chaly YV, 2000). Another two α -defensins (HD-5 & 6) are known as enteric defensins. The C terminal part in their structure is responsible for antimicrobial activity (Contreras, 2005). They can take part in the oxygen independent killing of phagocytosed microbes in neutrophil. Furthermore, human beta defensins (hBD) can also be classified into four and they are hBD-1, hBD-2, hBD-3 and hBD-4, expressed by different intestinal epithelial cells (K.D. Smet, 2005).

Location: HNP1, HNP2, HNP3 and HNP4 are found in azurophilic granules of neutrophils as mature peptides (Nizet, 2003). Again, HD-5 & 6 are found in the granules of peneth cells of small intestine and epithelial cells of female urogenital tract (Jones DE, 1992). On the other hand, HBD-1, 2, 3 & 4 can be found in several epithelial cells of respiratory tract, gastrointestinal tract, cornea, skin and kidney etc.

Mechanism of Action: Defensins electrostatically bind to membranes, causing the formation of multimeric pores and the leakage of essential minerals and metabolites (Lehrer R I, 1989). They also cause membrane depolarization, decreased cytoplasmic ATP levels, and inhibited cellular respiration (Cociancich S, 1993). The entrance of defensins into cells has caused DNA damage (Gera J F, 1991) (Lehrer R I, 1985).

Activity: HBD-1 and 2 are more bactericidal against gram negative bacteria rather than gram positive. On the other hand, HBD-3 kills broad ranges of microbes and opportunistic pathogenic yeast like *Candida albicans*. Other than antimicrobial activity, defensins also show their potentiality in stimulation of cell proliferation, chemo attraction of immune cells, induction of cytokine and cell proliferation (Contreras, 2005).

Table 3.1: Structure and sequences of human cathelicidin and defensins

Peptide	Structure	Amino acid sequence
LL-37	α -helix	LLGDFFRKSKEKIGKEFKRIVQRIKDFLRNLPRTES
HNP-1	β -sheet	ACYCRIPACIAGERRYGTCTIYQGRLWAFCC
HNP-4	Same	VCSCRLVFCRTELRVGNCLIGGVSFTYCCTRVD
HD-5	Same	ARATCYCRTGRCATRESLSGVCEISGRLYRLCCR
HD-6	Same	RAFTCHCRRS-CYSTEYSYGTCTVMGN-HRFCCL
HBD-1	Same	DHYNCVSSGGQCLYSACPIFTKIQTGTCYRGKAKCKK
HBD-2	Same	DPVTCLKSGAICHPVFCPRRYKQIGTCGLPGTKCCKKP
HBD-3	Same	QKYYCRVRGGRC AVL SCLPKEEQIGKCSTRGRKCCRKK
HBD-4	Same	LDRICGYGTARCRKK-CRSQEYRIGRCPNTYA CCLRKPWDESLLNRTK

Reference: Kris De Smet & Roland Contreras (2005) Human antimicrobial peptides: defensins, cathelicidins and histatins. J. Springer. Biotechnology Letters 27: 1337-1347

4. Clinical Importance of AMPs:

Antimicrobial peptides have a greater clinical importance in medical sector.

4.1 Natural antibiotics:

Antimicrobial peptides is such natural antibiotic which have greater importance in killing multidrug-resistant microorganisms that bring it into high spot in treating diseases. Now a days, antibiotic resistance has become a major issue in the field of treatment. As we know, antibiotics do not only useful in killing bacteria but also in surgery, chemotherapy, chronic infections and patients suffering from diabetes, end-stage renal disease, rheumatoid arthritis and organ transplant etc (J., 2016) (The lancet infectious diseases comission, 2013). From recent review on antimicrobial resistance, it has been known that about 700,000 annual deaths are currently attributable to drug-resistant pathogenic infections (Gould IM, 2013). The overuse of antibiotics, environmental factors and also uncontrolled sales in many low or middle income countries are the possible reasons behind antibiotic resistance development (Radek K, 2007). Penicillin, tetracycline, gentamycin, vancomycin and levofloxacin etc. antibiotic have been already resistant to the human body in past few decades. People are not using medicines schedule wise and not completing course as well which is starting a base line for resistance and making easier for microbes to grow again with more virulence. Besides human body, antibiotics are polluting environment because of their introduction into human wastes, animals and the pharmaceutical industry (Martinez, 2012). To handle this critical situation we can use natural antibiotics which can kill microbes with same efficiency and does not produce resistant so quickly because of their various mode of action against pathogens.

4.2 Immunomodulator:

These peptides have ability to boost immunity by behaving as immunomodulator. Human cathelicidin, LL-37 is a chemoattractant for mast cells, monocytes, T lymphocytes and neutrophils (Yang D, 2000). LL37 contributes to host defense against microbial invasion, by participating in the recruitment of leukocytes to sites of infection. This mechanism is potentially important in vivo, because the chemotactic activity of LL37, unlike its antimicrobial action, is not significantly inhibited by the presence of human

serum. The activation of FPRL-1 requires relatively high concentrations of LL-37 (10–5M) when compared to other classical chemoattractant agents, suggesting a low-affinity peptide-receptor interaction (De Y, 2000). LL-37 modulates cellular immune responses by stimulating chemokine production. It activates airway epithelial cells through the activation of mitogen activated protein kinase (MAPK) and increases the release of potent chemoattractant IL-8 (De Y, 2000). In keratinocytes, LL-37 induces IL8, and promotes migration and wound healing, and these activities depend on ADAM family metalloproteinase, EGFR and FPRL1 (Carretero M, 2008).

4.3 Anti-inflammatory agent:

They are widely used in preventing skin diseases. Studies have shown that LL-37 and HBD-2 are greatly increased in patients with inflammatory skin conditions (Stolzenberg ED, 1997) (Gallo RL, 1940) (Froh M, 1997). LL-37 is induced in human keratinocytes during psoriasis, lupus erythematosus and contact dermatitis (FROHM M, 1997). It is also able to induce angiogenesis, a process important for wound healing and tissue repair (KOCZULLA R, 2003).

4.4 Anti-infective agent:

They are used as anti-infective agents in the medical sector (Hancock, R. E.W., 2006). Though the major importance of antimicrobial peptides are as antibiotics but they also effectively act as antiviral and antifungal. Now a days, various manipulations with chemical structure have been done to create designer synthetic peptides for the development of AMPs as a new class of drugs to prevent and treat systemic and topical infection.

Antimicrobial peptides can be used as intestinal infections. Paneth cells, which are a characteristic epithelial lineage of the small intestine and localize to the bottom of the intestinal crypts, secrete α -defensins in response to bacterial antigens including lipopolysaccharide and muramyl dipeptide (Ayabe T, 2000). HD-5 plays multiple roles in inflammation and infection. It is found in metaplastic Paneth cells in the colon of IBD patients and presumably it serves as a protective role in response to bacterial

challenges during colitis (Cunliffe RN, 2001). The cationic proteins extracted from colonic mucosal biopsies of PPAR γ deficient mice showed a lack of bactericidal activity against *Candida albicans*, *Bacteroides fragilis*, *Enterococcus faecalis* and *E. coli*, suggesting that HBD-1 may play a role in colonic inflammation and infection (Peyrin-Biroulet L, 2010).

4.5 Cancer biomarker:

AMPs can be used as anticancer agent against various types of cancer. A recent report showed that LL-37 activated p53 dependent but caspase independent activation of apoptosis in human cancer cells including HCT116 and LoVo. In addition, LL-37 expression in human colonic tumors is down-regulated while LL-37 is expressed strongly in normal colonic tissues (Ren SX, 2012). On the other hand, alpha defensin may serve as a biomarker for colorectal cancer which is based on a study with 100 colorectal cancer patient (Kemik O, 2011).

5. Factors Generating Antimicrobial Peptide:

As AMPs are part of our body defense, the peptides are regulated by numerous factors which can be controlled. All the listed factors work on either Cathelicidin or Defensins. Following is a table belongs to the list the factors and some basic information to express specific antimicrobial peptides in brief.

Table 5.1: List of factors, targeted AMP, targeted site and related experiments.

Factors	Targeted AMP	Targeted site in Human	References
Vitamin D3	Cathelicidin	Keratinocytes of human skin	Svensson D, Nebel D, Voss U, Ekblad E & Nilsson B. Vitamin D-induced up-regulation of human keratinocyte cathelicidin antimicrobial peptide expression involves retinoid X receptor <i>α</i> . 2016.
Phenylbutyrate	Cathelicidin and beta defensin 1	Human bronchial epithelial cell line, renal carcinoma cell line, colonic adenocarcinoma cell line and leukemic monocyte lymphoma cell line	Steinmann J, Halldorsson S, Agerberth B, and Gudmundsson G H. Phenylbutyrate Induces Antimicrobial Peptide Expression. Institute of Biology, University of Iceland, Reykjavik, Iceland, and Medical Biophysics and Biochemistry, Karolinska Institutet, Stockholm, Sweden. 2009. Vol. 53, No. 12.
Bile Salt	Cathelicidin	Human liver	D'ALDEBERT E, BIYEYEME M J, MERGEY M, WENDUM D, FIRRINCIELI D, COILLY A, FOUASSIER L, CORPECHOT C, POUPON R, HOUSSET C and CHIGNARD N. Bile Salts Control the Antimicrobial Peptide Cathelicidin Through Nuclear Receptors in the Human Biliary

			Epithelium. <i>Gastroenterology</i> 2009; 136:1435–1443.
Epigallocatechin-3-Gallate	Beta defensin	Porcine intestinal epithelial cell	Murphy L. Y. Wan, K. H. Ling, M. F. Wang and Hani El-Nezami., Green tea polyphenol epigallocatechin-3-gallate improves the epithelial barrier function by inducing the production of antimicrobial peptide pBD-1 and pBD-2 in monolayers of porcine intestinal epithelial IPEC-J2 cells. 2016. <i>Mol. Nutr. Food Res.</i> 60, 1048–1058.
Isoleucine	Beta defensin	Bovine kidney epithelial cell	Pascale Fehlbaum, Meena Rao, Michael Zasloff, and G. Mark Anderson. An essential amino acid induces epithelial beta-defensin expression. Magainin Research Institute and Magainin Pharmaceuticals Inc. 2000.
Sulphoraphane	Beta defensin 2	Human colorectal cancer cell lines	Schwab M, Reynders V, Loitsch S, Steinhilber D, Schroder O and Stein J. The dietary histone deacetylase inhibitor sulforaphane induces human β -defensin-2 in intestinal epithelial cells. 2008. <i>Immunology</i> , 125, 241–251.
Platelet Release Growth Factor	Beta defensin 2	Primary keratinocyte	Andreas Bayer, Justus Lammel, Franziska Rademacher, Justus Groß, Markus Siggelkow, Sebastian Lippross, Tim Kluter, Deike Varoga, Mersedeh Tohidnezhad, Thomas Pufe, Jochen Cremer, Regine Glaser, and Jurgen Harder. Platelet-released growth factors induce the antimicrobial peptide human beta-defensin-2 in primary keratinocytes. <i>Experimental Dermatology</i> , 2016, 25, 460–465.

5.1 Epigallocatechin-3-Gallate

Tea is one of the popular beverages consumed by almost everyone. Green, black and Oolong tea are the most common form all around the world. Among all of them, the consumption of green tea has significant effects on human health (Cabrera C, 2006). Generally green tea composed of protein, amino acid, lipids, minerals, carbohydrates and many others compounds. Green tea also contains polyphenols which may present up to 30% of dry weight. It is evident that polyphenols protect against a wide range of oxidative stress related disorders, such as cardiovascular diseases, cancers, inflammation, neurodegenerative diseases, and diabetes (Scalbert, 2005). Most of the green tea polyphenols are flavonols, commonly called catechins. There are four types of catechins mainly find in green tea among them one of is epigallocatechin-3-gallate (EGCG) (Sano M, 2001). A cup of green tea may contain 100-200 mg of EGCG (Zaveri, 2006).

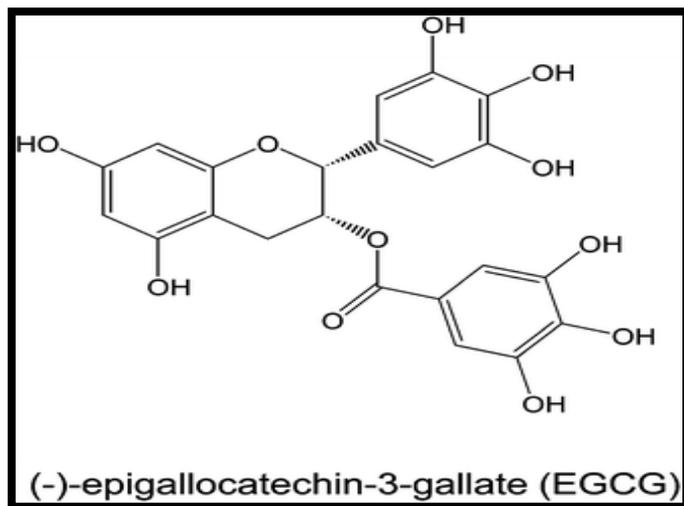


Figure 5.1: Epigallocatechin-3-gallate (EGCG)

Epidemiological surveys show that regular green tea consumption is related to lower chance of cardiovascular diseases, cancers, and obesity (Bell, 2002) (McKay, 2002). It is assumed that the ability of EGCG to bind many biological molecules and provoke the activity of various enzymes and signal transduction pathways at the micromolar and nanomolar level is the reason behind this beneficial effect of green tea (Lee MJ, 2002). As EGCG is a food-derived component, intestinal

epithelial cells are continuously and directly exposed to EGCG. This is the reason it can exert beneficial effects to the intestine. A previous study has been done demonstrating that the antimicrobial activity of AMPs, such as beta-defensins, may be associated with bacterial populations in the intestine, which naturally present a barrier limiting undesirable gut mucosal infections and preventing against bacterial translocation (Veldhuizen, 2008). Another study has also shown that EGCG induced defensin secretion on gingival epithelial cell (Lombardo Bedran, 2014).

A study was conducted based on the hypothesis that EGCG has protective response on intestinal barrier functions against bacterial translocation. The experiment was done by using porcine jejunal epithelial cells, IPEC-J2. IPEC-J2 cells isolated from neonatal piglet mid-jejunum. The cell line is unique in that it is derived from small intestinal tissue. IPEC-J2 cell line was chosen for the current study because it is a better model of normal IEC line than transformed cell line such as Caco-2 to study pathogen–host interactions (Geens, 2011). It conserves its epithelial nature, and is well documented to be able to express and secrete AMPs such as porcine beta-defensins 1 and 2 (pBD-1 and 2) as part of the innate immune response (Mariani, 2009).

EGCG had reduced bacterial translocation at a noncytotoxic concentration. It enhanced epithelial immunological barrier but not epithelial physical barrier. It may also be attributed to the secretions of defensins which was examined by ELISA with the amount of 12, 25 and 50 μ M of EGCG. Compare with control group, 25 μ M induced higher level of pBD-1 and 2 secretion significantly. In case of expression of pBD-1 and 2, EGCG successfully enhanced the expression of pBD-2 by 50% but no significant result for pBD-1.

This test was done by specific MAPK pharmacological inhibitor which shown that inhibition to p38 MAPK reduced the expression of pBD-2 and protein secretion. The induction of pBD-1 expression plays a potential role in surveillance and maintenance of a homeostatic state of gut micro biota on the mucosal epithelium (Veldhuizen E. H., 2006) (Elahi, 2006), whereas elevated pBD-2 expression will likely inhibit pathogenic bacteria (Zhang, 2011).

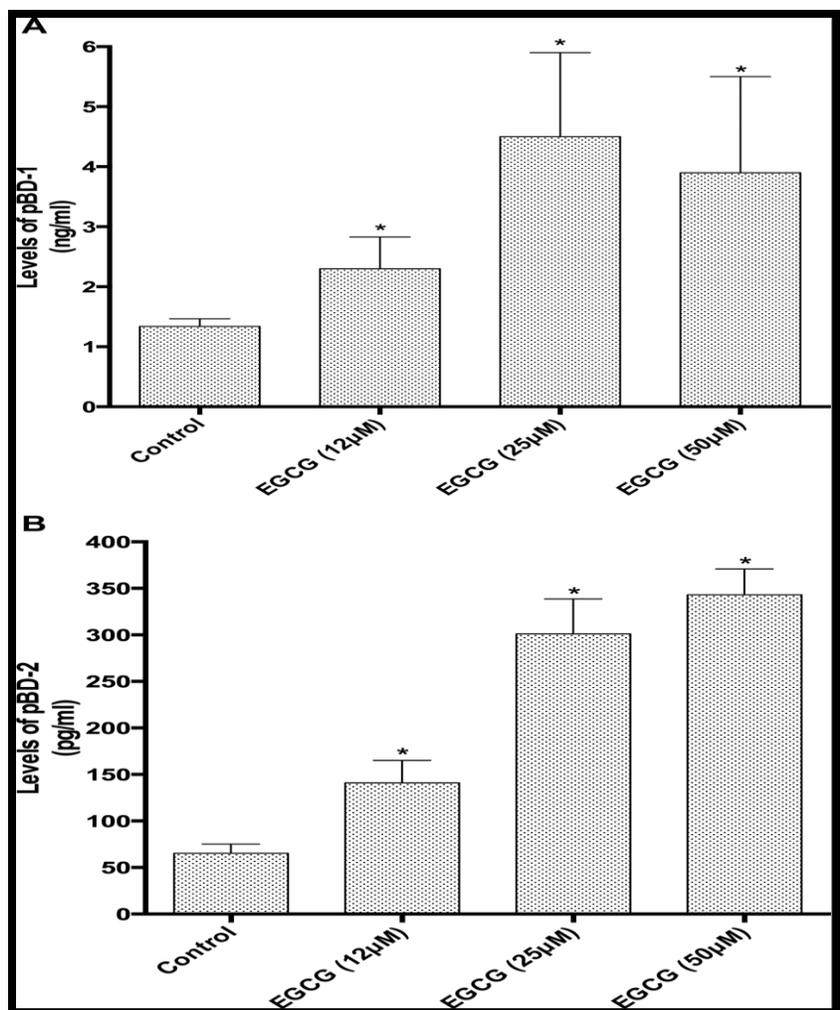


Figure 5.2: Showing result of pBD 1 & pBD 2 level after treating with EGCG (Murphy L. Y. Wan, 2016).

It was the first study to illustrate the protective capacity of EGCG on intestinal cells. The obtained results were satisfied enough to prove the fact that how much beneficial effect can be taken by adding green tea into our daily diet chart. Moreover, it helps to increase the production of mRNA and protein in both defensins. EGCG may be useful for prevention of intestinal disorder or bacterial infection in animals/humans (Murphy L. Y. Wan, 2016).

receptor signaling in human gingival and skin biopsies and to investigate the importance of RXRs for 1,25D3-induced CAMP gene activity in human keratinocyte HaCaT cells. The HaCaT cell line is a spontaneously immortalized human keratinocyte cell line from adult skin showing complete epidermal differentiation capacity, and thus these cells are most probably representative of human keratinocytes in vivo (Boukamp et al, 1988). Eight students were selected for the experiment and biopsies were collected from skin and gingiva by using a sterile biopsy punch. After going through the process, the obtained result was analyzed by various tests to ensure the effectivity of the study.

VDR immunoreactivity was tested from those individuals where positive cells were observed within the spectrum of both skin and gingiva. The mRNA expression for CAMP was about eight times higher ($P < 0.01$) in skin compared with gingiva. The expression of mRNA for VDR was identical in human skin and gingival biopsies confirming the VDR protein expression data. On the other hand, the RXR α expression was about three times higher ($P < 0.01$) in skin vs. gingiva, whereas the RXR β expression was similar in skin and gingival biopsies.

Another study was conducted at 2008 and based on administration of oral vitamin D induces cathelicidin production in atopic individuals that basically a skin problem. Atopic dermatitis is a type of inflammation which results in itchy, red, swollen and cracked skin. It affects 10% to 20% of children and 1% to 3% of adults (Schultz-Larsen FV, 2002). Individuals with atopic dermatitis are at an increased risk for cutaneous infections with *Staphylococcus aureus*, herpes simplex, and the small pox or vaccinia virus (Boguniewicz M, 2006). It has been shown that defects in the capacity to increase the production of cathelicidin, may account for this increase in infections (Ong PY, 2002) (Schauber J, 2008). The experiment was done on 14 normal controls and 14 atopic individuals suffering from moderate to severe atopic dermatitis. Firstly, their biopsies were taken and baselines of calcium and vitamin D were obtained. 4000 IU/day of oral vitamin D3 was given to the subjects for 21 days. After the finite time, biopsies were collected again along with serum calcium and vitamin D level. Those samples had gone under analysis with proper methods and techniques.

From the result of this experiment, AD lesional skin showed a significant increase in cathelicidin expression from a median of 3.53 relative copy units (RCU) before supplementation to a median of 23.91 RCU post supplementation and for nonlesional skin median value was from 1.50 RCU to 1.75 RCU. On the other side, normal skin showed a modest increase from a median of 1.0 RCU to

1.78 RCU. Immunofluorescence staining of duplicate skin biopsies from lesional skin confirmed results from RT-PCR and demonstrated an increase in cathelicidin protein after oral vitamin D supplementation (Tissa R. Hata, 2008). The intake of oral supplement also increased the level of 25-hydroxyvitamin D level (Vieth R, 2001). During this time, their serum 25-hydroxyvitamin D levels rose from a median of 24.5 mg/mL to 37 mg/mL. Similarly, atopic subjects' serum calcium levels also decreased from a median of 9.6 mg/dL to a median of 9.4 mg/dL post supplementation, and their serum 25-hydroxyvitamin D levels rose from a median of 22.5 mg/mL to 35.5 mg/mL. Analysis of previous data has shown that response to wound or disruption of the epidermal barrier, cathelicidin is induced (Schauber J D. R., 2007). But this data had shown that there is a small (but not statistically significant) increase in cathelicidin in AD lesional skin before treatment with vitamin D indicative of some ability of the atopic subject to induce cathelicidin with disruption of the epidermal barrier (Tissa R. Hata, 2008).

5.3 Sodium Phenylbutyrate

Sodium phenylbutyrate or 4-phenylbutyrate or phenylbutyrate (PBA) is a salt of an aromatic fatty acid and chemical formula is $C_{10}H_{11}NaO_2$. Basically it is converted into phenylacetate by β -oxidation (Spira AI, 2003). It is produced from dietary fibers by bacterial fermentation in the colon.

PBA is a histone deacetylase inhibitor which can be used as a mood stabilizer and anti-epileptic (Iannitti & Palmieri, 2011).

Phenylbutyrate has several clinical applications in which some of them are already applied and some are still under research. One of the important role of phenylbutyrate has been found that it can stimulate the production of LL-37 which is the only antimicrobial peptide of cathelicidin family in human body. To establish the fact, PBA was treated with various cell lines (Jonas Steinmann, 2009). It also has been said that vitamin D3 worked synergistically with PBA to boost up antimicrobial peptide expression at mRNA level. Phenylbutyrate induces cathelicidin in four cell lines and those are human bronchial epithelial cell line (VA10), renal carcinoma cell line (A498), colonic adenocarcinoma cell line (HT-29) and leukemic monocyte lymphoma cell line (U937). Same dose and time duration was given to all of the cells and increased level of cathelicidin was related to the dose and increased with time. It increased gradually with a period

of 48hours and PBA concentration was up to 4mM. At the opposite side, this gene expression by PBA can be hampered when treated with cyclohexidine and MAP kinase inhibitors because from previous studies it revealed that MAP kinase signaling pathway has effect on PBA enhanced CAMP expression (Schauber, 2003).

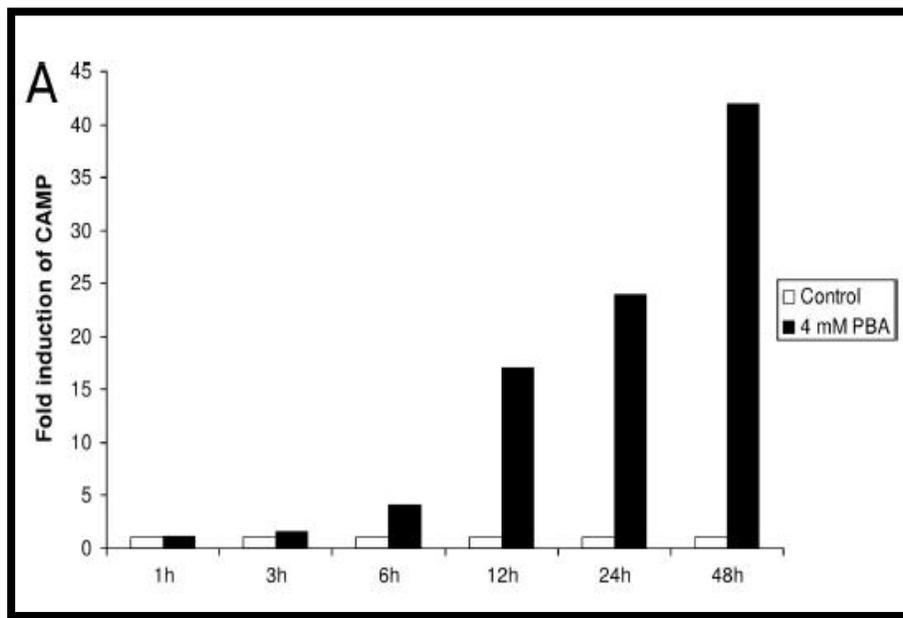


Figure 5.4: Showing cell stimulation with 4mM PBA and solvent alone (Jonas Steinmann, 2009).

The PBA concentration used in the experiment was approximately close to the reachable plasma level after oral administration. Also, it can be brought into medical field to boost up innate immunity as it has control over AMPs expression. Unfortunately, phenylbutyrate has limitation due to rapid degradation (Gilbert, 2001). To defeat the problem, some synthetic analogues of PBA has been used that shown half -lives longer stability than original in vivo (Pace, 2002). Moreover, vitamin D3 and phenylbutyrate works synergistically to improve CAMP gene expression. This can help tuberculosis patient to settle down vitamin D3 level by taking PBA as a medicine (Jonas Steinmann, 2009). From another study, an optimum dose has been found for PBA and vitamin D3 combination to boost up AMPs activity in tuberculosis which is 500 mg PB b.d. plus 5000 IU vitamin D3 o.d. taken orally (Akhirunnesa Mily, 2013). This combination can proceed intracellular killing of Mycobacterium Tuberculosis because of the induction of LL-37 expression. It also has been said that intracellular killing is mediated by autophagy and apoptosis.

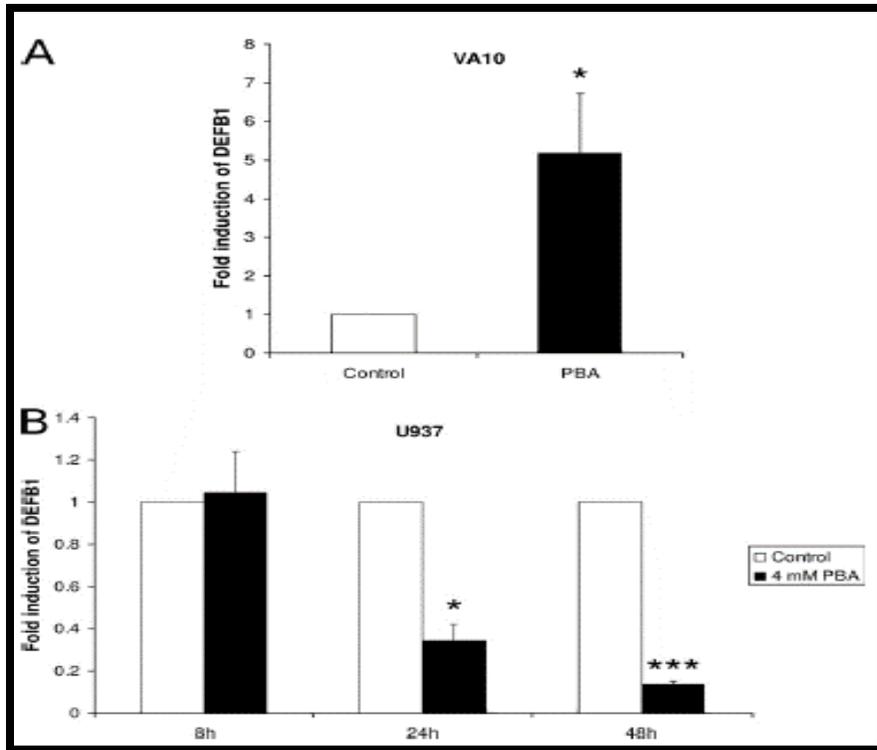


Figure 5.5: A) Showing result for expression of beta defensin 1 in VA10 cells; B) Showing results for expression of beta defensin 1 in U937 cells (Jonas Steinmann, 2009).

On the other hand, phenylbutyrate does not induce defensin family rather than beta defensin-1. At the same dose, PBA only enhances beta defensin-1 but not beta defensin-2, 3 or 4 in VA10 cells. But it gave negative result in U937 cells. So, it varies from cell to cell type.

5.4 Isoleucine

Isoleucine is an essential, nonpolar, uncharged and branched chain aliphatic amino acid. It is the isomer of leucine and encoded like ATT-ATC-ATA (Center for Biological Sequence Analysis, University of Denmark). In the chemical structure, two groups and one side chain has been found which are α -amino group, α -carboxylic acid group and hydrocarbon side chain as consecutively. As it is essential amino acid, daily diet or supplements can fill up the gap of isoleucine in human body. The diet source for this element are meat, fish, cheese, eggs, lamb, soy protein and soy seeds (List is in order of highest to lowest of per 200 Calorie serving of the food, not volume or weight).

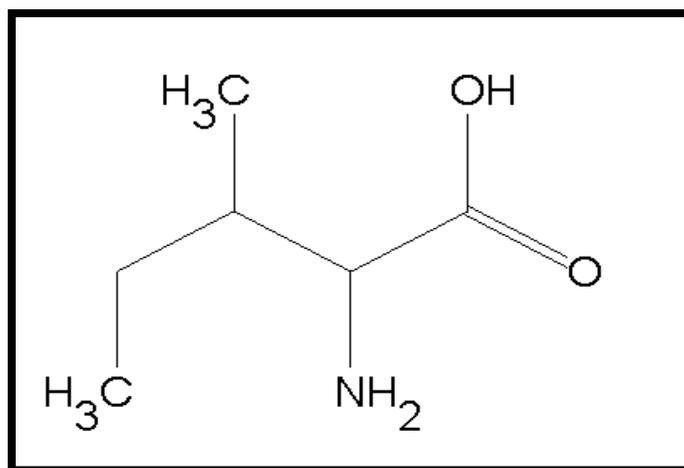


Figure 5.6: Chemical structure of isoleucine.

Isoleucine has various physiological functions including protein synthesis, induction of muscle protein synthesis, increase glucose uptake, detoxification, stimulate immune function and wound healing etc. It can be synthesized from plants and microorganisms following some steps which starts with pyruvic acid and alpha-ketoglutarate (Kisumi, Komatsubara, & Chibata, 1977).

In the middle of two enantiomers (L & D) of isoleucine, L-isoleucine is the potent inducer of epithelial beta defensin expression. The amount of isoleucine needed to reach at the peak level is between 3.12 - 12.5 $\mu\text{g/ml}$ in MDBK (Madin–Darby bovine kidney) cells (Pascale Fehlbaum, 2000). On the other hand, D-isoleucine require more concentration which is approximately 200 $\mu\text{g/ml}$. Moreover, the lower defensin activity has been observed with high concentration of this enantiomer. The micromolar quantity of L-isoleucine can serve as a marker for detecting microbes

and activate the antimicrobial action. It works through binding with receptor or enzyme intracellularly but the nature of receptor was unknown (Pascale Fehlbaum, 2000). It is also revealed that the inactivation of NF- κ B by pharmacologic inhibitor causes down regulation of isoleucine mediated defensin production. So, isoleucine can be used as an immunostimulant for this particular AMP production.

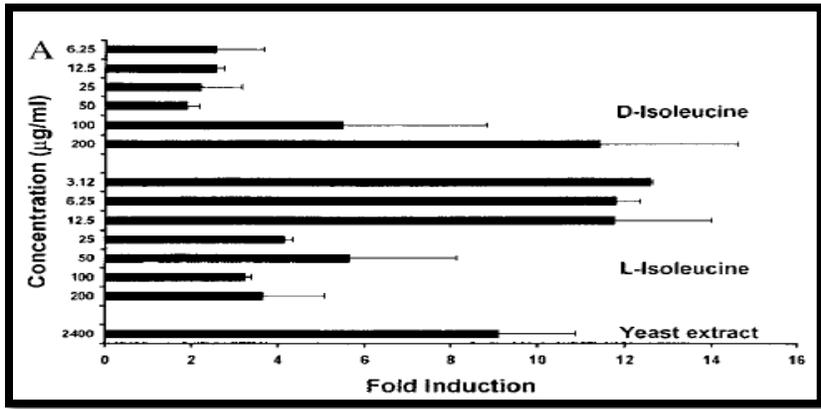


Figure 5.7: Relative expression of epithelial beta defensin in treatment with both L & D- isoleucine.

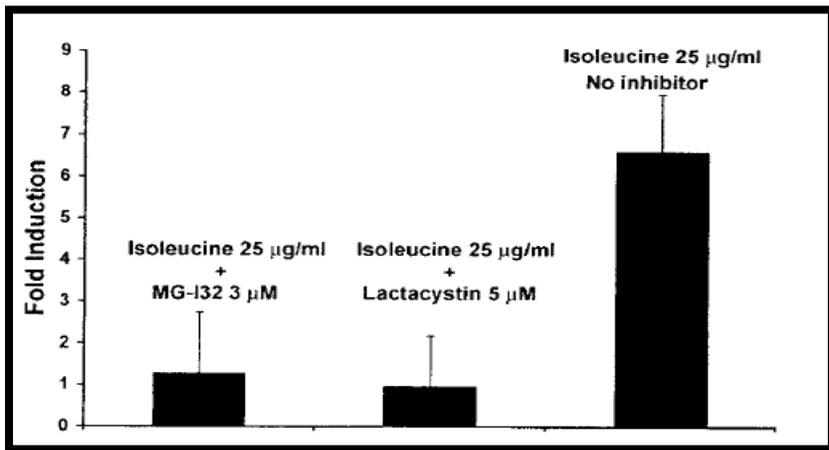


Figure 5.8: Proteasome inhibitors MG132 and Lactacystin block isoleucine activation of the defensin promoter.

Being inspired from the positive result, L- isoleucine was used in a murine model with progressive pulmonary tuberculosis to see the level of beta defensins expression (C. E. Rivas-Santiago, 2011). Isoleucine was injected into the mice after 8 weeks of infection. Significantly, both beta defensin 3 & 4 were induced with less tissue destruction even in mice infected with multidrug resistance bacilli. Though it could not fully cure the disease but might be used as a control therapy against this infectious one.

5.5 Sulforaphane

Sulforaphane (SFN) is a phytochemical agent containing isothiocyanate group in the chemical structure. The main source of sulforaphane is cruciferous vegetable such as, broccoli, Brussels sprout, cauliflower and cabbage etc. It is also known as broccoli extract because of their highly abundance in broccoli sprouts in the form of sulforaphane glucosinolate. It appears to be well absorbed from the intestine after oral administration and bioavailability is about 74% (Petri N, 2003). Sulforaphane works as an anticancer agent by epigenetic changes through inhibiting histone deacetylase (HDAC) activity. In colorectal cancer, it restricts cell proliferation and stimulates apoptosis and also does in prostate cancer (Myzak MC, 2006). This histone deacetylase inhibitor sulforaphane is responsible for inducing human beta defensin 2 in intestine epithelial cell.

In order to explore the above fact, an experiment was conducted which was based on colorectal cancer (Markus Schwab, 2008). In the experiment, human colorectal cancer cell lines Caco-2, HT-29 and SW480 were used and processed to make ready for doing various tests. From the result, it had been known that SFN can directly induce HBD-2 mRNA and protein in colonocyte. In HT-29 cells, HBD-2 mRNA was upregulated after 24 and 48hrs of treatment in a dose-dependent manner significantly in $\geq 5 \mu\text{M}$ concentration.

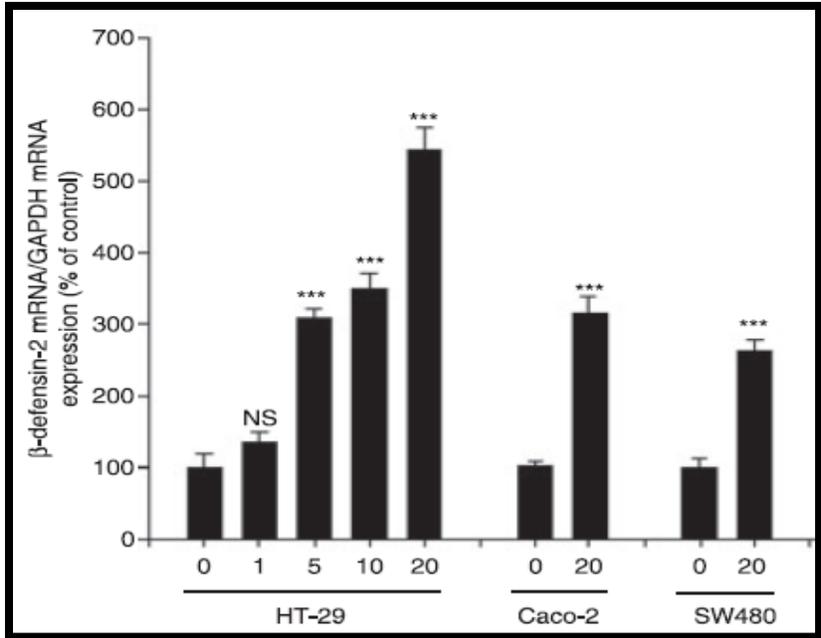


Figure 5.9: Dose dependent expression of sulphoraphane (1-20 μM) beta-defensin 2 in colorectal cell lines.

Also, HBD-2 gene was expressed upward in Caco-2 and SW480 after 24hrs. Following that, protein level was measured by ELISA where increasing HBD-2 level has been show. SFN influences the production of AMPs via vitamin D receptor (VDR) protein/ extra cellular-regulated kinase 1/2 and nuclear factor-κB signaling pathway. Upon the blocking of VDR, SFN mediated HBD-2 expression was down regulated. But in case of PPARγ and p38 MAPK, it remains unchanged. At the opposite side, NF-κB pathway is recognized as important factor for HBD-2 in colon epithelial cells (Wehkamp J, 2004). So, SFN can be useful in the therapy of colonic Crohn's disease.

5.6 Bile Salt

Bile salts are the derivatives of bile acids which contains sodium and potassium salts. Bile acids are steroid in nature which can be found in the bile of animals. There is basically three types of bile acids: primary, conjugated and secondary bile acids. Among them chenodeoxycholic acid is a primary bile acid which is synthesized in the liver. It has various function such as dissolve gallstones, treatment of cerebrotendineous xanthomatosis and constipation etc (Thistle JL, 1973) (Berginer VM, 1984).

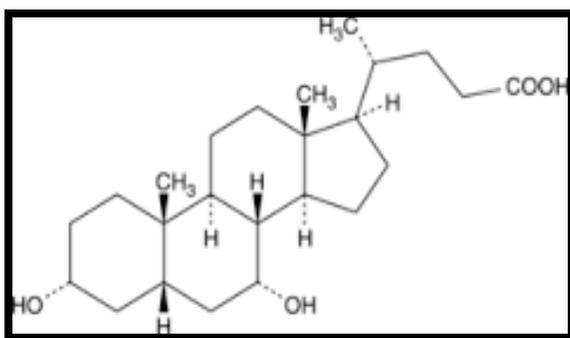


Figure 5.10: Chemical structure of chenodeoxycholic acid.

Ursodeoxycholic acid is a secondary bile acid which is metabolized from primary bile acids by intestinal bacteria. It has some significant medical uses for example reduction in gallstone formation, treatment of primary biliary cholangitis and improve bile flow in cystic fibrosis etc.

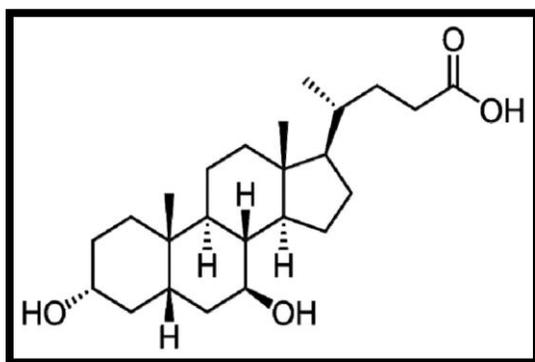


Figure 5.11: Chemical structure of ursodeoxycholic acid.

Bile salts may contribute to the cathelicidin expression in human biliary epithelium. In case of pathogen attacks, cathelicidin always expresses in the lumen of the biliary tract to defeat them. Again, we know that cathelicidin has affinity for VDR in biliary epithelial cells. Study showed

that both CDCA and UDCA induce the expression of AMP through two different nuclear receptors and they are farnesoid X receptor and vitamin D receptor (EMILIE D'ALDEBERT, 2009). Farnesoid X receptor is activated by CDCA and consequently act upon cathelicidin expression. Again, UDCA increases VDR protein expression in biliary epithelial cells resulting in AMP production. So, UDCA is dependent on VDR but CDCA is not.

It also has been found that combination therapy of CDCA, UDCA and VD3 can give better result in inflammatory biliary diseases, like primary biliary cirrhosis (Poupon RE, 1991) (Corpechot C, 2005).

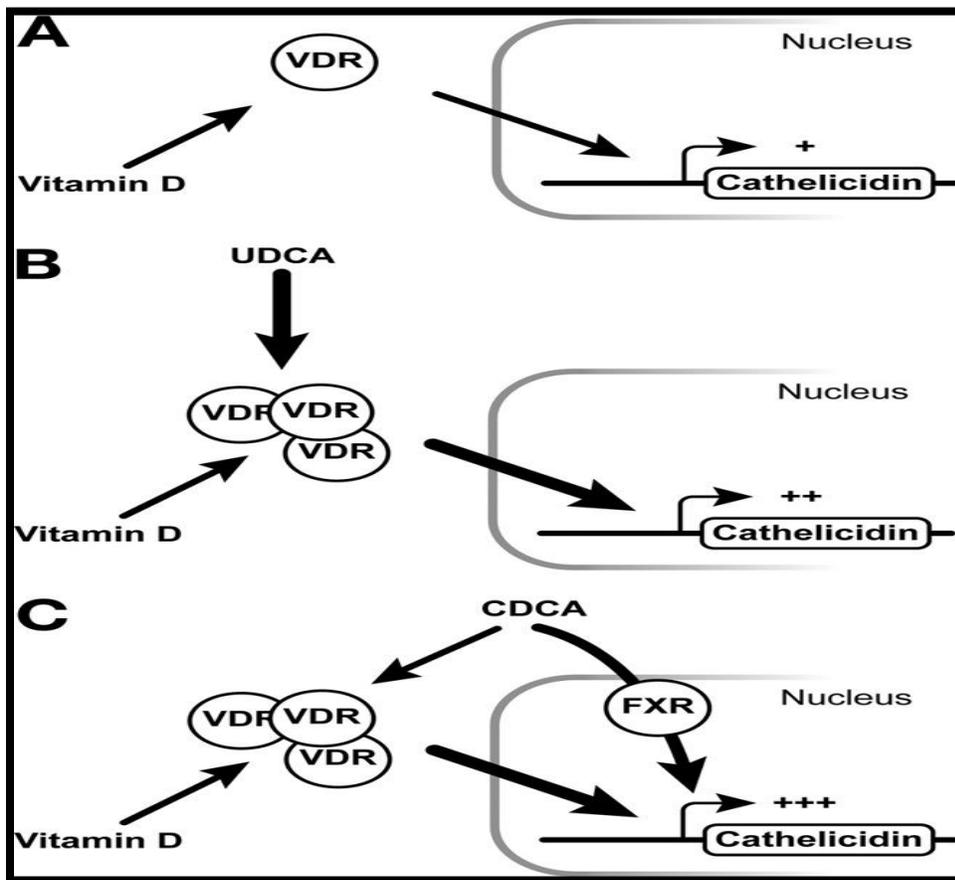


Figure 5.12: Showing possible effect of CDCA and UDCA along with vitamin D on cathelicidin's activity (EMILIE D'ALDEBERT, 2009).

5.7 Platelet Release Growth Factor

Platelet release growth factor is a constituent of whole blood serum which is purified from human platelets (Heldin C.H, 1999). It works as a mitogen for fibroblast, smooth muscle cell and glial cells. It helps in cell growth, division and angiogenesis. Studies shown that recombinant PRGF used to treat bone loss and chronic ulcer (GE, 2013).

The induction of HBD-2 occurs with the introduction of PRGF and in primary keratinocyte but in dose and time dependent manner (Andreas Bayer, 2016). When PRGF was treated with sample in vitro, HBD-2 gene expression was shown to increase after 24, 48 and 72 hours of PRGF stimulation. On the other hand, Vivostat PRF® also show positive result towards proliferation of HBD-2 in keratinocytes in the treatment of cutaneous wounds.

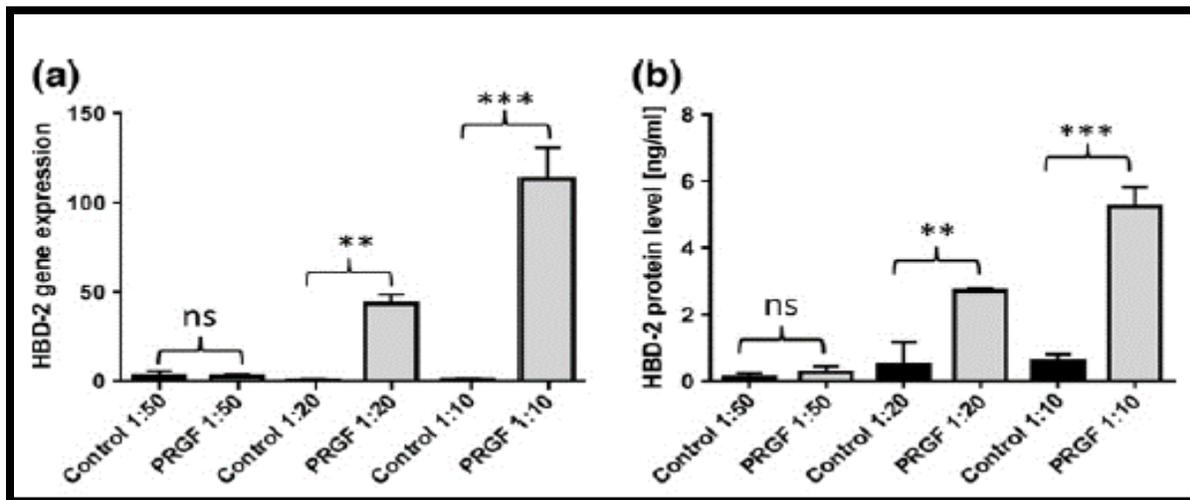


Figure 5.13: a) Showing HBD-2 gene expression with different concentration of PRGF; b) Showing protein level of HBD-2 with same concentration of PRGF as before (Andreas Bayer, 2016).

On the other hand, Vivostat PRF® also show positive result towards proliferation of HBD-2 in keratinocytes in the treatment of cutaneous wounds. In control part, wounds were treated with NaCl 0.9% and another part was treated with Vivostat PRF®. After ten days they were examined to see the HBD-2 level and the result was so much satisfactory.

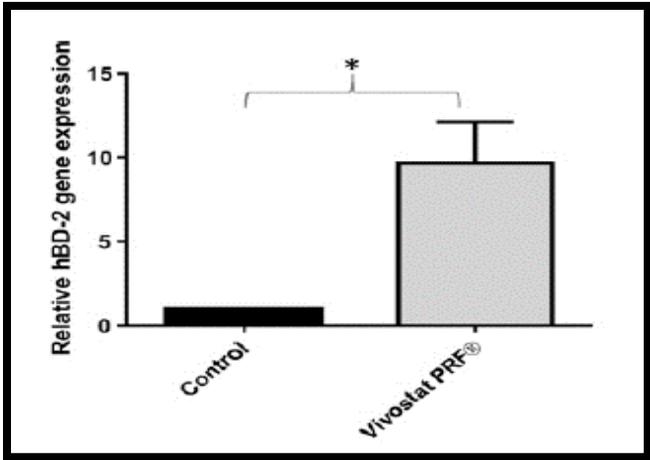


Figure 5.14: Showing the level of HBD-2 gene expression after treatment with Vivostat PRF® (Andreas Bayer, 2016).

In addition, epidermal growth factor receptor (EGFR) is potentially involved in this induction mechanism. This fact had been confirmed when an antibody (Cetuximab) is used to inactivate EGFR and HBD-2 level was drastically fallen down as a consequence. Moreover, induction of interleukin 6 (IL-6) was found in PRGF mediated experiment after 4hours stimulation. IL-6 is able to further increase activated protein 1 activity (Chuang J-Y, 2014).

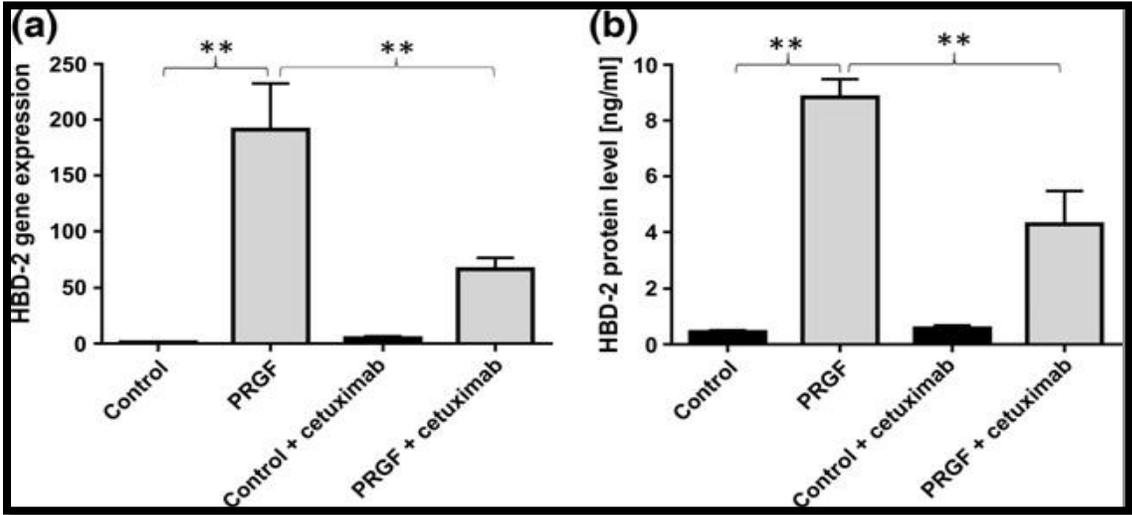


Figure 5.15: a) Showing HBD-2 gene expression upon activation of EGFR with cetuximab; b) Showing protein level of HBD-2 over that condition (Andreas Bayer, 2016).

6. Discussion:

In this study, some important factors are discussed in regulating antimicrobial peptide's production in human body. All of them work on cathelicidin and/or beta defensins. Epigallocatechin-3-gallate, isoleucine, sulphoraphane and platelet release growth factor act upon beta defensins family. On the other hand, vitamin D3 and bile salt act upon cathelicidin (LL-37). Meanwhile, phenylbutyrate effects on both cathelicidin and beta defensin 1.

Epigallocatechin-3-gallate which is a polyphenol present in green tea improve the immunological activity of intestinal epithelial cell. Approximately, 25 μ M is needed to get significant expression in both beta defensins. This effect can be useful to treat Crohn's diseases and inflammatory bowel diseases (Wehkamp, 2005). L-isoleucine can increase the level of beta defensins within a specific limit. This isoleucine can work as microbial marker to increase defensin activity and they act through a receptor which is still unknown. Sulphoraphane can directly induce gene expression of beta defensin 2. It also has dose dependent activity which is more than 5 μ M concentration. Platelet release growth factor also increase beta defensin 2 in keratinocytes in vitro. Again, Vivostat PRF® helps to regulate hBD-2 and treat chronic wounds.

Vitamin D3 is a factor which act upon cathelicidin expression. Though we know vitamin D3 binds with vitamin D receptor to become activate but in case of this regulation it must be bind with retinoid X receptor α . Moreover, the CAMP expression will increase where RXR activity is highest which is in skin. So, this vitamin D3 mediated cathelicidin expression can be used to treat atopic dermatitis. Moreover, supplementation of vitamin D injection can increase LL-37 level in ulcerative colitis patients (Sharifi A, 2016). Bile salts also induce cathelicidin expression in human biliary epithelium through farnesoid X receptor and vitamin D receptor. But a combination therapy of bile salts and vitamin D3 can give satisfactory result in primary biliary cirrhosis.

Phenylbutyrate is such a factor which can increase both cathelicidin and beta defensin 1. It induces cathelicidin expression in bronchial epithelial cell line. It works in dose and time dependent manner. Various synthetic analogs is used in medical field to avoid degradative nature of original one. Moreover, tuberculosis patient might be helpful by taking vitamin D3 and phenylbutyrate combination as a medicine because it reinforces the activity of phenylbutyrate. On the contrary, beta defensin 1 is induced by phenylbutyrate which again depend on cell to cell type. But no activity had been observed in beta defensin 2 expression with this factor.

Though all the factors are identified as the effective one but still some lackings are lying there. Many of them are not gone for human study or other animal study rather than one. So, the opportunity to get new medicines in relative diseases is dying. Phenylbutyrate is not stable in normal condition which is a big fault for introducing them as a drug or supplementary. Few of them need more intensive study to understand their proper mechanism of action. If all the drawbacks are taken into account and solved them in most possible way then these factors become more effective.

In the conclusion, it can be said that antimicrobial peptide has become a need for patient suffering of resistance issue from various antibiotics. It is the natural killer which can diminish microbial colonization in the body and keep free from being infected without developing resistance. Proper handling of these factors can make them useful for our benefits.

7. References:

- (R.J, 1939) Studies on a bactericidal agent extracted from a soil bacillus: I. Preparation of the agent. Its activity in vitro. *J. Exp. Med.* 1939; 70:1–10. doi: 10.1084/jem.70.1.1.43,300–304. 70, 11–17.
- Akhirunnesa Mily, Rokeya Sultana Rekha, S M Mostafa Kamal, Evana Akhtar, Protim Sarker, Zeaur Rahim, Gudmundur H Gudmundsson, Birgitta Agerberth and Rubhana Raqib. Oral intake of phenylbutyrate with or without vitamin D₃ upregulates the cathelicidin LL-37 in human macrophages: a dose finding study for treatment of tuberculosis. Mily et al. *BMC Pulmonary Medicine* 2013, 13:23.
- Andreas Bayer, Justus Lammel, Franziska Rademacher, Justus Groß, Markus Siggelkow, Sebastian Lippross, Tim Kluter, Deike Varoga, Mersedeh Tohidnezhad, Thomas Pufe, Jochen Cremer, Regine Glaser, and Jurgen Harder. Platelet-released growth factors induce the antimicrobial peptide human beta-defensin-2 in primary keratinocytes. *Experimental Dermatology*, 2016, 25, 460–465.
- Ayabe T, Satchell DP, Wilson CL, Parks WC, Selsted ME, Ouellette AJ. Secretion of microbicidal α -defensins by intestinal Paneth cells in response to bacteria. *Nature Immunology*. 2000;1(2):113–118.
- Bals R, Weiner DJ, Meegalla RL et al (1999) Transfer of a cathelicidin peptide antibiotic gene restores bacterial killing in a cystic fibrosis xenograft model. *J Clin Invest* 103:1113–1117.
- Bell, S. J., Goodrick, G. K., A functional food product for the management of weight. *Crit. Rev. Food. Sci. Nutr.* 2002, 42, 163–178.
- Berginer VM, Salen G, Shefer S (December 1984). "Long-term treatment of cerebrotendinous xanthomatosis with chenodeoxycholic acid". *N. Engl. J. Med.* 311 (26): 1649–52.
- Bergman P, Walter-Jallow L, Broliden K et al (2007) The antimicrobial peptide LL-37 inhibits HIV-1 replication. *Curr HIV Res* 5:410–415.
- Bodner L, Dayan D, Rothchild D et al. Extraction wound healing in desalivated rats. *Journal of oral pathology & medicine : official publication of the International Association of Oral Pathologists and the American Academy of Oral Pathology* 1991; 20: 176-8.
- Boguniewicz M, Leung DY. Atopic dermatitis. *J Allergy Clin Immunol* 2006;117:S475–S480.

- Boukamp P, Petrussevska RT, Breitkreutz D, Hornung J, Markham A, Fusenig NE (1988) Normal keratinization in a spontaneously immortalized aneuploid human keratinocyte cell line. *J Cell Biol* 106: 761–771
- Brogden KA. Antimicrobial peptides: pore formers or metabolic inhibitors in bacteria? *Nature Reviews Microbiology*. 2005;3(3):238-250.
- Buphenyl. Drugs@FDA. United States Food and Drugs Administration. Retrieved 26 October 2013.
- C. E. Rivas-Santiago, B. Rivas-Santiago, D. A. León, J. Castañeda-Delgado and R. Hernández Pando. Induction of b-defensins by l-isoleucine as novel immunotherapy in experimental murine tuberculosis. *Clinical and Experimental Immunology* © 2011 British Society for Immunology, *Clinical and Experimental Immunology*, 164: 80–89.
- Cabrera C, Artacho R, Giménez R: Beneficial effects of green tea: a review. *J Am Coll Nutr*. 2006, 25: 79-99
- Carlberg C, Bendik I, Wyss A, Meier E, Sturzenbecker LJ, Grippo JF, Hunziker W (1993) Two nuclear signaling pathways for vitamin D. *Nature* 361:657–660
- Carlberg C, Campbell MJ (2013) Vitamin D receptor signaling mechanisms: integrated actions of a well-defined transcription factor. *Steroids* 78:127–136
- Carretero M, Escamez MJ, Garcia M, Duarte B, Holguín A, Retamosa L, et al. In vitro and in vivo wound healing-promoting activities of human cathelicidin LL-37. *J Invest Dermatol*. 2008;128:223–36
- Center for Biological Sequence Analysis, University of Denmark.
- Chaly YV, Paleolog EM, Kolesnikova TS, et al.: Neutrophil alpha-defensin human neutrophil peptide modulates cytokine production in human monocytes and adhesion molecule expression in endothelial cells. *Eur Cytokine Netw* 2000, 11:257–266.
- Chuang J-Y, Huang Y-L, Yen W-L et al. *Int J Mol Sci* 2014: 15: 545–559.
- Cociancich S, Ghazi A, Hetru A, Hoffman J A, Letellier L. Insect defensin, an inducible antibacterial peptide, forms voltage-dependent channels in *Micrococcus luteus*. *J Biol Chem*. 1993;260: 19239–19245.
- Conner K, Nern K, Rudisill J et al (2002) The antimicrobial peptide LL-37 is expressed by keratinocytes in condyloma acuminatum and verruca vulgaris. *J Am Acad Dermatol* 47:347–350.

- Corpechot C, Carrat F, Bahr A, et al. The effect of ursodeoxycholic acid therapy on the natural course of primary biliary cirrhosis. *Gastroenterology* 2005;128:297–303.
- Cunliffe RN, Rose FR, Keyte J, Abberley L, Chan WC, Mahida YR. Human defensin 5 is stored in precursor form in normal Paneth cells and is expressed by some villous epithelial cells and by metaplastic Paneth cells in the colon in inflammatory bowel disease. *Gut* 2001;48: 176-85.
- Dafar A, Rico P, Isik A et al. Quantitative detection of epidermal growth factor and interleukin-8 in whole saliva of healthy individuals. *J Immunol Methods* 2014; 408: 46-51.
- Daniel Svensson; Daniel Nebel; Ulrikke Voss; Eva Ekblad & Bengt-Olof Nilsson. Vitamin D-induced up-regulation of human keratinocyte cathelicidin anti-microbial peptide expression involves retinoid X receptor α . 2016.
- De Y, Chen Q, Schmidt AP, Anderson GM, Wang JM, Wooters J, et al. LL-37, the neutrophil granule and epithelial cell-derived cathelicidin, utilizes formyl peptide receptor-like 1 (FPRL1) as a receptor to chemoattract human peripheral blood neutrophils, monocytes, and T cells. *J Exp Med*. 2000;192: 1069–74.
- De Y, Chen Q, Schmidt AP, Anderson GM, Wang JM, Wooters J, Oppenheim JJ, Chertov O. LL-37, the neutrophil granule- and epithelial cell-derived cathelicidin, utilizes formyl peptide receptor-like 1 (FPRL1) as a receptor to chemoattract human peripheral blood neutrophils, monocytes, and T cells. *J Exp Med*. 2000;192:1069-1074.
- den Hertog AL, van Marle J, Veerman EC et al (2006) The human cathelicidin peptide LL-37 and truncated variants induce segregation of lipids and proteins in the plasma membrane of *Candida albicans*. *Biol Chem* 387:1495–1502.
- Dorland's Illustrated Medical Dictionary, under Vitamin (Table of Vitamins).
- Dubos, R.J. Studies on a bactericidal agent extracted from a soil bacillus: II. Protective effect of the bactericidal agent against experimental *Pneumococcus* infections in mice. *J. Exp. Med.* 1939,
- Elahi, S., Buchanan, R. M., Attah-Poku, S., Townsend, H. G. G. et al., The host defense peptide beta-defensin 1 confers protection against *Bordetella pertussis* in newborn piglets. *Infect. Immun.* 2006, 74, 2338–2352.
- EMILIE D'ALDEBERT, MARIE-JEANNE BIYEYEME BI MVE, MARTINE MERGEY, DOMINIQUE WENDUM, DELPHINE FIRRINCIELI, AUDREY COILLY, LAURA

- FOUASSIER, CHRISTOPHE CORPECHOT, RAOUL POUPON, CHANTAL HOUSSET and NICOLAS CHIGNARD. Bile Salts Control the Antimicrobial Peptide Cathelicidin Through Nuclear Receptors in the Human Biliary Epithelium. *GASTROENTEROLOGY* 2009;136:1435–1443.
- Enoch S, Stephens P. Scarless healing: oral mucosa as a scientific model. *Wounds UK* 2009; 5: 42-8.
- Essentials of Human Physiology.
- Friedlaender GE et al. The Role of Recombinant Human Platelet-derived Growth Factor-BB (rhPDGF-BB) in Orthopaedic Bone Repair and Regeneration *Curr Pharm Des.* 2013;19(19):3384-90.
- FROHM M, AGERBERTH B, AHANGARI G et al.: The expression of the gene coding for the antibacterial peptide LL-37 is induced in human keratinocytes during inflammatory disorders. *J. Biol. Chem.*(1997) 272:15258-15263
- Frohman M, Agerberth B, Ahangari G, et al. The expression of the gene coding for the antibacterial peptide LL-37 is induced in human keratinocytes during inflammatory disorders. *J Biol Chem* 1997;272:15258-15263
- Gallo RL, Ono M, Povsic T, et al. Syndecans, cell surface heparan sulfate proteoglycans, are induced by a proline-rich antimicrobial peptide from wounds. *Proc Natl Acad Sci U S A* 1994;91:11035-11039
- Ganz T, Selsted M E, Lehrer R I. Defensins. *Eur J Haematol.* 1990;44: 1–8.
- Ganz, T. The role of antimicrobial peptides in innate immunity. *Integr. Comp. Biol.* 2003,
- Geens, M. M., Niewold, T. A., Optimizing culture conditions of a porcine epithelial cell line IPEC-J2 through a histological and physiological characterization. *Cytotechnology* 2011, 63, 415–423.
- Gennaro R, Zanetti M (2000). "Structural features and biological activities of the cathelicidin-derived antimicrobial peptides". *Biopolymers.* 55 (1): 31–49
- Gera J F, Lichenstein A. Human neutrophil peptide defensins induce single strand (Gudmundsson GH, 1995)d DNA breaks in target cells. *Cell Immunol.* 1991;138:108–120
- Gilbert, J., S. D. Baker, M. K. Bowling, L. Grochow, W. D. Figg, Y. Zabelina, R. C. Donehower, and M. A. Carducci. 2001. A phase I dose escalation and bioavailability

- study of oral sodium phenylbutyrate in patients with refractor solid tumor malignancies. *Clin. Cancer Res.* 7:2292–2300.
- Gould IM, Bal AM. New antibiotic agents in the pipeline and how they can overcome microbial resistance. *Virulence* 2013;4(2):185–19
- Gudmundsson GH, Magnusson KP, Chowdhary BP, Johansson M, Andersson L, Boman HG (1995) Structure of the gene for porcine peptide antibiotic PR-39, a cathelin gene family member: comparative mapping of the locus for the human peptide antibiotic FALL-39. *Proc Natl Acad Sci USA* 92:7085–7089
- Hancock RE, Diamond G. The role of cationic antimicrobial peptides in innate host defences. *Trends in Microbiology.* 2000;8(9):402-410.
- Hancock RE, Lehrer R. Cationic peptides: a new source of antibiotics. *Trends in Biotechnology.* 1998; 16(2):82-88.
- Hancock, R. E. W. & Sahl, H. G. Antimicrobial and host-defence peptides as new anti-infective therapeutic strategies. *Nat. Biotechnol.* 24, 1551–1557 (2006)
- Harris M, Mora-Montes HM, Gow NA, Coote PJ (2009) Loss of mannosylphosphate from *Candida albicans* cell wall proteins results in enhanced resistance to the inhibitory effect of a cationic antimicrobial peptide via reduced peptide binding to the cell surface. *Microbiology* 155: 1058–1070.
- Haussler MR, KerrWhitfield G, Kaneko I, Haussler CA, Hsieh D, Hsieh JC, Jurutka PW (2013) Molecular mechanisms of vitamin D action. *Calcif Tissue Int* 92:77–98
- Håvard Jenssen, Pamela Hamill, and Robert E. W. Hancock. Peptide Antimicrobial Agents. *CLINICAL MICROBIOLOGY REVIEWS*, July 2006, p. 491–511.
- Heldin C.H, Westermark B. Mechanism of Action and In Vivo Role of Platelet-Derived Growth Factor. *Physiological Reviews* Published 10 January 1999 Vol. 79 no. 4, 1283-1316.
- Iannitti, Tommaso; Beniamino Palmieri (September 2011). Clinical and Experimental Applications of Sodium Phenylbutyrate. *Drugs in R & D. Adis Data Information BV.* 11 (3): 227–249. *Immunol* 2008;122:261–266.
- Jonas Steinmann, Skarpheðinn Halldórsson, Birgitta Agerberth, and Gudmundur H. Gudmundsson. Phenylbutyrate Induces Antimicrobial Peptide Expression. Institute of

- Biology, University of Iceland, Reykjavik, Iceland, and Medical Biophysics and Biochemistry, Karolinska Institutet, Stockholm, Sweden. 2009. Vol. 53, No. 12.
- Jones DE, Bevins CL (1992) Paneth cells of the human small intestine express an antimicrobial peptide gene. *J. Biol. Chem.* 267: 23216-23225.
- Kemik O, Kemik AS, Sumer A, Begenic H, Purisa S, Tuzun S. Human neutrophil peptides 1, 2 and 3 (HNP 1-3): elevated serum levels in colorectal cancer and novel marker of lymphatic and hepatic metastasis. *Hum Exp Toxicol* 2011;32:167-71
- Kisumi, M; Komatsubara, S; Chibata, I (Jul 1977). Pathway for isoleucine formation from pyruvate by leucine biosynthetic enzymes in leucine accumulating isoleucine revertants of *Serratia marcescens*. *J. Biochem.* 82: 95–103.
- KOCZULLA R, VON DEGENFELD G, KUPATT C et al. : An angiogenic role for the human peptide antibiotic LL-37/hCAP-18. *J. Clin. Invest.* (2003) 111:1665-1672.
- Kris De Smet & Roland Contreras (2005) Human antimicrobial peptides: defensins, cathelicidins and histatins. *J. Springer. Biotechnology Letters* 27: 1337-1347.
- Lee CC, Sun Y, Qian S, Huang HW. Transmembrane pores formed by human antimicrobial peptide LL-37. *Biophys J* 2011; 100: 1688-96.
- Lee MJ, Maliakal P, Chen L, Meng X, Bondoc FY, Prabhu S et al. (2002). Pharmacokinetics of tea catechins after ingestion of green tea and (-)-epigallocatechin-3-gallate by humans: formation of different metabolites and individual variability. *Cancer Epidemiol Biomarkers Prev* 11: 1025–103
- Lehrer R I, Barton A, Daher K A, Harwig S S L, Ganz T, Selsted M E. Interaction of human defensins with *Escherichia coli*. Mechanism of activity. *J Clin Invest.* 1989;84:553–561
- Lehrer R I, Szklarek D, Ganz T, Selsted M E. Correlation of binding of rabbit granulocyte peptides to *Candida albicans* with candidacidal activity. *Infect Immun.* 1985;49:207–211
- Liu PT, Stenger S, Li H et al (2006) Toll-like receptor triggering of a vitamin D-mediated human antimicrobial response. *Science* 311:1770–1773.
- Liu, P. T., S. Stenger, D. H. Tang, and R. L. Modlin. 2007. Cutting edge: vitamin D-mediated human antimicrobial activity against *Mycobacterium tuberculosis* is dependent on the induction of cathelicidin. *J. Immunol.* 179:2060–2063.

- Liu, P. T., S. Stenger, H. Li, L. Wenzel, B. H. Tan, S. R. Krutzik, M. T. Ochoa, J. Schaubert, K. Wu, C. Meinken, D. L. Kamen, M. Wagner, R. Bals, A. Steinmeyer, U. Zügel, R. L. Gallo, D. Eisenberg, M. Hewison, B. W. Hollis, J. S. Adams, B. R. Bloom, and R. L. Modlin. 2006. Toll-like receptor triggering of a vitamin D-mediated human antimicrobial response. *Science* 311: 1770–1773.
- Lombardo Bedran, T. B., Feghali, K., Zhao, L., Palomari Spolidorio, D. M. et al., Green tea extract and its major constituent, epigallocatechin-3-gallate, induce epithelial beta-defensin secretion and prevent beta-defensin degradation by *Porphyromonas gingivalis*. *J. Periodontal Res.* 2014, 49, 615–623.
- Lopez-Garcia B, Lee PH, Yamasaki K et al (2005) Anti-fungal activity of cathelicidins and their potential role in *Candida albicans* skin infection. *J Invest Dermatol* 125:108–115.
- Mariani, V., Palermo, S., Fiorentini, S., Lanubile, A., Giuffra, E., Gene expression study of two widely used pig intestinal epithelial cell lines: IPEC-J2 and IPI-2I. *Vet. Immunol. Immunopathol.* 2009, 131, 278–284.
- Martinez, J. L., & Olivares, J. (2012). Environmental Pollution By Antibiotic Resistance Genes. In P. L. Keen, & M. H. Montforts, *Antimicrobial Resistance in the Environment* (pp. 151- 171). Hoboken, N.J.: John Wiley & Sons.
- Maton, Anthea (1993). *Human Biology and Health*. Prentice Hall
- Matsuzaki K, Yoneyama S, Fujii N, Miyajima K, Yamada K, Kirino Y, Anzai K. Membrane permeabilization mechanisms of a cyclic antimicrobial peptide, tachyplesin I, and its linear analog. *Biochemistry.* 1997; 36(32):9799-9806.
- McKay, D. L., Blumberg, J. B., The role of tea in human health: an update. *J. Am. Coll. Nutr.* 2002, 21, 1–13.
- Mendez-Samperio P, Miranda E, Trejo A (2008) Expression and secretion of cathelicidin LL-37 in human epithelial cells after infection by *Mycobacterium bovis* Bacillus Calmette-Guerin. *Clin Vaccine Immunol* 15:1450–1455.
- Murphy L. Y. Wan, K. H. Ling, M. F. Wang and Hani El-Nezami., Green tea polyphenol epigallocatechin-3-gallate improves epithelial barrier function by inducing the production of antimicrobial peptide pBD-1 and pBD-2 in monolayers of porcine intestinal epithelial IPEC-J2 cells. *Mol. Nutr. Food Res.* 2016, 60, 1048–1058.

- Myzak MC, Ho E, Dashwood RH. Dietary agents as histone deacetylase inhibitors. *Mol Carcinog* 2006; 45:443–6.
- Nijnik A, Hancock RE (2009) The roles of cathelicidin LL-37 in immune defences and novel clinical applications. *Curr Opin Hematol* 16:41–47.
- Niyonsaba, F.; Iwabuchi, K.; Matsuda, H.; Ogawa, H.; Nagaoka, I. Epithelial cell-derived human beta-defensin-2 acts as a chemotaxin for mast cells through a pertussis toxin-sensitive and phospholipase c-dependent pathway. *Int. Immunol.* 2002, 14, 421–426.
- O’Neill J. Tackling drug-resistant infections globally: Final report and recommendations. The review on antimicrobial resistance; London: HM Government and the Wellcome Trust; 2016.
- Ong PY, Ohtake T, Brandt C, Strickland I, Boguniewicz M, Ganz T, et al. Endogenous antimicrobial peptides and skin infections in atopic dermatitis. *N Engl J Med* 2002;347:1151–1160.
- Overhage J, Campisano A, Bains M et al (2008) Human host defense peptide LL-37 prevents bacterial biofilm formation. *Infect Immun* 76:4176–4182.
- Pace, B. S., G. L. White, G. J. Dover, M. S. Boosalis, D. V. Faller, and S. P. Perrine. 2002. Short-chain fatty acid derivatives induce fetal globin expression and erythropoiesis in vivo. *Blood* 100:4640–4648.
- Pascale Fehlbaum, Meena Rao, Michael Zasloff, and G. Mark Anderson. An essential amino acid induces epithelial beta-defensin expression. Magainin Research Institute and Magainin Pharmaceuticals Inc. 2000.
- Petri N, et al Absorption/metabolism of sulforaphane and quercetin, and regulation of phase II enzymes, in human jejunum in vivo . *Drug Metab Dispos.* (2003)
- Peyrin-Biroulet L, Beisner J, Wang G, Nuding S, Oommen ST, Kelly D, Parmentier-Decrucq E, Dessein R, Merour E, Chavatte P, Grandjean T, Bressenot A, Desreumaux P, Colombel JF, Desvergne B, Stange EF, Wehkamp J, Chamailard M. Peroxisome proliferator-activated receptor gamma activation is required for maintenance of innate antimicrobial immunity in the colon. *Proc Natl Acad Sci U S A* 2010;107:8772-7
- Poupon RE, Balkau B, Eschwege E, et al. A multicenter, controlled trial of ursodiol for the treatment of primary biliary cirrhosis. UDCA-PBC Study Group. *N Engl J Med* 1991;324:1548–1554.

- Radek, K.; Gallo, R. Antimicrobial peptides: Natural effectors of the innate immune system. *Semin. Immunopathol.* 2007, 29, 27–43.
- Rao AG. Conformation and antimicrobial activity of linear derivatives of tachyplesin lacking disulfide bonds. *Archives of biochemistry and biophysics.* 1999; 361(1):127-134.
- Ren SX, Cheng AS, To KF, Tong JH, Li MS, Shen J, Wong CC, Zhang L, Chan RL, Wang XJ, Ng SS, Chiu LC, Marquez VE, Gallo RL, Chan FK, Yu J, Sung JJ, Wu WK, Cho CH. Host immune defense peptide LL-37 activates caspase-independent apoptosis and suppresses colon cancer. *Cancer Res* 2012;72: 6512-23.
- Richard L. Gallo, MD, PhD, and Victor Nizet, MD. *Endogenous Production of Antimicrobial Peptides in Innate Immunity and Human Disease.* 2003.
- Rivas-Santiago B, Hernandez-Pando R, Carranza C et al (2008) Expression of cathelicidin LL-37 during *Mycobacterium tuberculosis* infection in human alveolar macrophages, monocytes, neutrophils, and epithelial cells. *Infect Immun* 76:935–941.
- Roupe KM, Nybo M, Sjöbring U et al. Injury is a major inducer of epidermal innate immune responses during wound healing. *J Invest Dermatol* 2010; 130: 1167-77.
- Sano M, Tabata M, Suzuki M, Degawa M, Miyase T, Maeda-Yamamoto M: Simultaneous determination of twelve tea catechins by high-performance liquid chromatography with electrochemical detection. *Analyst.* 2001, 126: 816-820. 10.1039/b102541b.
- Scalbert, A., Manach, C., Morand, C., Remesy, C. et al., Dietary polyphenols and the prevention of diseases. *Crit. Rev. Food Sci. Nutr.* 2005, 45, 287–306.
- Schaller-Bals S, Schulze A, Bals R (2002) Increased levels of antimicrobial peptides in tracheal aspirates of newborn infants during infection. *Am J Respir Crit Care Med* 165:992–995.
- Schauber J, Dorschner R, Coda A, Büchau A, Liu P, Kiken D, et al. Injury enhances TLR2 function and antimicrobial peptide expression through a vitamin D-dependent mechanism. *J Clin Invest* 2007;117:803–811.
- Schauber J, Gallo RL. Antimicrobial peptides and the skin immune defense system. *J Allergy Clin*
- Schultz-Larsen FV, Hanifin JM. Epidemiology of atopic dermatitis. *Immunol Allergy Clin North Am* 2002;22:1–24.

- Schwab M, Reynders V, Loitsch S, Steinhilber D, Schroder O and Stein J. The dietary histone deacetylase inhibitor sulforaphane induces human β -defensin-2 in intestinal epithelial cells. 2008. *Immunology*, 125, 241–251.
- Sharifi A, Hosseinzadeh-Attar MJ, Vahedi H, Nedjat S. A randomized controlled trial on the effect of vitamin D3 on inflammation and cathelicidin gene expression in ulcerative colitis patients. *Saudi J Gastroenterol* 2016;22:316-23.
- Smet, K. D., Contreras, R., Human antimicrobial peptides: defensins, cathelicidins and histatins. *Biotechnol. Lett.* 2005, 27, 1337–1347.
- Sørensen OE, Cowland JB, Theilgaard-Mönch K et al. Wound healing and expression of antimicrobial peptides/polypeptides in keratinocytes, a consequence of common growth factors. *J Immunol.* 2003; 170: 5583-9.
- Sørensen OE, Thapa DR, Rosenthal A et al. Differential regulation of β -defensin expression in human skin by microbial stimuli. *J Immunol.* 2005; 174: 4870-9.
- Spira AI, Carducci MA. Differentiation therapy. *Curr Opin Pharmacol* 2003; 3: 338–43.
- Stolzenberg ED, Anderson GM, Ackermann MR, Whitlock RH, Zasloff M. Epithelial antibiotic induced in states of disease. *Proc Natl Acad Sci U S A* 1997;94:8686-8690
- Szpaderska AM, Zuckerman JD, DiPietro LA. Differential injury responses in oral mucosal and cutaneous wounds. *J Dent Res* 2003; 82: 621-6.
- T. Mohanty, P. Alberius, A. Schmidtchen, K. Reiss⁴, J.-M. Schröder, O.E. Sørensen. Saliva induces expression of antimicrobial peptides and promotes intracellular killing of bacteria in keratinocytes by EGFR-transactivation. 2016.
- The Lancet Infectious Diseases Commission. Antibiotic resistance—the need for global solutions. *Lancet Infect Dis* 2013; 13: 1057–98.
- Thistle JL, Hofmann AF (September 1973). "Efficacy and specificity of chenodeoxycholic acid therapy for dissolving gallstones". *N. Engl. J. Med.* 289 (13): 655–9.
- Tissa R. Hata, Paul Kotol, Michelle Jackson, Meggie Nguyen, Aimee Paik, Don Udall, Kimi Kanada, Kenshi Yamasaki, Phd, Doru Alexandrescu, and Richard L. Gallo. Administration of oral vitamin D induces cathelicidin production in atopic individuals. 2008.

- Veldhuizen, E. J. A., Rijnders, M., Claassen, E. A., van Dijk, A. et al, Porcine α -defensin 2 displays broad antimicrobial activity against pathogenic intestinal bacteria. *Mol. Immunol*, 2008, 45, 386–394.
- Veldhuizen, E., Hendriks, H., Hogenkamp, A., van Dijk, A. et al., Differential regulation of porcine [beta]-defensins 1 and 2 upon Salmonella infection in the intestinal epithelial cell line IPI-2I. *Vet. Immunol. Immunopathol.* 2006, 114, 94–102.
- Vieth R, Chan P, MacFarlane G. Efficacy and safety of vitamin D3 intake exceeding the lowest observed adverse effect level. *Am J Clin Nutr* 2001;73:288–294.
- Wade WG. The oral microbiome in health and disease. *Pharmacol Res.* 2013; 69: 137-43.
- Wehkamp J, Harder J, Wehkamp K et al. NF-kappaB- and AP-1-mediated induction of human beta defensin-2 in intestinal epithelial cells by Escherichia coli Nissle 1917: a novel effect of a probiotic bacterium. *Infect Immun* 2004; 72:5750–8.
- Wehkamp, J., Fellermann, K., Herrlinger, K. R., Bevins, C. L. et al., Mechanisms of disease: defensins in gastrointestinal diseases. *Nat. Clin. Pract. Gastroenterol. Hepatol.* 2005, 2, 406–415.
- White S H, Wimley W C, Selsted M E. Structure, function, and membrane integration of defensins. *Curr Opin Struct Biol.* 1995;5: 521–527.
- Yang D, Chen Q, Schmidt AP, Anderson GM, Wang JM, Wooters J, Oppenheim JJ, Chertov O. LL-37, the neutrophil granule- and epithelial cell-derived cathelicidin, utilizes formyl peptide receptor-like 1 (FPRL1) as a receptor to chemoattract human peripheral blood neutrophils, monocytes, and T cells. *Journal of Experimental Medicine.* 2000;192: 1069-1074.
- Yeaman MR, Yount NY (2003) Mechanisms of antimicrobial peptide action and resistance. *Pharmacol Rev* 55: 27–55.
- Zaveri, N. T., Green tea and its polyphenolic catechins: medicinal uses in cancer and noncancer applications. *Life Sci.* 2006, 78, 2073–2080.
- Zhang, J., Deng, J., Li, Y., Yang, Q., The effect of Lactobacillus on the expression of porcine α -defensin-2 in the digestive tract of piglets. *Livest. Sci.* 2011, 138, 259–265.

